

1 **Network complexity of rubber plantations is lower than tropical forests for soil**
2 **bacteria but not fungi**

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9 **Running title:** Forest conversion alters soil microbial networks

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17 **Abstract:**

18 Soil microbial communities play a crucial role in ecosystem functioning. Past
19 research has examined the effects of forest conversion on soil microbial composition
20 and diversity, but it remains unknown how networks within these communities
21 respond to forest conversion, including when tropical rainforests are replaced with
22 rubber plantations. Microbial networks are viewed as critical indicators of soil health
23 and quality. They consist of two parts: nodes and edges. In this study, we used data
24 from Illumina sequencing and metagenome shotgun sequencing to analyze bacterial
25 and fungal community network structure in a large number of soil samples from
26 tropical rainforests and rubber plantation sites in Hainan Island, China. Our results
27 showed only 5-10% shared network edges (i.e., links between species A and B
28 existing in both rubber plantations and rainforests) were observed in both bacterial
29 and fungal communities, which indicates that forest conversion altered soil microbial
30 network structure. The identity of keystone OTUs differed entirely between rubber
31 plantation and rainforest sites, further underscoring the altered network structure.
32 More edges and more negative correlations within the soil bacterial-fungal networks
33 were observed in rubber plantation sites (dry season: 4,284 total edges, 844 negative;
34 rainy season: 7,257 total, 1,744 negative) than in rainforest sites (dry season: 3,650
35 total, 149 negative; rainy season: 6,018 total, 489 negative), demonstrating that soil
36 bacterial-fungal network structure was more complex and stable in rubber plantations
37 than in rainforests. For bacteria, a larger number of network edges were observed
38 among bacterial networks in samples from tropical rainforest than in samples from
39 rubber plantations, indicating that rainforest bacterial networks were more complex
40 than those from rubber plantations. However, soil fungal networks from rubber
41 plantations showed more links, suggesting that forest conversion increased fungal
42 network complexity. More edges of network and more links between species and
43 functions were observed in the rainy season than in the dry season indicating that
44 seasonal changes had a strong effect on network structure and function. Further

45 analysis shows soil pH, potassium (AK), and total nitrogen (TN) had more links with
46 species of some phyla. In conclusion, forest conversion results in an increase in soil
47 pH, and a decrease in AK and TN, and these changes as well as seasonal variations
48 had a great impact on soil microbial composition, network structure and function.

49 **Copyright statements:** The copyright statement will be included by Copernicus, if
50 applicable.

51 **Keyword:** Tropical rainforest, Rubber plantations, Networks, Soil microbial
52 community, Forest conversion

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54 **1. Introduction**

55 ‘Soil microbiome’ refers to the enormous variety of microorganisms inhabiting the
56 complex soil environment. It is highly diverse and comprises up to one quarter of
57 Earth’s species (Wagg et al., 2019). Soil microbes play a critical role in the
58 maintenance of soil quality and function, and they represent more than half of the
59 biodiversity of terrestrial ecosystems (Philippot et al., 2013). A number of studies
60 have investigated how the conversion of tropical forests to rubber plantations impacts
61 soil microbial composition and diversity (Schneider et al., 2015; Kerfahi et al., 2016,
62 Lan et al., 2017a; 2017b; 2017c; Lan et al., 2020a; 2020b; 2021). Studies conducted
63 in Indonesia (Schneider et al. 2015), Malaysia (Kerfahi et al. 2016) and South China
64 (Lan et al. 2017a; Lan et al. 2017b; Lan et al. 2017c) have found significant
65 differences between rubber plantations and tropical forests, specifically that the
66 diversity of soil bacteria was higher in rubber plantations than in rainforest. Compared
67 to primary forests, agricultural systems tend to have higher bacterial richness but
68 lower fungal richness (Lan et al., 2017a; Cai et al., 2018; Tripathi et al., 2012; Kerfahi
69 et al., 2016). Compared to Eucalyptus plantations, rubber plantations have been found
70 to possess higher diversity of both bacteria and fungi (Ma et al., 2019). Song et al.
71 (2019) reported that tropical forest conversion to rubber plantation results in reduced
72 fungal microbial community network complexity. However, there are few studies on
73 the impact of forest conversion on soil bacterial community network structure, nor on
74 the drivers leading to changes in network structure.

75 The soil microbial network is viewed as a critical indicator of soil health and
76 quality (Kuperman et al., 2014). Microbial networks consist of two parts: nodes and
77 edges. Nodes indicate microbe types, generally OTUs or species, but can also indicate
78 environmental variables of interest, such as soil pH. Edges (or links) indicate
79 significant correlations between nodes. The number of links connected to a node is
80 known as the node’s degree (Röttjers and Faust, 2018). Usually, the more links a
81 network has, the more complex the community is. Similarly, the higher the degree of

82 a node (OTU or species), the more important the species is to the network structure
83 (Berry and Widder, 2014). In recent years, microbial network analysis has been used
84 to identify keystone taxa (Banerjee et al., 2018) and provide comprehensive insight
85 into the microbial community structure and assembly (Fuhrman, 2019). The
86 complexity of network structure is understood by calculating the number of edges,
87 while keystone species of the community are found by estimating the degree of each
88 species. The stability of the microbial community is determined by calculating the
89 ratio of positive and negative correlation of the network because a large proportion of
90 positive correlations in a microbial community are thought to be unstable (Coyte et al.,
91 2015).

92 Previous work has shown that agricultural intensification can reduce microbial
93 network complexity (Banerjee et al., 2019). Logging alters soil fungal network in
94 tropical rainforests, i.e., a better-organized fungal community in cut stands when
95 compared with primary stands (Chen et al., 2019). Soil bacterial networks are less
96 stable under drought than fungal networks (De Vries et al., 2018). While the exact
97 drivers of microbial network structure still remain unknown. Previous work has
98 shown that soil nutrients, such as soil phosphorus content, and soil pH, are the main
99 drivers for the network structure for microbial communities (Banerjee et al., 2019).
100 Seasonal variation also affects the network structure by changing the identity of
101 keystone species in the community, given that a keystone species might be present
102 only in a specific season or time period (Banerjee et al., 2018).

103 Hainan Island is home to a large area of tropical rainforest rich in biodiversity. It is
104 a part of the Indian-Malay rainforest system at the northern edge of the world's
105 rainforest distribution. However, rubber plantations now account for almost a quarter
106 of the total extent of vegetated areas on Hainan Island (Lan et al., 2021). Previous
107 work on the island has shown that forest conversion from rainforest to rubber
108 plantations can result in shifts in bacterial composition from the Proteobacteria to
109 Chloroflexi, and fungal composition from Basidiomycota to Ascomycota (Lan et al.,

110 2021). Other research has shown microbial (including bacteria and fungi) diversity
111 was higher in rainforest soils than in rubber plantation (Lan et al., 2021). Therefore,
112 we hypothesize the following: (1) the conversion of rainforest to rubber plantations
113 results in changes in the dominant phyla of soil microbes, with concomitant changes
114 in network structure and identity of keystone species. (2) Soil bacterial-fungal
115 network structure is less complex but more stable in rubber plantation than in
116 rainforest because high species diversity leads to complex network structure and
117 unstable microbiome communities (Coyte et al., 2015). (3) Soil bacterial networks in
118 rubber plantations are less complex than in rainforests because intensive cropping
119 reduces the complexity of bacterial network, even as richness increases (Karimi et al.,
120 2019). (4) Soil fungal network structure in rubber plantation sites is more complex
121 than in the rainforest because organic farming showed a much more complex fungal
122 network than conventional or no-tillage farm systems resulting from organic fertilizer
123 and the return of litter to the field (Banerjee et al., 2019). By testing these hypotheses,
124 we want to clarify the drivers and mechanisms of microbial community assembly that
125 link forest conversion to differences in soil microbial network structure. This study
126 will provide critical information for understanding and managing microbial
127 communities in tropical forests of China and elsewhere.

