

# What comes after the sun? – On the integration of soil biogeochemical pre-weathering into microplastic experiments

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10 **Abstract.** Recent studies have been engaged in estimating the adverse effects of  
microplastic (MP) on soil quality parameters. Mass concentrations of MP as found in highly  
contaminated soils have been shown to weaken the soil structure, and parts of the edaphon  
are adversely affected by mainly the < 100 µm MP size fraction. However, the vast majority of  
15 these studies used pristine particles, which have surface characteristics different from that of  
environmental MP. Exposed to UV radiation, plastic undergoes photochemical weathering  
with embrittlement and the formation of surface charge leading to an alteration of  
physiochemical behavior. When plastic particles then enter the soil environment, further aging  
factors appear with yet unknown efficacy. This little explored soil biogeochemical phase  
20 includes biofilm cover, decay with enzymes (as shown in laboratory experiments with both  
conventional and biodegradable plastics), contact with biotic and abiotic acids, oxidants as  
well as uptake by the soil fauna that causes physical fragmentation. Such transformation of  
surfaces is assumed to affect soil aggregation processes, soil faunal health and the transport  
of plastic colloids and adsorbed solubles. This perspective article encourages to consider the  
25 weathering history of MP in soil experiments and highlights the need for reproducing the  
surface characteristics of soil MP to conduct laboratory experiments with close-to-nature  
results.

### 30 **Did we neglect biogeochemical aging factors?**

Since the mass production of plastic articles of daily use started in the early 1950<sup>th</sup> (Thompson et al., 2009), a number of processes cause the contamination of ecosystems such as inland and coastal waters, sediments, the open and deep seas, soils and even the atmosphere with MP (e.g. Cole et al., 2011; Woodall et al., 2014; Wu et al., 2018; Büks and Kaupenjohann, 2020; Trainic et al., 2020). The formation of soil MP pools occur through littering and dispersion from landfills, the application of wastewater, contaminated surface water, sewage sludge, composts, digestates, mulching foils, seed and fertilizer coatings, road dust as well as atmospheric deposition (Eerkes-Medrano et al., 2015; Huerta Lwanga et al., 2017a; Weithmann et al., 2018; Corradini et al., 2019; Dierkes et al., 2019; He et al., 2019; Edo et al., 2020; Huang et al., 2020; Bertling et al., 2021; Katsumi et al., 2021; Szewc et al., 2021).

Today, we are faced to a global contamination of soil ecosystems with MP, that averages 1.7 mg kg<sup>-1</sup> dry soil in agricultures (Büks and Kaupenjohann, 2020), exceeds this value by several orders of magnitude in heavily contaminated soils at road sides and industrial areas (Fuller and Gautam, 2016; Dierkes et al., 2019), and reaches even remote areas (Abbasi et al., 2021). Several laboratory studies showed adverse effects of high MP concentrations on the soil fauna (Büks et al., 2020a) and soil structure (e.g. de Souza Machado et al., 2018; de Souza Machado et al., 2019; Liang et al., 2019; Lozano et al., 2021) and underlined the relevance of especially the small-sized fraction (MP<100 µm) (Büks et al., 2020b).

Although these results are rightly alarming due to the function of soil structure and the edaphon as soil fertility parameters (Bronick and Lal, 2005; Thiele-Bruhn et al., 2012), their informative value is limited by the fact, that the vast majority of experiments used pristine plastic and a short run time that does not allow for further weathering (e.g. de Souza Machado et al., 2018; de Souza Machado et al., 2019; Liang et al., 2019; Büks et al., 2020a; Lozano et al., 2021). For a better matching with of environmental conditions, some studies have used photooxidatively weathered plastic. In soil, however, the bulk of MP is additionally exposed to biogeochemical alteration for years. Its surface characteristics and role within soil ecosystems thereby possibly change compared to solely above ground weathering.

### 60 **Underground weathering – a second phase of aging?**

On a microscopic scale, the surfaces of pristine plastic items are normally smooth with nearly no surface charge (e.g. Fotopoulou and Karapanagioti, 2012; Fotopoulou and Karapanagioti, 2015). Exposed to sunlight, depletion of UV absorbers and HALS (hindered amine light stabilizers) leads to enhanced photooxidation (e.g. Kokott, 1989; Pickett, 2018). From the point of view of the macroscopic observer, the plastic becomes less hydrophobic, stiff and more prone to fragmentation by wind and water erosion.

Standardized approaches from materials science are newly used in soil science to reproduce natural photooxidative aging characteristics (BMBF initiative “Plastic in the Environment”, <https://www.bmbf-plastik.de/en>, e.g. Büks et al., 2021). They recommend xenon arc lamps with borosilicate filters, that adjust the emitted spectrum tighter to the natural UV spectrum, or fluorescent UV lamps (DIN EN ISO 4892-2/3: “Plastics – Methods of exposure to laboratory light sources”). The performance of these approaches is enhanced by use of modern daylight filters, a steady temperature of 38°C, relative air humidity of 25 to 50 % and regular washing of the sample surfaces by artificial rain (Pickett, 2018). Beside the use of UV,  $\gamma$ -irradiation treatment is reported to imitate the carbonyl stretch in PE samples similar to a long-term UV-B exposition (Johansen et al., 2019). Furthermore, Zhou et al. (2020) could demonstrate that discharged plasma oxidation (DPO) is likewise suitable to increase surface area, crystallinity and carbonyl indices of plastic particles within hours.