128 **2. Methods**

129 **2.1 Study site**

130 This study was conducted on Hainan Island (18°10'–20°10'N and
131 108°37'–111°03'E), south China. The total area of Hainan Island is about 34,000 km²
132 (Lopez et al., 2009). Hainan Island is the largest island within the Indo-Burma
133 Biodiversity Hotspot of tropical Asia (Francisco-Ortega et al., 2010) and has a
134 tropical monsoon climate. Hainan Island has a warm and humid climate all year round,
135 with an average annual temperature of 22-26°C. Rainfall is abundant, ranging from
136 1,000 mm to 2,600 mm yearly, with an average annual precipitation of 1,639 mm.
137 The rainy season occurs from May to October, with a total precipitation of about 1500

138 mm, accounting for 70-90% of the total annual precipitation. Only 10-30% of the total
139 annual precipitation falls within the dry season, from November to April. The central
140 part of Hainan Island is mountainous and contains old-growth tropical rainforests and
141 monsoon forests. Rubber plantations are found on the plateaus surrounding the central
142 mountainous zone.

143 **2.2 Soil sampling**

144 The tropical rainforests of Hainan are mainly distributed in Bangwangling, Diaoloshan,
145 Wuzhishan, Yinggeling and Jianfengling. Therefore, we selected a tropical rainforest in
146 each of these sites for our study objectives. There are about 530,000 hectares of rubber
147 plantations on Hainan Island. Five rubber plantations were selected, one each in
148 Wanning, Ledong, Danzhou, Haikou, Qiongzong. These five rubber plantations are
149 located in the east, south, west, north and middle of Hainan, respectively. (Figure S1).
150 We selected 25-30 years old rubber trees (i.e., mature rubber plantations) for our study
151 objectives. Latex harvest and fertilizer application are common management practices
152 in rubber plantations. Usually, compound fertilizer (1-1.5 kg per tree) and organic
153 fertilizers (20-25 kg per tree) were applied once or twice a year. More information on
154 the study sites is provided in Table S1. Before soil sampling, we sterilized the soil drill
155 with 75% alcohol. After the removal of the litter layer using a 5-cm diameter steel drill,
156 top soil (0 to 20 cm) was collected, then homogenized and passed through a 2-mm
157 mesh sieve. For each site, thirteen soil samples were collected, thus there were a total
158 of 130 samples collected between the rubber plantation and tropical rainforest sites.
159 Soil sampling was performed twice in 2018, once in January (dry season) and once in
160 July (rainy season). There were thus a total of 260 soil samples (130 per forest type).
161 Soil samples were divided into two parts. One was used to analyze soil water content,
162 soil pH, total nitrogen, total phosphorus (TP), total potassium (TK), nitrate nitrogen
163 (NN), ammonium nitrogen (AN), available phosphorus (AP), potassium (AK). Soil
164 water content (%) was measured gravimetrically. Soil pH was measured in a soil/water
165 suspension (1: 2.5, w/w) using a pH meter. TN was determined using a micro-Kjeldahl

166 digestion followed by steam distillation. TP and TK were measured following
167 digestion with NaOH. NN and AN were determined by steam distillation and
168 indophenol-blue colorimetry, respectively. Soil samples were extracted with NaHCO₃
169 and the extract was then used to measure available soil phosphorus AP via
170 molybdate-blue colorimetry. To measure soil AK, soil was extracted with
171 ammoniumacetate and then the extract was loaded onto an atomic absorption
172 spectrometer with ascorbic acid as a reductant (Chen et al., 2019). Soil properties of the
173 rubber plantation and rainforest sites are shown in Table S2. The other part of each soil
174 sample was stored in ultra-low temperature (- 80 °C) refrigerator for later DNA
175 extraction.

176 **2.3 DNA extraction and PCR amplification**

177 Microbial DNA was extracted from 0.5 g of soil using E.Z.N.A.® Soil DNA Kits
178 (Omega Bio-tek, Norcross, GA, U.S.) following the manufacturer's protocol. The
179 fungal ITS1 hypervariable region was amplified using the PCR primers ITS1F
180 (5'-CTTGGTCATTTAGAGGAAGTAA-3') and ITS2R
181 (5'-GCTGCGTTCTTCATCGATGC-3') (Adams et al., 2013). For bacteria and
182 archaea, the V4 hypervariable region of the bacterial 16S rRNA gene was amplified
183 using the PCR primers 515FmodF (5'-GTGYCAGCMGCCGCGGTAA-3') and
184 806RmodR (5'-GGACTACNVGGGTWTCTAAT-3') (Walters et al., 2016; Sampson
185 et al., 2016). The PCR reactions were conducted using the following approach: an
186 initial 3 min denaturation at 95°C, followed by 27 cycles of 30s at 95°C, 30s of
187 annealing at 55°C, and 45s of elongation at 72°C, and a 10 min final extension at
188 72°C.

189 **2.4 Illumina MiSeq sequencing**

190 Amplicons were extracted from 2% agarose gels, purified using the AxyPrep DNA
191 Gel Extraction Kit (Axygen Biosciences, Union City, CA, U.S.) and quantified using
192 a QuantiFluor™ -ST Fluorometer (Promega, U.S.). Purified amplicons were pooled in

193 an equimolar solution and then sequenced (paired-end, 2×250 bp) on an Illumina
194 MiSeq platform according to standard protocols.

195 Metagenomic shotgun sequencing libraries were prepared and then sequenced by
196 Majorbio, Inc. (Shanghai, China) using the Illumina HiSeq 2000 platform. The NR
197 gene catalog was aligned against the Kyoto Encyclopedia of Genes and Genomes
198 (KEGG) database using BLAST (Version 2.2.28+) and then functionally annotated
199 using KOBAAS 2.0 according to previously described methods (Qin et al., 2010)

200 **2.5 Bioinformatics and data analysis**

201 Raw fastq files were demultiplexed and quality-filtered using QIIME (Caporaso et al.,
202 2010) (version 1.17). Operational Taxonomic Units (OTUs) were clustered with a
203 97% similarity cut-off using UPARSE (Edgar, 2013), and chimeric sequences were
204 identified and removed using UCHIME. Using the RDP Classifier, the phylogenetic
205 affiliation of each 16S rRNA gene and ITS gene sequence was determined using a
206 confidence threshold of 70% with the SILVA 16S rRNA database and UNITE
207 database, respectively (Amato et al., 2013). For each site, the relative abundance of
208 different taxa (Good, 1953) and the Shannon diversity index were calculated based on
209 re-sampled sequence data using MOTHUR (Schloss et al., 2009). The raw reads were
210 deposited into the NCBI Sequence Read Archive (SRA) database (Accession Number:
211 SRP108394, SRP278296, SRP278319).

212 **2.6 Statistical analysis**

213 To make the co-occurrence network analyses simple, we only used the top 300 most
214 abundant bacterial and fungal OTUs to analyze the network structure of the two forest
215 types. These OTUs are roughly match the OTUs with a relative abundance greater
216 than 0.05% (Jiao et al., 2016). Four networks, representing each habitat during each
217 sampling period (rainforest and rubber plantations in the dry and rainy seasons), were
218 constructed with 65 samples each. We also performed the bacterial-fungal community
219 network analysis to investigate soil microbial network complexity of each of the four.