However, it is unknown whether these protocols properly reproduce the additional influence of underground aging, which occurs under different physiochemical conditions. When plastic is exposed to the dark world of soil fauna, microorganisms, roots and frequent leaching, the composition of weathering parameters changes significantly (Table 1). The plastic is now faced to new mechanical stresses such as (bio)turbation, largely moist conditions and exposed to a variety of soil biogeochemical processes.

One of these potential aging factors is the diverse and active soil fauna, that has been shown to ingest, digest and excrete plastic particles (Büks et al., 2020). It is an ensemble of small, mobile bioreactors, that incubate soil particles including MP within a habitat of high microbial diversity – their gastrointestinal tract – and distribute them throughout the soil by excretion. A well known example for this multifaced functionality is the earthworm. Some taxa like woodlice, termites, mealworms and earthworms have been additionally found to comminute plastic by gnawing and, hence, actively produce MP (e.g. Lenz et al., 2012; Zhang et al., 2018; Büks et al., 2020a). There are also indications that the mealworm microbiome is able to degrade not only additives, but also PE and PS polymers (e.g. Brandon et al., 2018).

While moisture evaporates quickly on sun-exposed, heated plastic surfaces, in soils it is the ubiquitous condition for microbial life, extracellular metabolic processes and the release and transport of chemical agents, that react with the plastic outside the fauna. Microbial colonization and biofilm formation on surfaces of MP particles have been shown in studies on various aquatic ecosystems (e.g. Zettler et al., 2013; McCormick et al., 2014; Oberbeckmann et al., 2015; Dussud et al., 2018; Jiang et al., 2018). Recent studies on soil ecosystems have also demonstrated that MP surfaces of different origin are covered with microbial communities. This could hypothetically cause a masking of plastic surface characteristics by the biofilm matrix. The composition of surface MP communities is very different from that of the soil matrix (Chai et al., 2020; Zhang et al., 2019). The altered soil microbial community is thereby not only determined by the physiochemical properties of the surrounding soil, but also by the type of plastic and its additives (Chai et al., 2020; Ng et al., 2020; Wang et al., 2020;

Wiedner and Polifka, 2020; Yan et al., 2020; Yi et al., 2020). This might lead to a physiochemical behavior of plastic particles, that differs not because of the plastic type, but because of its biofilm cover.

110 Although plastic is unlikely to serve as a major substrate for microbes due to its large  
molecular size, high chemical stability, and low bioavailability (Oberbeckmann and Labrenz  
2020), there is indication that biofilm cover causes the alteration of its chemical properties.  
Not only a viscous matrix, that protects bacteria against mechanical stress, predators,  
desiccation and irradiation, biofilm is also an extracellular reaction space that facilitates the  
115 concentration and metabolization of nutrients and the recycling of dead cell material  
(Flemming and Wingender, 2010). For this purpose, manifold extracellular enzymes are  
produced by the biofilm community to decompose food sources or modify the biofilm matrix in  
face of e.g. oxygen or nutrient gradients (Flemming and Wingender, 2010). Among these are  
esterases, proteases and amidases that target on substrates like polysaccharides, proteins,  
120 extracellular DNA, lipids and urea, but also allow the (co)metabolization of artificial polymers  
such as diverse polyesters, ester-based PU and PET in laboratory experiments (Shimao,  
2001; Wei and Zimmermann, 2017; Danso et al., 2019).

Given a poor biodegradability of polymers with C-C backbones and no hydrolysable functional  
groups such as pristine PE, PP, PS and PVC, laboratory experiments showed an unexpected  
125 degradation of PE by a bacterial alkan hydroxylase (Yoon et al., 2012), and, beyond this, the  
specific targeting of PET with a bacterial PETase (Yoshida et al., 2016). In contrast, neither  
degrading enzymes nor observed biodegradation have been reported in case of PP and PVC  
(Danso et al., 2019). Unspecific lignin-degrading enzymes such as laccases, manganese  
peroxidases, hydroquinone peroxidases and lignin peroxidases produced by actinomycetes,  
130 other bacteria as well as fungi, have been further shown to depolymerize even plastics such  
as PE, PS and PA, that are considered recalcitrant (Bhardwaj et al., 2013; Wei and  
Zimmermann, 2017). Beside the direct proof of enzymatic degradation pathways there are  
numerous references on the metabolization of (bio)plastic samples by bacterial and fungal  
strains (e.g. Bhardwaj et al., 2013; Kale et al., 2015; Raziya-fathima et al., 2016; Roohi et al.,  
2017). However, since many studies applied commercial polymers, that have concealed  
135 compositions (Danso et al., 2019), there is often poor insight to what degree the measured  
mass loss is caused by microbial/enzymatic decomposition of the polymer or additives.  
These findings imply, that biodegradation of plastic surfaces in soil is conceivable.

**Table 1:** Development of surface characteristics during the three phases of aging (pristine, photooxidative and soil biogeochemical phase). Data of soil biogeochemical weathering are only known from aquatic systems. (?) marks assumptions based on soil biogeochemical processes found in soils. Some references are: <sup>1</sup>Fotopoulou and Karapanagioti (2012), <sup>2</sup>Fotopoulou and Karapanagioti (2015), <sup>3</sup>ter Halle et al. (2017), <sup>4</sup>Dong et al. (2020), <sup>5</sup>Pickett (2018), <sup>6</sup>Andrady et al. (1993).

characteristic	pristine phase	photooxidative phase	soil biogeochemical phase
topography	smooth <sup>1,2,4</sup>	rough <sup>5</sup>	rough <sup>1,2,4</sup>
surface charge, carbonyl index	no <sup>1,2,3,4</sup>	yes <sup>6</sup>	increasing <sup>1,2,3,4</sup>
crystallinity, crosslinks, chain scissions	low <sup>3</sup>	high <sup>5</sup>	increasing <sup>3,4</sup>
biofilm cover	low	low	growing or mature <sup>2,5</sup>
aging factors	no	UV radiation <sup>5</sup> blue/violet spectrum <sup>5</sup> frequent leaching <sup>5</sup> wind/water erosion	enzymes <sup>(?)</sup> organic acids <sup>(?)</sup> inorganic acids <sup>(?)</sup> bases <sup>(?)</sup> oxidants <sup>(?)</sup> bioturbation <sup>(?)</sup> feeding by the edaphon <sup>(?)</sup> frequent leaching <sup>(?)</sup> freeze-thaw-cycles <sup>(?)</sup>

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Beside the soil biome, soil born acids, bases and oxidants are expected to directly influence the belowground alteration of plastic surfaces. While – to the best of our knowledge – there has been no systematic examination of the effects of such agents within natural ranges of concentration and time of exposure, the treatment of plastic with highly concentrated reagents caused damaging effects from color leaching and expansion to total dissolution (Enders et al., 2017). However, pre- and post-treatment with oxidants such as H<sub>2</sub>O<sub>2</sub> are common parts of the extraction of MP from soil samples with density fractionation (Büks and Kaupenjohann, 2020).

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In winter, when the biotic effects are reduced, freeze-thaw-cycles might be an additional factor of fragmentation. Studies on the effect of alternating freezing and thawing on the structure of plastic surfaces are sparse and only focus on composite materials that include non-plastic components (Wang et al., 2007; Adhikary et al., 2009; Zhou et al., 2014). However, water, that has entered cracks of brittle plastic, most likely contributes to its fragmentation through freezing and expansion and, thus, increases the surface area exposed to other aging factors.

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## Pre-weathering under soil conditions: A methodology for future approaches?

160 While plastics are considered persistent, the above in-vitro experiments indicate, that degradation in soil is possible. The difference in factors leading to photooxidative and biochemical weathering make it plausible for MP surface characteristics to develop differently in above- and belowground environments. If the physicochemical behavior of the microplastic is significantly affected by this, the effects must be considered in the design of laboratory and field experiments. The focus on surfaces is particularly important in studies on (1) soil aggregation and structure, that strongly depend on biofilm cover and surface charge/polarity of the involved primary particles, (2) adverse effects on the soil fauna, that might be  
165 influenced by particle shape and sorption of (in)organic pollutants, (3) interactions with plants and microorganisms as well as (4) the transport of colloidal MP within the soil pore space.

170 However, there are currently no studies that evaluate the efficacy of specific soil biogeochemical aging mechanisms. Recent work only showed the alteration of plastic surfaces during environmental weathering, indicating that future experiments have to be conducted with pre-weathered instead of pristine MP. It is still an open question, if there is effective soil biogeochemical aging beyond the photooxidative phase or whether the DIN EN ISO 4892-2/3 approach, as applied in recent work, is sufficient to imitate soil weathering conditions in future studies.

175 Only a few studies have integrated soil biogeochemical factors into pre-weathering approaches of artificial MP so far (Table 2, Büks et al., 2020a), alas fragmentary, heterogeneous and often directly applied to pristine plastic: Tsunoda et al. (2010) heated plastic items within a water bath at 90 °C for 3 weeks and abraded the surface prior to feeding experiments with termites. This treatment was aimed to make the surface more accessible for gnawing and might also extract soluble additives from the pristine plastic. In another  
180 experiment, the formation of biofilms on MP surfaces was induced by four weeks of incubation in seawater to make the material more attractive as a food source for the lugworm *Arenicola marina* (Gebhardt and Forster, 2018), an approach that can be likewise applied with soil solution. In order to remove soluble substances and fine particles from artificial MP, pristine plastics have been treated with organic solvents (Huerta Lwanga et al., 2016; Huerta Lwanga et al., 2017b; Rodrigues-Seijo et al. 2018; Rodrigues-Seijo et al., 2019; Wang et al.,  
185 2019; Yang et al., 2019). If the plastic type is prone to the solvents, the surface is roughened by the dissolution of oligomers and, thus, increased. However, these techniques are not assumed to increase the number of carbonyl groups and surface charge. Therefore, they do not change the interaction with the soil matrix and the soil fauna, and have never been tested  
190 for similarity to natural weathering.

In contrast, some authors avoided artificial aging and instead applied natural weathering over shorter periods between two weeks and 12 month, which can be used as a kind of “plastic breeding” (e.g. Martin-Closas et al., 2016; Zhang et al., 2018). This treatment changes the

195 physiochemical characteristics of plastics similar to environmental short-term weathering  
 belowground and is suitable for the alteration of large amounts of plastic. But, it is very costly  
 in terms of time when the production of strongly weathered MP is needed.

200 Once we know the important biogeochemical aging factors, long-term weathering  
 experiments will be extremely helpful to understand the dynamics of surface alteration of soil  
 MP. These experiments must take into account not only ecosystem parameters (e.g. humidity,  
 edaphon activity and soil organic carbon) and start conditions such as plastic type, particle  
 surface and protection by specific additives. The increase of surface area and charge density  
 over time might cause a non-linear aging, while biofilm-cover cloaks the real MP surface  
 characteristics – issues that should also be carefully included into the experimental design.