220 Interactions consisted of Spearman's rank correlations. Co-occurrence networks were
221 constructed using only significant correlations of $\rho > 0.6$ ($P < 0.01$) (Barberan et al.
222 2012), because this cutoff includes a range of interactions strengths (De Vries et al.,
223 2018). The networks were then visualized in R using the *igraph* package. To reveal
224 the distribution pattern of correlation coefficients, the frequency of the coefficients of
225 $\rho > 0.3$ ($P < 0.01$) were plotted. The Network Analyzer tool in Cytoscape (version
226 3.4.0) was used to calculate network topology parameters including number of nodes,
227 edges, degree, betweenness, closeness. The number of shared edges and unique edges
228 as well as keystone OTUs were calculated to evaluate whether the network structure
229 changed. We defined a shared network edge as the link (edge) between species A and
230 species B not only existing in rubber plantation network, but also in rainforest
231 network. Similarly, a unique edge was a link that only existing in either rubber
232 plantations or rainforests. The number of shared edges and unique edges were
233 calculated using *igraph* and visualized using Venn diagrams. Keystone OTUs are
234 known to be important for ecosystem structure and function and were selected on the
235 basis of high network degree, high closeness centrality, and low betweenness
236 centrality as defined by Berry and Widder (2014). To evaluate the proportional
237 influence of each phylum on bacterial and fungal network structure, node degrees of
238 each phylum were calculated and bar plots were created. Correlation coefficients
239 between species and functions were calculated based on metagenomics data. Here we
240 used the top 50 most abundant species and top 50 KEGG functions (pathway level 3).
241 Then the species and function correlation network was constructed on the Major bio
242 cloud platform (<https://cloud.majorbio.com/>). To reveal the relationship between
243 microbial taxa and environment variables, two-way correlation networks were also
244 constructed on the Major bio cloud platform. The topological role of each node in a
245 network was assessed by the Z_i and P_i values, where Z_i represents the connectivity
246 of node i within a module, and P_i measures the degree of node i connected with other
247 modules (Roger and Amaral, 2005). All species can be divided into four groups
248 according to the simplified criteria (Olesen et al., 2007), namely peripherals ($Z_i < 2.5$

249 and $P_i < 0.62$), connectors ($P_i > 0.62$), module hubs ($Z_i > 2.5$) and network hubs ($Z_i >$
250 2.5 and $P_i > 0.62$). The Z_i and P_i values were calculated using GIANT package of
251 Cytoscape. The Z_i - P_i plot was created with *ggplot2* in R.

252 **3. Results**

253 **3.1 Bacterial and fungal networks**

254 Our results showed that the bacterial-fungal community network were more complex
255 in rubber plantations (4,284 edges in the dry season, 7,257 in the rainy season) than in
256 primary forests (3,650 in the dry season, 6,507 in the rainy season), and more
257 complex during the rainy season than the dry season (Figure 1). Most nodes of
258 bacterial networks (Figure 2) and fungal networks (Figure 3) varied with forest type
259 in both the dry and rainy seasons. For bacterial networks, there were 2,559 and 2,501
260 edges in the forests and rubber plantations, respectively, in the dry season, but these
261 two networks only shared 262 edges, accounting for only about 10% the total edges
262 (Figure 4A-B). Similarly, these networks only shared 519 edges during the rainy
263 season. For fungal networks, there were only 4 and 5 shared edges (i.e., no more than
264 5% of the total edges) in the dry and rainy seasons, respectively (Figure 4C-D).

265 The number of edges of bacterial and fungal networks were almost equivalent
266 during the dry season. However, in the rainy season, there were more edges in the
267 rainforest bacterial network than in the rubber plantation network (Table 1 & Figure
268 4B). For the network structure of the fungal community, more edges were observed in
269 rubber plantations in the rainy season (Table 1 & Figure 4D). Similarly, there were no
270 significant differences in either the degree of the bacterial and fungal networks
271 between the tropical rainforest sites and rubber plantations in the dry season (Figure
272 5A, C). In the rainy season, rainforest sites had a higher bacterial network degree,
273 while rubber plantations showed higher fungal network degree (Figure 5B, D). For
274 bacterial networks, more nodes (OTUs) with high degree (rubber plantation had 2
275 nodes with degree greater than 75, rainforest had 8 such nodes) were observed in the

276 rainforest during the rainy season (Figure S2B). For fungal networks, 15 nodes of
277 higher degree (degree greater than 25) were observed in rubber plantations, however,
278 only 7 such nodes existed for rainforest sites (Figure S2D). These results indicate that
279 the fungal network structure in the rubber plantations was more complex than for
280 tropical rainforests during the rainy season, but that the reverse was true for bacteria.

281 When considering the ratio of positive to negative correlation coefficients, the
282 bacterial-fungal community network revealed that rubber plantations (844 edges in
283 dry season, 1744 edges in rainy season) have more negative links than rainforest (149
284 edges in dry season, 489 edges in rainy season) indicating that the network of rubber
285 plantation was more stable than that of the rainforests (Figure 1). More correlations
286 (greater than 0.3, $P < 0.01$) were also calculated to investigate the ratio of positive to
287 negative correlation and the results showed that negative correlations between
288 bacterial and fungal OTUs in the rubber plantations were consistently stronger than in
289 the tropical rainforests in both dry season and rainy season (Figure 6).

290 **3.2 Keystone taxa**

291 Module hubs and network hubs have been commonly identified as keystone taxa in
292 networks. However, for both the bacterial and fungal communities, neither tropical
293 rainforest nor rubber plantation networks possessed module hubs and network hubs
294 (Figure S3-S4). The keystone taxa of bacteria were very different between rubber
295 plantations and tropical rainforest sites in both the dry and rainy seasons. For bacteria,
296 there were more keystone taxa in tropical rainforest sites than in the rubber
297 plantations in both the dry and rainy seasons, indicating that the tropical rainforest
298 networks had higher complexity. We found that some groups of Acidobacteria are
299 keystone taxa in tropical rainforest sites but disappeared after forest conversion. There
300 were more Actinobacteria in rubber plantations than in tropical rainforest sites (Table
301 S3).

302 For fungi, more keystone taxa were observed in rubber plantations than in tropical

303 rainforest sites during both the dry and rainy seasons, indicating that the rubber
304 plantation networks were more complex. Most keystone taxa belong to Ascomycota,
305 suggesting that member of this group are very important for network structure. In
306 addition to forest conversion, seasonal changes also affect the keystone taxa of the
307 fungal community network. There were more Basidiomycota OTUs in the dry season,
308 but more Ascomycota in rainy season (Table S4).

309 **3.3 Drivers of microbial composition and network structure**

310 Two-way correlation network analysis of the 300 most abundant OTUs and
311 environmental variables revealed elevation (106 degree), AK (104), and soil pH (86)
312 were the main drivers of bacterial community composition (Figure S5A). For fungi,
313 elevation had the highest degree (61), followed by AK (51) (Figure S5B). The
314 relationship between OTU abundance and soil pH revealed that the soil pH negatively
315 correlated with some species of Acidobacteria, but positively correlated with some
316 species of Chloroflexi and some species of Ascomycota (Figure S6). AK positively
317 correlated with species of Planctomycetes, Verrucomicrobia and Basidiomycota, but
318 negatively correlated with Chloroflexi and Ascomycota. In words, AK and soil pH
319 were important driver for microbial composition.

320 The total degree of each phylum were used to reveal the influence of each phylum
321 on network structure (Figure 7). For bacteria, Proteobacteria, Actinobacteria and
322 Acidobacteria had a large influence on network structure. Acidobacteria and
323 Planctomycetes contributed more to rainforest networks than rubber plantation
324 networks. However, Actinobacteria and Chloroflexi showed the opposite pattern. For
325 fungi, Ascomycota and Basidiomycota had large influences on network structure.
326 Both Ascomycota and Basidiomycota had stronger influences on rainforest networks
327 than rubber plantation networks. The influence of Ascomycota was stronger during
328 the rainy season than in the dry season, indicating seasonal change also had impact on
329 fungal community networks. The degree of each phylum of bacterial and fungal
330 network also revealed seasonality had effects on network degree (Figure S7).

331 **3.4 Interaction between species and functions**

332 Two-way network analysis of the 50 most abundant species (all belonging to bacteria
333 groups based on metagenomic data) and the 50 most abundant KEGG functions
334 revealed the interaction between microbial composition and functions. The results
335 showed that soil network structure at rainforests sites (460 edges in the dry season,
336 1,750 in the rainy season) was more complex than that of rubber plantations (223 in the
337 dry season, 451 in the rainy season) (Figure 8), indicating that more links between
338 microbial species and functions were observed in soils of rainforest sites. Both rubber
339 plantation and rainforest networks were more complex in the rainy season than in the
340 dry season. We also found that metabolism was the most important function in soil
341 microbial networks. Surprisingly, species of Actinobacteria negatively correlated with
342 function in rubber plantations.