205 There is a great incentive to develop pre-weathering approaches to create designer-MP for  
 laboratory experiments. Those close-to-nature weathering protocols might contain full chains  
 of aboveground and in-soil aging factors and can be diversified according to actual material  
 and environmental conditions. Applied to coming experiments, they will help us to better  
 understand and predict short- and long-term effects of soil MP, the concentration of which is  
 the result of decades of contamination and is still increasing.

**Table 2:** Approaches of surface (pre)weathering in recent experiments with soil microplastic. The abbreviations used in this table are as follows: UV – ultraviolet, TBBPA – tetrabromobisphenol A, FE – feeding experiment. Polymers: BD – biodegradable plastics, OP – oxodegradable plastics, PA – polyamide, PE – polyethylene, PO – polyolefins, PP – polypropylene, PVC – polyvinyl chloride, TCE – thermoplastic copolyester elastomers. NA denotes that information was not available.

aging factor	applied plastic type	aging time (d)	resulting characteristics	experimental focus	reference
UV radiation (climate chamber)	diverse	variable	photooxidative aging	diverse	DIN EN ISO 4892-2, DIN EN ISO 4892-3
gamma irradiation ( <sup>60</sup> Co source)	PE, PP	NA	photooxidative aging	cation adsorption	Johansen et al. (2019)
discharged plasma oxidation (DPO)	PVC	0.02	photooxidative aging	TBBPA adsorption of and toxicity to algae	Zhou et al. (2020)
wather bath (90°C) + abrasion	PO, PA, PE, TCE	21	extraction of additives, increased accessibility for feeding organisms	feeding experiment with termites	Tsunoda et al. (2010)
incubation in seawater	PA, PS	28	surface biofilm formation	FE lugworms	Gebhardt and Forster (2018)
incubation in aquatic systems	PE, PP	19	surface biofilm formation	cation adsorption	Johansen et al. (2019)
methanol treatment	PE, PS	NA	extract soluble additives	FE earthworms	Wang et al. (2019)
ethanol treatment	PE	NA	extract soluble additives	FE earthworms	Rodrigues-Seijo et al. (2018)
pentane + octane treatment	PE	NA	extract soluble additives	FE earthworms	Rodrigues-Seijo et al. (2019)
		NA	extract soluble additives	FE earthworms	Huerta Lwanga et al. (2016)
		NA	extract soluble additives	FE earthworms	Huerta Lwanga et al. (2017b)
plastic nursing (soil)	BD, OD, PE	NA	extract soluble additives	FE earthworms	Yang et al. (2019)
		~150	belowground weathering	mulch foil degradation experiment	Martin-Closas et al. (2016)
		14-365	belowground weathering	feeding experiment with earthworms	Zhang et al. (2018)

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### **Data availability**

All of the data are published within this paper.

### **Author contributions**

215 FB developed the article concept, collected data and prepared the manuscript. MK supervised the study by participating in structural discussions on the idea and concept of the paper and the final corrections.

### **Competing interests**

220 The authors declare that they have no conflict of interest.



## References

- Abbasi, S., Turner, A., Hoseini, M. and Amiri, H.: Microplastics in the Lut and Kavir Deserts, Iran, *Environ. Sci. Technol.*, 55(9), 5993-6000, <https://doi.org/10.1021/acs.est.1c00615>, 2021.
- 225 Adhikary, K. B., Pang, S. and Staiger, M. P.: Effects of the accelerated freeze-thaw cycling on physical and mechanical properties of wood flour-recycled thermoplastic composites, *Polym. Compos.*, 31(2), 185-194, <https://doi.org/10.1002/pc.20782>, 2009.
- Andrady, A. L., Pegram, J. E. and Tropscha, Y.: Changes in carbonyl index and average molecular weight on embrittlement of enhanced-photodegradable polyethylenes, *J. Environ. Polym. Degrad.*, 1(3), 171-179, <https://doi.org/10.1007/bf01458025>, 1993.
- 230 Bertling, J., Zimmermann, T. and Rödiger, L.: *Kunststoffe in der Umwelt: Emissionen in landwirtschaftlich genutzte Böden*, Fraunhofer Institut, UMSICHT Bericht, 2021.
- Bhardwaj, H., Gupta, R. and Tiwari, A.: Communities of microbial enzymes associated with biodegradation of plastics, *J. Polym. Environ.*, 21(2), 575-579, <https://doi.org/10.1007/s10924-012-0456-z>, 2013.
- 235 Brandon, A. M., Gao, S. H., Tian, R., Ning, D., Yang, S. S., Zhou, J., Wu, W. M. and Criddle, C. S.: Biodegradation of polyethylene and plastic mixtures in mealworms (larvae of *Tenebrio molitor*) and effects on the gut microbiome, *Environ. Sci. Technol.*, 52(11), 6526-6533, <https://doi.org/10.1021/acs.est.8b02301>, 2018.
- Bronick, C. J. and Lal, R.: Soil structure and management: a review. *Geoderma*, 124(1-2), 3-22, <https://doi.org/10.1016/j.geoderma.2004.03.005>, 2005.
- 240 Büks, F. and Kaupenjohann, M.: Global concentrations of microplastics in soils – a review, *SOIL*, 6, 649–662, <https://doi.org/10.5194/soil-6-649-2020>, 2020.
- Büks, F., van Schaik, N., and Kaupenjohann, M.: What do we know about how the terrestrial multicellular soil fauna reacts to microplastic?, *SOIL*, 6, 245–267, <https://doi.org/10.5194/soil-6-245-2020>, 2020a.
- 245 Büks, F., van Schaik, N. L., and Kaupenjohann, M.: Mikroplastik aus Klärschlämmen hat das Potential Bodenleben zu schädigen, *KW Korrespondenz Wasserwirtschaft*, <https://doi.org/10.3243/kwe2020.09.001>, 2020b.
- Büks, F., Kayser, G., Zieger, A., Lang, F. and Kaupenjohann, M.: Particles under stress: ultrasonication causes size and recovery rate artifacts with soil-derived POM but not with microplastics, *Biogeosciences*, 18, 159–167, <https://doi.org/10.5194/bg-18-159-2021>, 2021.
- 250 Chai, B., Li, X., Liu, H., Lu, G., Dang, Z. and Yin, H.: Bacterial communities on soil microplastic at Guiyu, an E-Waste dismantling zone of China, *Ecotox. Environ. Safe.*, 195, 110521, <https://doi.org/10.1016/j.ecoenv.2020.110521>, 2020.
- Cole, M., Lindeque, P., Halsband, C. and Galloway, T. S.: Microplastics as contaminants in the marine environment: a review, *Mar. Pollut. Bull.*, 62(12), 2588-2597, <https://doi.org/10.1016/j.marpolbul.2011.09.025>, 2011.
- 255 Corradini, F., Meza, P., Eguiluz, R., Casado, F., Huerta-Lwanga, E., and Geissen, V.: Evidence of microplastic accumulation in agricultural soils from sewage sludge disposal, *Sci. Total Environ.*, 671, 411–420, <https://doi.org/10.1016/j.scitotenv.2019.03.368>, 2019.
- 260 Danso, D., Chow, J. and Streit, W. R.: Plastics: environmental and biotechnological perspectives on microbial degradation, *Appl. Environ. Microbiol.*, 85(19), e01095-19, <https://doi.org/10.1128/AEM.01095-19>, 2019.
- de Souza Machado, A. A., Lau, C. W., Till, J., Kloas, W., Lehmann, A., Becker, R. and Rillig, M. C.: Impacts of microplastics on the soil biophysical environment, *Environ. Sci. Technol.*, 52(17), 9656-9665, <https://doi.org/10.1021/acs.est.8b02212>, 2018.
- 265 de Souza Machado, A. A., Lau, C. W., Kloas, W., Bergmann, J., Bachelier, J. B., Faltin, E., Becker, R., Görlich, A. S. and Rillig, M. C.: Microplastics can change soil properties and affect plant performance, *Environ. Sci. Technol.*, 53(10), 6044-6052, <https://doi.org/10.1021/acs.est.9b01339>, 2019.
- Dierkes, G., Lauschke, T., Becher, S., Schumacher, H., Földi, C. and Ternes, T.: Quantification of microplastics in environmental samples via pressurized liquid extraction and pyrolysis-gas chromatography, *Anal. Bioanal. Chem.*, 411, 6959–6968, <https://doi.org/10.1007/s00216-019-02066-9>, 2019.