343 **4. Discussion**

344 **4.1 Forest conversion reduces soil bacterial network complexity**

345 Land-use changes increasingly threaten biodiversity, particularly in tropical forests
346 (Gibson et al., 2011). However, we still have little understanding of how soil
347 networks respond to forest conversion, such when rainforests are converted to rubber
348 plantations. Our results showed that forest conversion had large effects on both soil
349 bacterial and fungal networks. Tropical rainforest bacterial networks had more edges
350 (Table 1) and a higher degree (Figure 4) than did rubber plantation networks,
351 especially during the rainy season, indicating that the rainforest bacterial network was
352 more complex than the rubber plantation network. This finding is consistent with
353 previous observations that soil bacterial networks were more complex in natural
354 systems than in crop soil (Karimi et al., 2019). Further study showed that soil
355 networks become more connected through the process of ecological restoration
356 (Morriën et al., 2017). The observed decrease in network complexity and cohesion
357 supports the hypothesis that cropping may enhance the isolation of bacterial taxa

358 (Karimi et al., 2019), which results in lower numbers of connections in the network.
359 In addition, at the microscale, the structure of tilled soils is more homogeneous, and
360 the soil pores are less connected than in soils that are not tilled (Pagliai et al., 2004),
361 such as rainforest soil. In nature, soil ecosystems are highly heterogeneous since hot
362 spots of soil microbial biodiversity can form spatial and temporally within soil
363 aggregates (Bach et al., 2018). This spatial heterogeneity likely plays an important
364 role for the interactions among microbes and the mechanisms by which more complex
365 and diverse communities drive various nutrient cycling processes on small spatial
366 scales (Wagg et al., 2018).

367 A large number of studies employing microbial network analysis have enriched
368 our understanding of microbial co-occurrence patterns in various soil ecosystems.
369 However, very little is known of whether differences in the structure of microbial
370 networks have consequences for microbiome functioning (Wagg et al., 2018). Our
371 results demonstrated that more species correlated with metabolism in natural system
372 than in the agricultural system, especially during the rainy season. This is in line with
373 a previous study conducted in Sumatra, Indonesia, which found that the
374 transformation of forest to rubber results in a 10-16% decrease in community
375 metabolism (Barnes et al., 2014). Fewer interactions between microbial species (most
376 of them are bacteria) and functions in rubber plantations demonstrated that forest
377 conversion reduced soil bacterial network complexity.

378 **4.2 Forest conversion increased soil fungal network complexity**

379 Surprisingly, rainforest bacterial networks were characterized by fewer edges (Table 1)
380 and a lower degree (Figure 4) than the rainforest bacterial networks, meaning that
381 rubber plantation bacterial networks were more complex than those of native forests.
382 Our results were not consistent with a study conducted in Xishuangbanna (Song et al.,
383 2019) which showed that tropical forest conversion reduced fungal network complex,
384 but is consistent with previous observations which found that fungal community
385 networks were better organized in disturbed forest compared to primary forest (Chen

386 et al., 2019). Banerjee et al. (2019)'s observation showed that organic agricultural
387 fields harbored much more complex fungal networks with many more keystone taxa
388 than conventionally managed fields. Forest conversion resulted in shifts in fungal
389 composition from Basidiomycota to Ascomycota (Figure S8), as seen in previous
390 investigations (Lan et al., 2021; Lan et al., 2020a). Previous work showed that
391 Basidiomycota species show higher drought sensitivity than Ascomycota species
392 (Taniguchi et al., 2018), this would result in a shift in richness and abundance of
393 Basidiomycota species (Figure S8). Many Basidiomycota species are capable of
394 long-distance dispersal (Egidi et al. 2019, Geml et al., 2012), which may result in a
395 decrease in fungal network complexity. This possibly explained why Ascomycota
396 OTUs contribute more to the network structure than Basidiomycota (Figure 6).
397 Overall, reductions in the abundance and richness of Basidiomycota species led to an
398 increase in fungal links in rubber plantations.

399 **4.3 Forest conversion enhanced the stability of soil network**

400 The ability of a microbial community to resist and recover from disturbances is
401 defined as the stability of the community (De Vries and Ashley, 2013). The ratio of
402 positive to negative network links constitutes the balance between facilitative and
403 inhibitive relationships within a network (Karimi et al., 2017). Theoretical studies, for
404 example, predict that ecological networks that consist of weak interactions are more
405 stable than those with strong interactions (Neutel et al., 2002, Coyte et al., 2015), and
406 that compartmentalization and presence of negative interactions increase the stability
407 of networks under disturbances (Coyte et al., 2015, Rooney et al., 2006, Stouffer &
408 Bascompte 2011). In our study, more negative correlations were detected in rubber
409 plantation, indicating the network structure of rubber plantation soils was more stable
410 than that of rainforest soils (De Vries et al., 2018). Forest conversion from rainforest
411 to rubber plantations resulted in shifts in bacterial composition from Proteobacteria to
412 Chloroflexi, and fungal composition from Basidiomycota to Ascomycota (Lan et al.,
413 2021). Proteobacteria and Basidiomycota are copiotrophic microorganisms, while

414 Chloroflexi and Ascomycota are oligotrophic (Yang et al., 2019; Lan et al., 2021).
415 This indicated forest conversion resulted in shifts in soil microbial composition from
416 more *k*-strategists to more *r*-strategists (De Vries et al., 2013). R-strategists are
417 quick-growing and less resistant but more resilient to disturbances. However, growth
418 is not the only mechanism that could maintain community stability because the
419 response of microorganisms to disturbance are very complex.

420 **4.4 Main drivers for soil microbial composition and network structure**

421 Forest conversion results in the loss of plant diversity, plant biomass and an increase
422 in soil pH (Lan et al., 2017a, 2017b). Rubber plantations had a significantly higher pH,
423 which explains the relative decrease in the abundance of Acidobacteria (Lan et al.,
424 2017a). Our results demonstrate that forest conversion drives a shift in which soil
425 microbes act as keystone species (Table 1). We found that many OTUs of
426 Acidobacteria fit our criteria as keystone species for rainforest sites, which is
427 consistent with previous findings (Banerjee et al., 2018). Unexpectedly, OTU11388
428 and OTU11373, both Acidobacteria, were observed in rainforest soils in both the dry
429 and rainy seasons, indicating Acidobacteria were very important for rainforest soil
430 bacterial networks (Figure 6 and table S3). Higher AK concentrations resulted in a
431 higher abundance and more taxa of Actinobacteria (Figure 9), which suggests that
432 Actinobacteria contributed more in rubber plantation than in rainforest (Figure 6).
433 Indeed, forest conversion reduced the abundance of Actinobacteria OTUs (Figure S9).
434 Due to the human disturbance in rubber plantations, the soil will inevitably be slightly
435 polluted with herbicides and domestic garbage. Previous study showed member of
436 Actinobacteria were observed in contaminated soil (Jiao et al., 2016).

437 Forest conversion also increases land use intensity (Brinkmann et al., 2019),
438 including the application of fertilizer and herbicide. The application of herbicides has
439 also been linked to significant decreases in root colonization and spore biomass of
440 arbuscular mycorrhizal fungi in tropical agriculture (Zaller et al., 2014). Soil nutrient
441 concentrations show a decline around the roots of rubber plantations compared to

442 those from rainforests (Sahner et al., 2015). Our study was no exception. For instance,
443 AK and TN concentration were significant lower in rubber plantations than in
444 samples from rainforest sites (Table S2). Higher concentrations of AK reasonably
445 explained the higher contribution of Basidiomycota to the network structure (Figure
446 8B) due to the positive association of AK with Basidiomycota.

447 Spatiotemporal heterogeneity can be a major driver of the abundance and
448 distribution of keystone taxa in soil, which is a highly heterogeneous and multifaceted
449 environment (Mills et al., 1993, Power et al., 1996; Mouquet et al., 2013). Seasonal
450 variability determines the structural and compositional properties of microbiomes in
451 an environment, and as such, a keystone species might be present only in a specific
452 season or time period (Banerjee et al., 2018). It was interesting that more bacterial
453 OTUs were identified as connectors during the rainy season than the dry season.
454 Connectors have been characterized as generalists (Olesen et al., 2007), and
455 generalists drive covariation among communities in a network (Chen et al., 2019).
456 Previous observations have demonstrated that some keystone taxa that were found in
457 the dry season disappeared during the rainy season (Lan et al., 2018). Seasonal
458 changes possibly explain why some keystone taxa were observed in the rainy season
459 but not in the dry season.