- 270 Dong, M., Zhang, Q., Xing, X., Chen, W., She, Z. and Luo, Z.: Raman spectra and surface changes of microplastics weathered under natural environments, *Sci. Total Environ.*, 139990, <https://doi.org/10.1016/j.scitotenv.2020.139990>, 2020.
- Dussud, C., Meistertzheim, A. L., Conan, P., Pujo-Pay, M., George, M., Fabre, P., Coudane, J., Higgs, P., Elineau, A., Pedrotti, M. L., Gorsky, G. and Ghiglione, J. F.: Evidence of niche partitioning among bacteria living on plastics, organic particles and surrounding seawaters, *Environ. Pollut.*, 236, 807-816, <https://doi.org/10.1016/j.envpol.2017.12.027>, 2018.
- 275 Edo, C., González-Pleiter, M., Leganés, F., Fernández-Piñas, F. and Rosal, R.: Fate of microplastics in wastewater treatment plants and their environmental dispersion with effluent and sludge, *Environ. Pollut.*, 259, 113837, <https://doi.org/10.1016/j.envpol.2019.113837>, 2020.
- 280 Eerkes-Medrano, D., Thompson, R. C. and Aldridge, D. C.: Microplastics in freshwater systems: a review of the emerging threats, identification of knowledge gaps and prioritisation of research needs, *Water res.*, 75, 63-82, <https://doi.org/10.1016/j.watres.2015.02.012>, 2015.
- Enders, K., Lenz, R., Beer, S. and Stedmon, C. A.: Extraction of microplastic from biota: recommended acidic digestion destroys common plastic polymers, *ICES J. Mar. Sci.*, 74(1), 326-331, <https://doi.org/10.1093/icesjms/fsw173>, 2017.
- 285 Flemming, H. C. and Wingender, J.: The biofilm matrix, *Nat. Rev. Microbiol.*, 8(9), 623-633, <https://doi.org/10.1038/nrmicro2415>, 2010.
- Fotopoulou, K. N. and Karapanagioti, H. K.: Surface properties of beached plastic pellets, *Mar. Environ. Res.*, 81, 70-77, <https://doi.org/10.1016/j.marenvres.2012.08.010>, 2012.
- 290 Fotopoulou, K. N. and Karapanagioti, H. K.: Surface properties of beached plastics, *Environ. Sci. Pollut. Res.*, 22(14), 11022-11032, <https://doi.org/10.1007/s11356-015-4332-y>, 2015.
- Fuller, S. and Gautam, A.: A procedure for measuring microplastics using pressurized fluid extraction, *Environ. Sci. Technol.*, 50, 5774–5780, <https://doi.org/10.1021/acs.est.6b00816>, 2016.
- 295 Gebhardt, C. and Forster, S.: Size-selective feeding of *Arenicola marina* promotes long-term burial of microplastic particles in marine sediments, *Environ. Pollut.*, 242, 1777–1786, <https://doi.org/10.1016/j.envpol.2018.07.090>, 2018.
- He, P., Chen, L., Shao, L., Zhang, H. and Lü, F.: Municipal solid waste (MSW) landfill: A source of microplastics? -Evidence of microplastics in landfill leachate, *Water res.*, 159, 38-45, <https://doi.org/10.1016/j.watres.2019.04.060>, 2019.
- 300 Huang, Y., Liu, Q., Jia, W., Yan, C., and Wang, J.: Agricultural plastic mulching as a source of microplastics in the terrestrial environment, *Environ. Pollut.*, 260, 114096, <https://doi.org/10.1016/j.envpol.2020.114096>, 2020.
- Huerta Lwanga, E., Gertsen, H., Gooren, H., Peters, P., Salánki, T., van der Ploeg, M., Besseling, E., Koelmans, A. A. and Geissen, V.: Microplastics in the terrestrial ecosystem: implications for *Lumbricus terrestris* (*Oligochaeta*, *Lumbricidae*), *Environ. Sci. Technol.*, 50, 2685–2691, <https://doi.org/10.1021/acs.est.5b05478>, 2016.
- 305 Huerta Lwanga, E., Vega, J. M., Quej, V. K., de los Angeles Chi, J., del Cid, L. S., Chi, C., Segura, G. E., Gertsen, H., Salánki, T., van der Ploeg, M., Koelmans, A. A., and Geissen, V.: Field evidence for transfer of plastic debris along a terrestrial food chain, *Sci. Rep.-UK*, 7, 14071, <https://doi.org/10.1038/s41598-017-14588-2>, 2017a.
- 310 Huerta Lwanga, E., Gertsen, H., Gooren, H., Peters, P., Salánki, T., van der Ploeg, M., Besseling, E., Koelmans, A. A. and Geissen, V.: Incorporation of microplastics from litter into burrows of *Lumbricus terrestris*, *Environ. Pollut.*, 220, 523–531, <https://doi.org/10.1016/j.envpol.2016.09.096>, 2017b.
- Jiang, P., Zhao, S., Zhu, L. and Li, D.: Microplastic-associated bacterial assemblages in the intertidal zone of the Yangtze Estuary, *Sci. Total Environ.*, 624, 48-54, <https://doi.org/10.1016/j.scitotenv.2017.12.105>, 2018.
- 315 Johansen, M. P., Cresswell, T., Davis, J., Howard, D. L., Howell, N. R. and Prentice, E.: Biofilm-enhanced adsorption of strong and weak cations onto different microplastic sample types: Use of spectroscopy, microscopy and radiotracer methods, *Water Res.*, 158, 392-400, <https://doi.org/10.1016/j.watres.2019.04.029>, 2019.
- 320 Kale, S. K., Deshmukh, A. G., Dudhare, M. S. and Patil, V. B.: Microbial degradation of plastic: a review, *J. Biochem. Technol.*, 6(2), 952-961, ISSN 0974-2328, 2015.

- Katsumi, N., Kusube, T., Nagao, S. and Okochi, H.: Accumulation of microcapsules derived from coated fertilizer in paddy fields. *Chemosphere*, 267, 129185, <https://doi.org/10.1016/j.chemosphere.2020.129185>, 2021.
- Kockott, D.: Natural and artificial weathering of polymers, *Polym. Degrad. Stabil.*, 25(2-4), 181-208, [https://doi.org/10.1016/S0141-3910\(89\)81007-9](https://doi.org/10.1016/S0141-3910(89)81007-9), 1989.
- 325 Lenz, M., Creffield, J. W., Evans, T. A., Kard, B., Vongkaluang, C., Sornnuwat, Y., Lee, C.-Y., Yoshimura, T. and Tsunoda, K.: Resistance of polyamide and polyethylene cable sheathings to termites in Australia, Thailand, USA, Malaysia and Japan: a comparison of four field assessment methods, *Int. Biodeter. Biodegr.*, 66, 53–62, <https://doi.org/10.1016/j.ibiod.2011.11.001>, 2012.
- 330 Liang, Y., Lehmann, A., Ballhausen, M., Muller, L. and Rillig, M. C.: Increasing temperature and microplastic fibers jointly influence soil aggregation by saprobic fungi, *Front. Microbiol.*, 10:2018, <https://doi.org/10.3389/fmicb.2019.02018>, 2019.
- Lozano, Y. M., Lehnert, T., Linck, L. T., Lehmann, A. and Rillig, M. C.: Microplastic shape, polymer type, and concentration affect soil properties and plant biomass, *Front. Plant Sci.*, 12, 169, <https://doi.org/10.3389/fpls.2021.616645>, 2021.
- 335 Martin-Closas, L., J., Costa, A., Cirujeda, Aibar, J., Zaragoza, C., A., Pardo, Suso, M. L., Moreno, M. M., Moreno, C., Lahoz, I., Macua, J. I. and Pelacho, A. M.: Above–soil and in-soil degradation of oxo- and bio-degradable mulches: a qualitative approach, *Soil Res.* 54, 225–236, <https://doi.org/10.1071/SR15133>, 2016.
- McCormick, A., Hoellein, T. J., Mason, S. A., Schlupe, J. and Kelly, J. J.: Microplastic is an abundant and distinct microbial habitat in an urban river, *Environ. Sci. Technol.*, 48(20), 11863-11871, <https://doi.org/10.1021/es503610r>, 2014.
- 340 Ng, E. L., Lin, S. Y., Dungan, A. M., Colwell, J. M., Ede, S., Lwanga, E. H., Meng, K., Geissen, V., Blackall, L. L. and Chen, D.: Microplastic pollution alters forest soil microbiome, *J. Hazard. Mater.*, 124606, <https://doi.org/10.1016/j.jhazmat.2020.124606>, 2020.
- Oberbeckmann, S., Löder, M. G. and Labrenz, M.: Marine microplastic-associated biofilms—a review, *Environ. Chem.*, 12(5), 551-562, <https://doi.org/10.1071/EN15069>, 2015.
- Oberbeckmann, S. and Labrenz, M.: Marine microbial assemblages on microplastics: diversity, adaptation, and role in degradation, *Annu. Rev. Mar. Sci.*, 12, 209-232, <https://doi.org/10.1146/annurev-marine-010419-010633>, 2020.
- Pickett, J. E.: Weathering of plastics, in *Handbook of Environmental Degradation of Materials* (pp. 163-184), William Andrew Publishing, Oxford, ISBN 978-0-323-52472-8, 2018.
- 350 RaziyaFathima, M., Praseetha, P. K. and Rimal, I. R. S.: Microbial degradation of plastic waste: a review, *J Pharm ChemBiol Sci* 2016; 4(2):231-242, 4, 231-42, ISSN 2 348-7658, 2016.
- Rodríguez-Seijo, A., da Costa, J. P., Rocha-Santos, T., Duarte, A. C. and Pereira, R.: Oxidative stress, energy metabolism and molecular responses of earthworms (*Eisenia fetida*) exposed to low-density polyethylene microplastics, *Environ. Sci. Pollut. R.*, 25, 33599–33610, <https://doi.org/10.1007/s11356-018-3317-z>, 2018.
- 355 Rodríguez-Seijo, A., Santos, B., da Silva, E. F., Cachada, A. and Pereira, R.: Low-density polyethylene microplastics as a source and carriers of agrochemicals to soil and earthworms, *Environ. Chem.*, 16, 8–17, <https://doi.org/10.1071/EN18162>, 2019.
- Roohi, M., Bano, K., Kuddus, M., R Zaheer, M., Zia, Q., F Khan, M., Gupta, A. and Aliev, G.: Microbial enzymatic degradation of biodegradable plastics, *Curr. Pharm. Biotechnol.*, 18(5), 429-440, <https://doi.org/10.2174/1389201018666170523165742>, 2017.
- 360 Shimao, M.: Biodegradation of plastics, *Curr. Opin. Biotech.*, 12(3), 242-247, [https://doi.org/10.1016/S0958-1669\(00\)00206-8](https://doi.org/10.1016/S0958-1669(00)00206-8), 2001.
- Szewc, K., Graca, B. and Dołęga, A.: Atmospheric deposition of microplastics in the coastal zone: Characteristics and relationship with meteorological factors, *Sci. Total Environ.*, 761, 143272, <https://doi.org/10.1016/j.scitotenv.2020.143272>, 2021.
- 365 ter Halle, A., Ladirat, L., Martignac, M., Mingotaud, A. F., Boyron, O. and Perez, E.: To what extent are microplastics from the open ocean weathered?, *Environ. Pollut.*, 227, 167-174, <https://doi.org/10.1016/j.envpol.2017.04.051>, 2017.
- 370 Thiele-Bruhn, S., Bloem, J., de Vries, F. T., Kalbitz, K. and Wagg, C.: Linking soil biodiversity and agricultural soil management, *Curr. Opin. Environ. Sustain.*, 4(5), 523-528, <https://doi.org/10.1016/j.cosust.2012.06.004>, 2012.