460 **4.5 Possible impact of forest conversion on microbial community function**

461 Forest conversion results in a decrease in the abundance of Proteobacteria and an
462 increase of Actinobacteria. Most species of Proteobacteria were positively correlated
463 with metabolic function, while most of Actinobacteria were negatively associated with
464 metabolic function (Figure 8). Therefore, the changes in the abundance of these two
465 phyla results in a reduction of microbial community function after forest conversion.
466 Because the metabolic function of a specific species is often affected by environmental
467 conditions (Louca et al., 2018), some species are not correlated with any function in the
468 dry season, but correlated with metabolic function in the rainy season, indicating there
469 some microorganisms do not participate in the metabolic process during the dry season,

470 especially for the rubber plantations. In conclusion, forest conversion as well as
471 seasonal variation had a great impact on soil microbial community functions.

472 **5. Summary**

473 Our knowledge about land-use impacts on soil ecosystems is mostly limited to
474 biodiversity and ecosystem functions, leaving uncertainty about how soil networks
475 change after forest conversion. This study is the most comprehensive report on
476 changes in network structure that occur when tropical rainforests are converted into
477 rubber plantations. Our study showed that forest conversion altered both bacterial and
478 fungal soil networks, reduced bacterial network complexity and enhanced fungal
479 network complexity, especially during the rainy season. One possible reason maybe
480 that forest conversion altered soil pH and other soil properties, which altered bacterial
481 composition, subsequent network structure as well as certain soil ecosystem functions.
482 Our study demonstrates the impact of forest conversion on soil network structure,
483 which has important implications for ecosystem functioning, such as metabolic
484 activity, and health of soil ecosystems in tropical regions.

485

486 **Code availability**

487 Not applicable

488 **Authors' contributions**

489 Guoyu Lan: Conceptualization, Methodology, Writing, Reviewing and Editing;
490 Chuan Yang, Rui Sun, Bangqian Chen, Xicai Zhang and Zhixiang Wu: Investigation

491 **Competing interests**

492 The authors declared that they have no conflicts of interest to this study.

493 **Disclaimer**

494 Publisher's note: Copernicus Publications remains neutral with regard to jurisdictional

495 claims in published maps and institutional affiliations.

496 **Acknowledgements**

497 We appreciate Dr. Hogan and another anonymous reviewer very much for their
498 positive and constructive comments and suggestions on our manuscript.

499 **Financial support**

500 This work was supported by National Natural Science Foundation of China
501 (31770661); High level Talents Project of Hainan Natural Science Foundation
502 (320RC733); Finance Science and Technology Project of Hainan Province
503 (ZDYF2019145); the Earmarked Fund for China Agriculture Research System
504 (CARS-33-ZP3).

505

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708

709 **Figure legend**

710 **Figure 1** The soil bacterial-fungal community network of rubber plantations and
711 rainforests in the dry and rainy seasons. Red solid circles indicate fungi, and green
712 solid circles indicate bacteria. Red lines indicate positive correlation between OTUs,
713 and green indicate negative correlation. Absolute value of correlation coefficient $\rho >$
714 $0.6, P < 0.01$

715 **Figure 2** Soil bacterial network structure of rubber plantations and tropical rainforests
716 in dry and rainy seasons. Red lines indicate positive correlation between OTUs, and
717 green indicate negative correlation. Absolute value of correlation coefficient $\rho > 0.6,$
718 $P < 0.01$

719 **Figure 3** Soil fungal network structure of rubber plantations and tropical rainforests
720 in the dry and rainy seasons. Red lines indicate positive correlation between OTUs,
721 and green indicate negative correlation. Absolute value of correlation coefficient $\rho >$
722 $0.6, P < 0.01$

723 **Figure 4** Number of shared and unique edges of soil bacterial and fungal networks in
724 rubber plantations and tropical rainforests in the dry and rainy seasons. The number
725 where the two circles cross is number of shared edges. Numbers in the red circle
726 presents the unique edges in rainforests, while in blue circle present the unique edges
727 in rubber plantations.

728 **Figure 5** Network degree of soil bacterial and fungal communities of rubber
729 plantations (blue) and tropical rainforests (red) in the dry and rainy seasons.

730 **Figure 6** Frequency distributions of correlation strength in bacterial (a: dry season, b:
731 rainy season) and fungal (c: dry season, d: rainy season) networks of rubber
732 plantations and tropical rainforests in the dry and rainy seasons. (absolute correlation
733 coefficient greater than 0.3, $P < 0.01$) Correlations in rainforest networks are red,
734 correlations in rubber plantation networks are blue.

735 **Figure 7** Proportional influence of different phyla on bacterial and fungal network
736 structure in both dry and rainy seasons. The influence was the number of degrees of
737 nodes belonging to a particular phylum. (a: bacteria in dry season, b: bacteria in rainy
738 season, c: fungal in dry season, d: fungal in rainy season).

739 **Figure 8** Network of the top 50 most abundant species (based on metagenomics data)
740 and top 50 most frequent KEGG functions (pathway level 3) of rubber plantations and
741 tropical rainforest sites in the dry and rainy seasons. (A: rubber in dry season; B:
742 rainforest in dry season; C: rubber in rainy season; D: rainforest in rainy season) The
743 size of the node indicates the species/function abundance. A red line indicates positive
744 correlation between species/functions, and green indicates negative correlation.
745 Absolute value of correlation coefficient $> 0.6, P < 0.01$

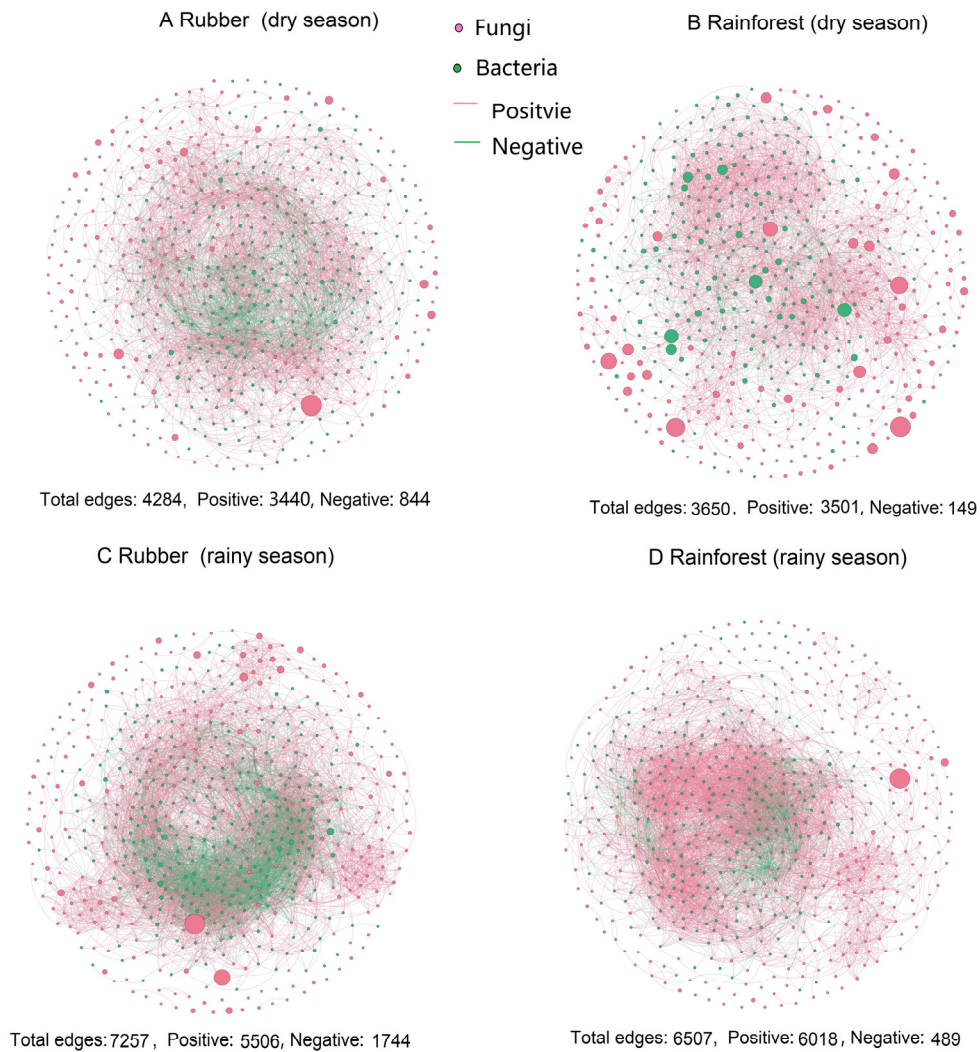
746

747 **Table 1** Topological properties of soil microbial (bacterial and fungi) network
 748 structure in rubber plantation and tropical rain forest in dry and rainy seasons

	Bacteria		Fungi					
	Rubber Dry	Rainforest Dry	Rubber Rainy	Rainforest Rainy	Rubber Dry	Rainforest Dry	Rubber Rainy	Rainforest Rainy
No. of nodes	291	287	296	296	220	235	243	244
No. of edges	2448	2559	4248	5019	791	769	1250	905
No. of positive edges	2052	2508.00	3385	4901	760	764	1195	897
No. of negative edges	396	51	863	118.00	31	5	55	8
Connectance	0.06	0.06	0.09	0.11	0.02	0.02	0.03	0.02
Average degree	16.82	17.83	16.67	33.91	16.67	6.54	10.28	7.41
Average betweenness	100.61	93.52	31.23	42.57	208.90	185.03	170.99	266.31
Average of shortest path length	2.92	2.92	2.56	2.45	4.00	3.626	3.81	5.27
Diameter	6.00	7.00	7.00	6.00	10.00	12.00	9.00	15.00
Cluster of coefficient	0.46	0.451	0.51	0.54	0.59	0.457	0.49	0.51
No of clusters	11.00	14.00	5.00	5.00	94.00	75.00	62.00	67.00
Degree centralization	0.12	0.11	0.17	0.18	0.08	0.08	0.10	0.10
Betweenness centralization.	0.0066	0.0067	0.0053	0.0049	0.02	0.03	0.121	0.04
Closeness centralization.	0.35	0.35	0.40	0.41	0.31	0.351	0.291	0.26
Neighborhood Connectivity	21.12	21.79	34.69	40.68	10.45	9.28	13.78	9.44
Topological coefficient	0.26	0.26	0.26	0.27	0.33	0.36	0.37	0.50

749

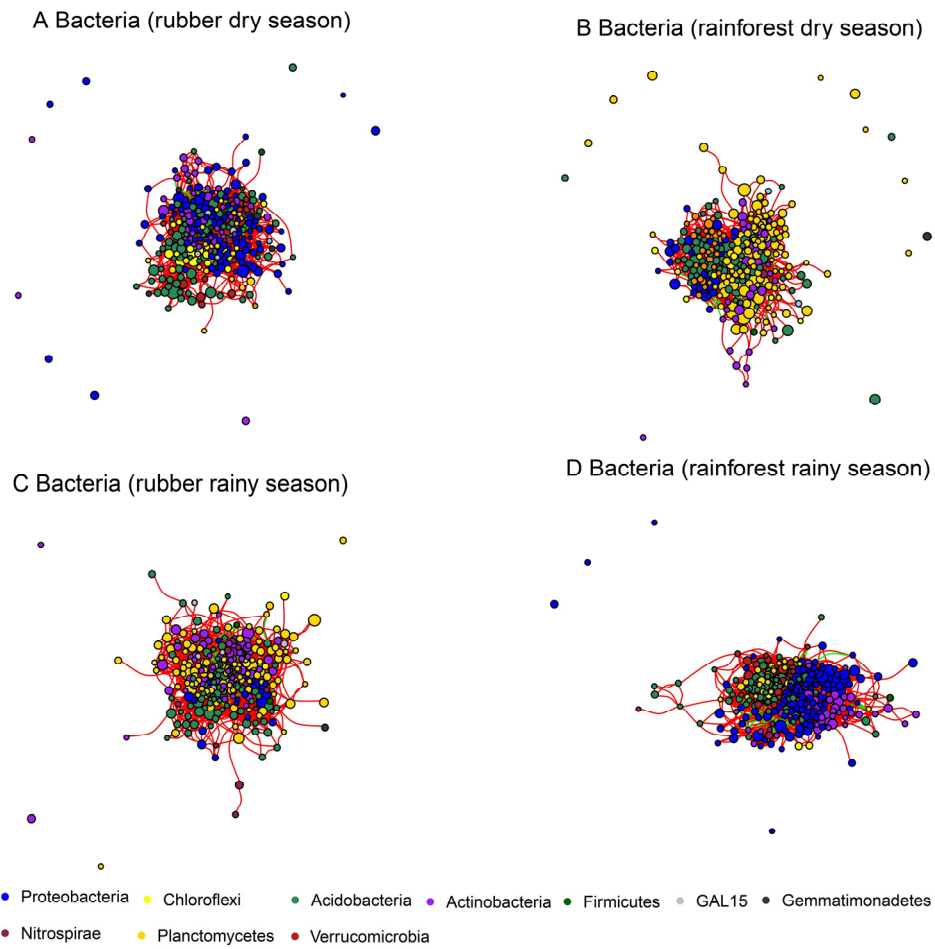
750



751

752 **Figure 1** The soil bacterial-fungal community network of rubber plantations and rainforests in the
 753 dry and rainy seasons. Red solid circles indicate fungi, and green solid circles indicate bacteria. Red
 754 lines indicate positive correlation between OTUs, and green indicate negative correlation. Absolute
 755 value of correlation coefficient $\rho > 0.6, P < 0.01$.

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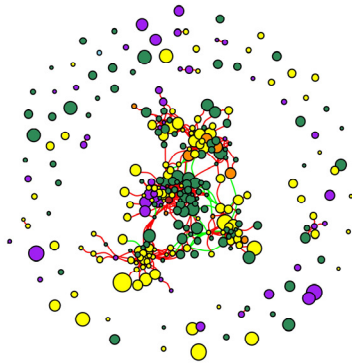


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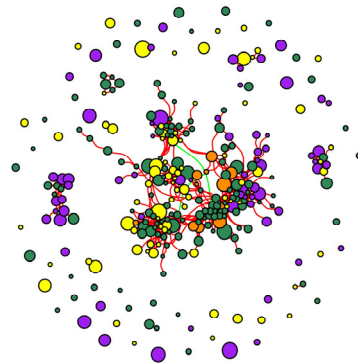
758 **Figure 2** Soil bacterial network structure of rubber plantations and tropical rainforest in the dry
 759 and rainy seasons. Red line indicates positive correlation between OTUs, and green indicates
 760 negative correlation. Absolute value of correlation coefficient $\rho > 0.6$, $P < 0.01$

761

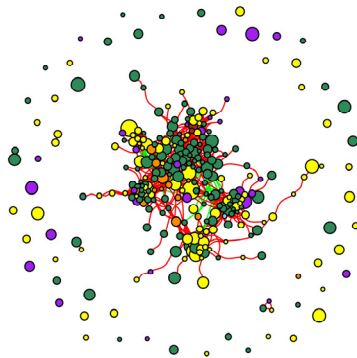
A Fungi (rubber dry season)



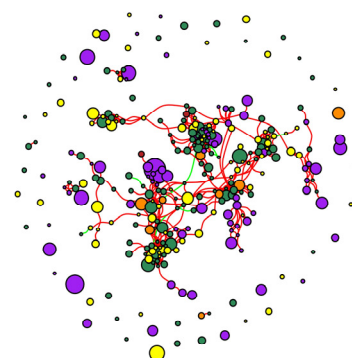
B Fungi (rainforest dry season)



C Fungi (rubber rainy season)



D Fungi (rainforest rainy season)