- Thompson R. C., Swan S. H., Moore C. J. and vom Saal F.S.: Our plastic age, *Phil. Trans. R. Soc. B.*, 364, 1973–1976, <https://doi.org/10.1098/rstb.2009.0054>, 2009.
- 375 Trainic, M., Flores, J. M., Pinkas, I., Pedrotti, M. L., Lombard, F., Bourdin, G., Gorsky, G., Boss, E., Rudich, Y., Lombard, F., Vardi, A. and Koren, I.: Airborne microplastic particles detected in the remote marine atmosphere, *Nature Comm. Earth Env.*, 1(1), 1-9, <https://doi.org/10.1038/s43247-020-00061-y>, 2020.
- Tsunoda, K., Rosenblat, G. and Dohi, K.: Laboratory evaluation of the resistance of plastics to the subterranean termite *Coptotermes formosanus* (Blattodea: Rhinotermitidae), *Int. Biodeter. Biodegr.*, 64, 232–237, <https://doi.org/10.1016/j.ibiod.2009.12.008>, 2010.
- 380 Wang, W. H., Wang, Q. W., Xiao, H. and Morrell, J. J.: Effects of moisture and freeze-thaw cycling on the quality of rice-hull-PE composite, *Pigment. Resin Technol.*, Vol. 36, No. 6, pp. 344-349, <https://doi.org/10.1108/03699420710831764>, 2007.
- Wang, J., Coffin, S., Sun, C., Schlenk, D. and Gan, J.: Negligible effects of microplastics on animal fitness and HOC bioaccumulation in earthworm *Eisenia fetida* in soil, *Environ. Pollut.*, 249, 776–784, <https://doi.org/10.1016/j.envpol.2019.03.102>, 2019.
- Wang, J., Huang, M., Wang, Q., Sun, Y., Zhao, Y. and Huang, Y.: LDPE microplastics significantly alter the temporal turnover of soil microbial communities, *Sci. Total Environ.*, 138682, <https://doi.org/10.1016/j.scitotenv.2020.138682>, 2020.
- 390 Wei, R. and Zimmermann, W.: Microbial enzymes for the recycling of recalcitrant petroleum-based plastics: how far are we?, *Microb. Biotechnol.*, 10(6), 1308-1322, <https://doi.org/10.1111/1751-7915.12710>, 2017.
- Weithmann N., Möller J. N., Löder M. G., Piehl S., Laforsch C., and Freitag R.: Organic fertilizer as a vehicle for the entry of microplastic into the environment, *Sci. Adv.*, 4, eaap8060, <https://doi.org/10.1126/sciadv.aap8060>, 2018.
- 395 Wiedner, K. and Polifka, S.: Effects of microplastic and microglass particles on soil microbial community structure in an arable soil (Chernozem), *SOIL*, 6, 315–324, <https://doi.org/10.5194/soil-6-315-2020>, 2020.
- Woodall, L. C., Sanchez-Vidal, A., Canals, M., Paterson, G. L., Coppock, R., Sleight, V., Calafat, A., Rogers, A.D., Narayanaswamy, B.E. and Thompson, R. C.: The deep sea is a major sink for microplastic debris, *R. Soc. Open Sci.*, 1(4), 140317, <https://doi.org/10.1098/rsos.140317>, 2014.
- 400 Wu, C., Zhang, K. and Xiong, X.: Microplastic pollution in inland waters focusing on Asia, In: Wagner M., Lambert S. (eds) *Freshwater Microplastics* (pp. 85-99), *The Handbook of Environmental Chemistry*, vol 58, Springer, Cham, [https://doi.org/10.1007/978-3-319-61615-5\\_5](https://doi.org/10.1007/978-3-319-61615-5_5), 2018.
- Yan, Y., Chen, Z., Zhu, F., Zhu, C., Wang, C. and Gu, C.: Effect of polyvinyl chloride microplastics on bacterial community and nutrient status in two agricultural soils, *B. Environ. Contam. Tox.*, 1-8, <https://doi.org/10.1007/s00128-020-02900-2>, 2020.
- 405 Yang, X., Lwanga, E. H., Bemani, A., Gertsen, H., Salanki, T., Guo, X., Fu, H., Xue, S., Ritsema, C. and Geissen, V.: Biogenic transport of glyphosate in the presence of LDPE microplastics: A mesocosm experiment, *Environ. Pollut.*, 245, 829–835, <https://doi.org/10.1016/j.envpol.2018.11.044>, 2019.
- Yi, M., Zhou, S., Zhang, L. and Ding, S.: The effects of three different microplastics on enzyme activities and microbial communities in soil, *Water Environ. Res.*, 93(1), 24-32, <https://doi.org/10.1002/wer.1327>, 2020.
- 410 Yoon, M. G., Jeon, H. J. and Kim, M. N.: Biodegradation of polyethylene by a soil bacterium and AlkB cloned recombinant cell, *J Bioremed. Biodegrad.*, 3(4), 1-8, <https://doi.org/10.4172/2155-6199.1000145>, 2012.
- Yoshida, S., Hiraga, K., Takehana, T., Taniguchi, I., Yamaji, H., Maeda, Y., Toyohara, K., Miyamoto, K., Kimura, Y. and Oda, K.: A bacterium that degrades and assimilates poly (ethylene terephthalate), *Science*, 351(6278), 1196-1199, <https://doi.org/10.1126/science.aad6359>, 2016.
- 415 Zettler, E. R., Mincer, T. J. and Amaral-Zettler, L. A.: Life in the “plastisphere”: microbial communities on plastic marine debris, *Environ. Sci. Technol.*, 47(13), 7137-7146, <https://doi.org/10.1021/es401288x>, 2013.
- Zhang, L., Sintim, H. Y., Bary, A. I., Hayes, D. G., Wadsworth, L. C., Anunciado, M. B. and Flury, M.: Interaction of *Lumbricus terrestris* with macroscopic polyethylene and biodegradable plastic mulch, *Sci. Total Environ.*, 635, 1600–1608, <https://doi.org/10.1016/j.scitotenv.2018.04.054>, 2018.
- 420 Zhang, M., Zhao, Y., Qin, X., Jia, W., Chai, L., Huang, M. and Huang, Y.: Microplastics from mulching film is a distinct habitat for bacteria in farmland soil, *Sci. Total Environ.*, 688, 470-478, <https://doi.org/10.1016/j.scitotenv.2019.06.108>, 2019.

- 425 Zhou, X., Huang, S., Su, G., Yu, Y. and Chen, L.: Freeze-thaw cycles weathering degrading properties of bamboo flour-polypropylene foamed composites, Transactions of the Chinese Society of Agricultural Engineering, 30(10), 285-292, <https://doi.org/10.3969/j.issn.1002-6819.2014.10.036>, 2014.
- Zhou, L., Wang, T., Qu, G., Jia, H. and Zhu, L.: Probing the aging processes and mechanisms of microplastic under simulated multiple actions generated by discharge plasma, J. Hazard. Mater., 398, 122956, <https://doi.org/10.1016/j.jhazmat.2020.122956>, 2020.