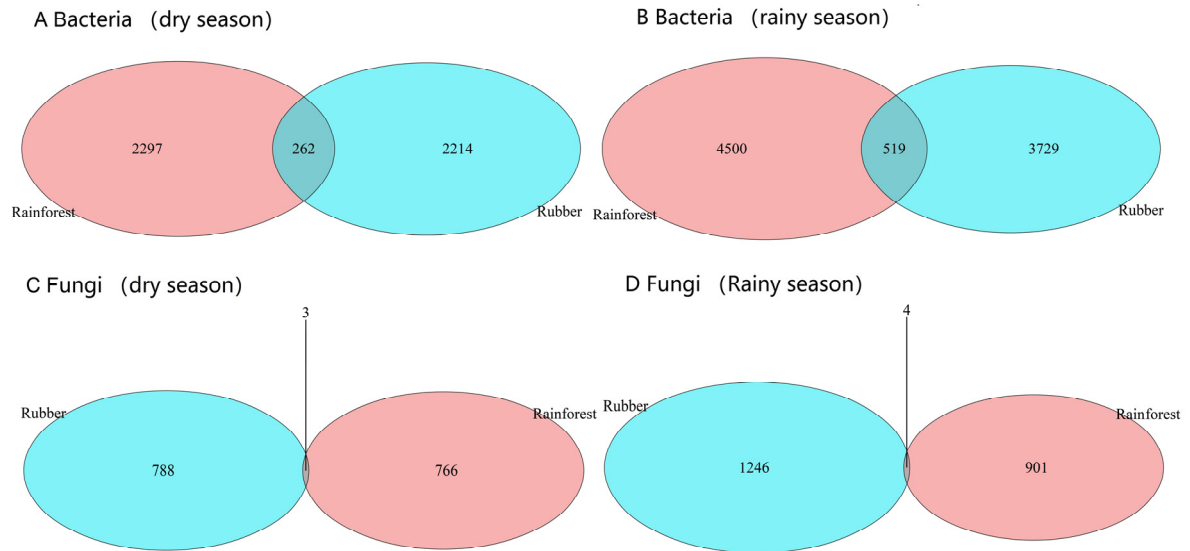
● Ascomycota ● Basidiomycota ● Chytridiomycota ● Rozellomycota ● Zygomycota ● Unclassified

762

763 **Figure 3** Soil fungal network structure of rubber plantations and tropical rainforests in the dry and
764 rainy seasons. Red line indicates positive correlation between OTUs, and green indicates negative

765 correlation. Absolute value of correlation coefficient $\rho > 0.6$, $P < 0.01$

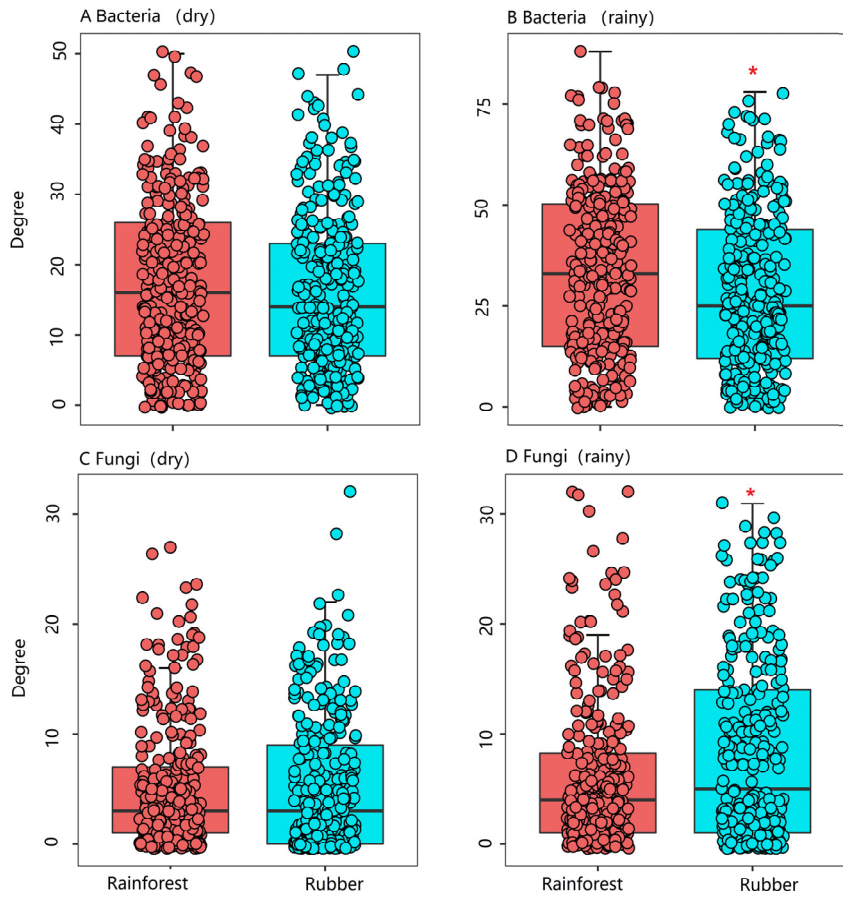
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768 **Figure 4** Number of shared and unique edges of soil bacterial and fungal networks in rubber
 769 plantations and tropical rainforests in the dry and rainy seasons. The number where the two circles
 770 cross is number of shared edges. Numbers in the red circle presents the unique edges in rainforests,
 771 while in blue circle present the unique edges in rubber plantations.

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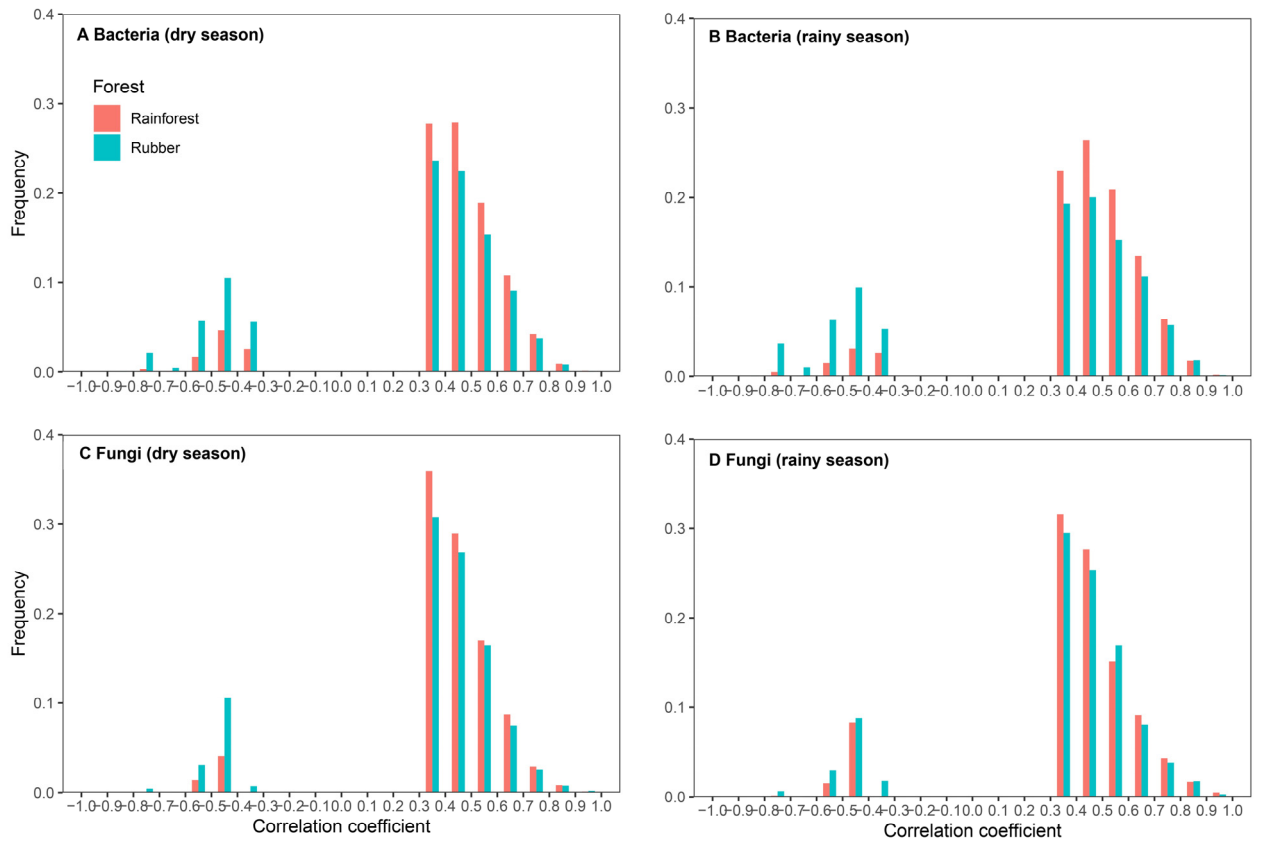
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Figure 5 Network degree of soil bacterial and fungal communities of rubber plantations (blue) and tropical rainforests (red) in the dry and rainy seasons.

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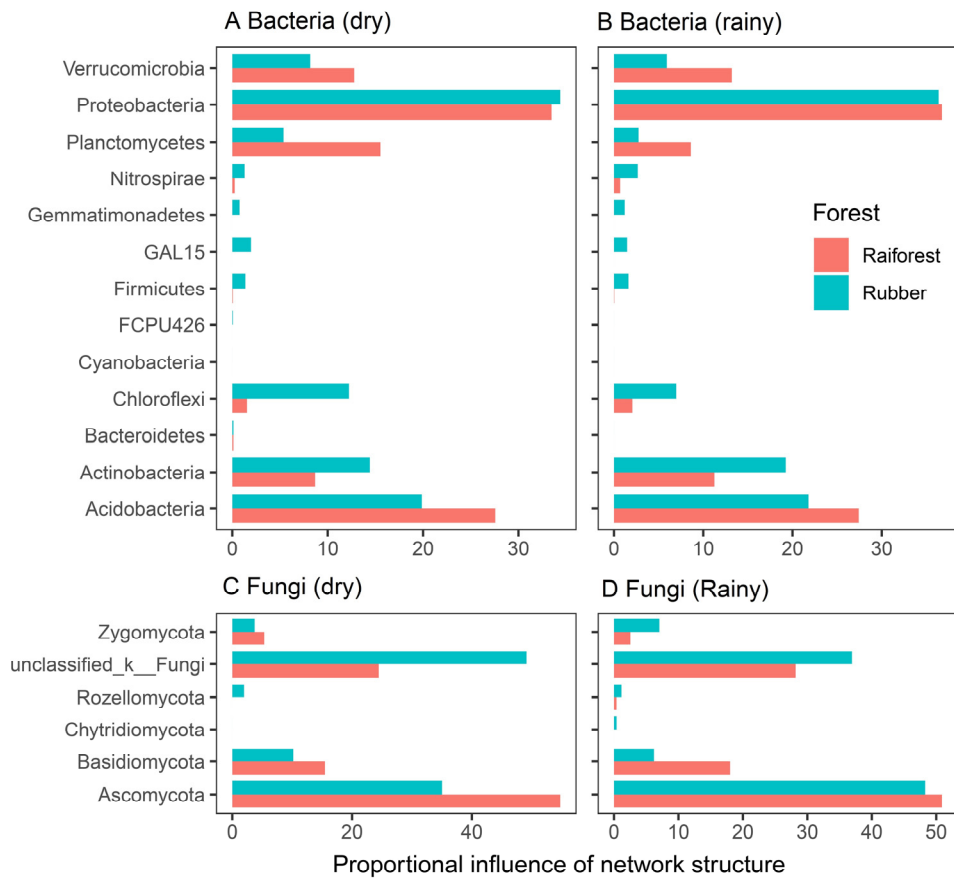
780 **Figure 6** Frequency distributions of correlation strength in bacterial (a: dry season, b: rainy season)

781 and fungal (c: dry season, d: rainy season) networks of rubber plantations and tropical rainforests

782 in the dry and rainy seasons (absolute correlation coefficient greater than 0.3, $P < 0.01$).

783 Correlations in rainforest networks are red, correlations in rubber plantation networks are blue.

784

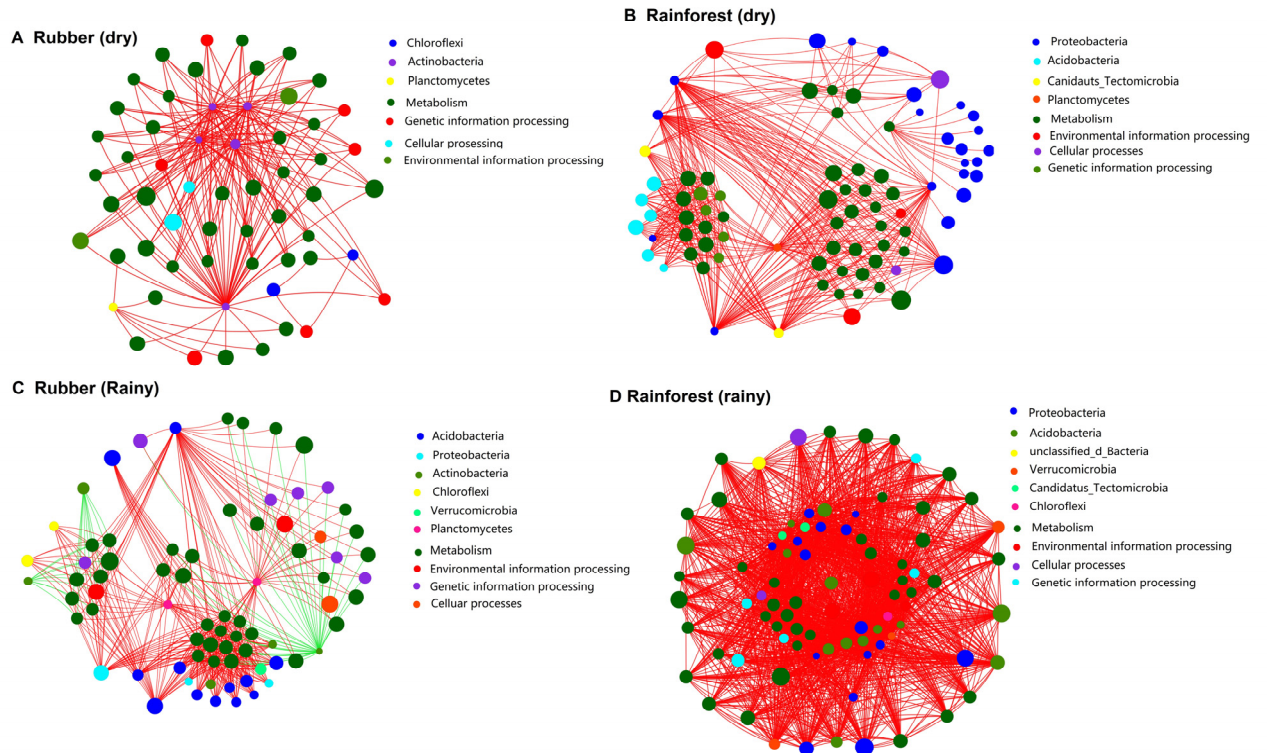


786

787 **Figure 7** Proportional influence of different phyla on bacterial and fungal network structure in
 788 both dry season and rainy season. The influence was the number of degrees of nodes belonging to
 789 a particular phylum. (a: bacteria in dry season, b: bacteria in rainy season, c: fungal in dry season,
 790 d: fungal in rainy season).

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792



793

794 **Figure 8** Network of the top 50 most abundant species (based on metagenomics data) and the top
 795 50 abundant KEGG functions (pathway level 3) of rubber plantations and tropical rainforest in the
 796 dry and rainy seasons (A: rubber in dry season; B: rainforest in dry season; C: rubber in rainy
 797 season; D: rainforest in rainy season). The size of the node indicates the species/function
 798 abundance. Red line indicates positive correlation between species/functions, and green indicates
 799 negative correlation. Absolute value of correlation coefficient > 0.6 , $P < 0.01$