



1 **Lower functional redundancy in “narrow” than “broad” functions in global soil**  
2 **metagenomics**

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24 **Abstract**

25 Understanding the relationship between soil microbial taxonomic compositions and  
26 functional profiles is essential for predicting ecosystem functions under various  
27 environmental disturbances. However, even though microbial communities are sensitive  
28 to disturbance, ecosystem functions remain relatively stable, as soil microbes are likely to  
29 be functionally redundant. Microbial functional redundancy may be more associated with  
30 “broad” functions carried out by a wide range of microbes, than with “narrow” functions  
31 specialized by specific microorganisms. Thus, a comprehensive study to evaluate how  
32 microbial taxonomic compositions correlate with “broad” and “narrow” functional  
33 profiles is necessary. Here, we evaluated soil metagenomes worldwide to assess whether  
34 functional and taxonomic diversities differ significantly between the five “broad” and the  
35 five “narrow” functions that we chose. Our results revealed that compared with the five  
36 “broad” functions, soil microbes capable of performing the five “narrow” functions were  
37 more taxonomically diverse, and thus their functional diversity was more dependent on  
38 taxonomic diversity, implying lower levels of functional redundancy in “narrow”  
39 functions. Co-occurrence networks indicated that microorganisms conducting “broad”  
40 functions were positively related, but microbes specializing “narrow” functions were  
41 interacting mostly negatively. Our study provides strong evidence to support our  
42 hypothesis that functional redundancy is significantly different between “broad” and  
43 “narrow” functions in soil microbes, as the association of functional diversity with  
44 taxonomy were greater in the five “narrow” rather than the five “broad” functions.  
45



46 **Keywords** Functional redundancy, Soil metagenomics, Functional traits, Taxonomic  
47 compositions,  
48

## 49 **1. Introduction**

50 Microbial communities often exhibit incredible taxonomic diversity, with one gram of  
51 soil harboring millions of microbial species (Gans et al., 2005). However, how such  
52 diversity governs microbial functional potential and ecosystem processes is largely  
53 unknown. Though microbial taxonomic composition is generally sensitive to disturbance  
54 and often do not rapidly recover (Allison and Martiny, 2008), it is unclear how changes  
55 in microbial community composition would regulate ecosystem functioning. Mechanistic  
56 understanding of microbial systems, including microbial taxonomic compositions and  
57 functional potential, is essential for predicting ecosystem functioning under various  
58 environmental disturbances (Torsvik and Øvreås, 2002; Wellington et al., 2003; McGill et  
59 al., 2006).

60 Though microbial community composition usually shift in response to disturbance,  
61 ecosystem functions could remain relatively stable due to functional redundancy (Allison  
62 and Martiny, 2008). Microbial functional redundancy is an inevitable emergent property  
63 of microbial systems (Louca et al., 2018), as some metabolic functions can be performed  
64 by multiple species, which may thus be substitutable in certain ecosystem processes  
65 (Rosenfeld, 2002), implying that microbial taxonomy and function can be decoupled  
66 (Louca et al., 2016; Louca et al., 2017). Microbial functional redundancy has been mainly  
67 observed in “broad” ecosystem processes (Yin et al., 2000; Rousk et al., 2009; Banerjee et  
68 al., 2016), but is perhaps less significant in “narrow” functions specialized by certain



69 microorganisms (Schimel, 1995;Balsler et al., 2002). However, some studies simulating  
70 microbial diversity reduction and physiological processes challenged the hypothesis of  
71 microbial redundancy in soil microbes (Peter et al., 2011;Philippot et al., 2013;Delgado-  
72 Baquerizo et al., 2016). Such apparent contradictory results suggest the degree of  
73 functional redundancy may depend on the function of interest. Microbes conducting  
74 “broad” metabolic functions, such as carbon decomposition, are likely to distribute across  
75 most taxa (Crowther et al., 2019) and associate with high level of functional redundancy  
76 (Beier et al., 2017;Rivett and Bell, 2018). “Narrow” functions, such as nitrification or  
77 methanogenesis, may be restricted to a few phylogenetic clades (Schimel and Gullede,  
78 1998), and are hypothesized to exhibit less redundancy than “broad” functions (Schimel,  
79 1995;Rocca et al., 2015). Today, multifunctionality (Hector and Bagchi, 2007) has to be  
80 accounted for to avoid overestimating functional redundancy (Gamfeldt et al., 2008). By  
81 assessing multiple functions, the relationship between microbial diversity and ecosystem  
82 function can be better quantified in the soil (Bastida et al., 2016;Delgado-Baquerizo et  
83 al., 2016).

84 Nowadays, metagenomics have been increasingly used as a promising comparative  
85 tool (Tringe et al., 2005) to study the relationship between functional and taxonomic  
86 diversities (Fierer et al., 2012a;Fierer et al., 2012b;Fierer et al., 2013;Pan et al., 2014;Leff  
87 et al., 2015;Souza et al., 2015). The growing wealth of soil metagenome data thus poised  
88 well to aid in the generalization of global patterns of microbial attributes and  
89 standardizing frameworks for consistent representation of microbial community (Chen et  
90 al., 2021a;Chen et al., 2021b). However, a synthetic metagenomic analysis to assess how



91 general microbial taxonomic and functional diversities differ between “broad” and  
92 “narrow” functions across the globe is still lacking.

93 Here, we constructed soil metagenomic datasets of taxonomic and functional  
94 diversities of five “broad” and five “narrow” functions across seventeen climate zones.  
95 We typically chose SEED Subsystems database (Overbeek et al., 2013) that has diverse  
96 classification at level 1, allowing us to conduct comparison between “broad” versus  
97 “narrow” functions. We selected five “narrow” functions, namely N (Nitrogen  
98 Metabolism), P (Phosphorous Metabolism), K (Potassium Metabolism), S (Sulfur  
99 Metabolism), and Fe (Iron Acquisition and Metabolism). These are typical functional  
100 categories of specific nutrient cycling in Subsystems Level 1 and are only performed by  
101 certain groups of soil microbes (Schimel, 1995). The five “broad” functions selected were  
102 AAD (Amino Acids and Derivatives), CHO (Carbohydrates), CBS (Clustering-based  
103 Subsystems), CVPGP (Cofactors, Vitamins, Prosthetic Groups, Pigments), and Protein  
104 (Protein Metabolism), which are the most abundant functional categories in Subsystems  
105 level 1, and represent broad-scale functions acquired by a relatively larger group of  
106 diverse soil microbes (Balsler et al., 2002). We further constructed the pairwise similarity  
107 of function and taxonomy based on the relative abundance of functional and taxonomic  
108 compositions, respectively, for the five “broad” and the five “narrow” functions. We  
109 hypothesized that the taxonomic similarity of soil microbes would be more linearly  
110 correlated to the functional similarity for the five “narrow” functions in comparison to the  
111 five “broad” functions. Therefore, using these global soil metagenomes, our objective  
112 was to test whether the taxonomic compositions of soil microbes that conduct the five  
113 “narrow” functions are more dependent on the functional compositions, leading to a



114 lower level of functional redundancy in the “narrow” functions than the “broad”  
115 functions.

116

## 117 **2. Materials and Methods**

### 118 **2.1. Data collection**

119 To ensure that the quality and completeness of the metagenomes analyzed were of  
120 standard, we carefully selected soil metagenomes in MG-RAST server that have been  
121 published in peer-reviewed journals. We searched peer-reviewed publications from 2012  
122 to 2018 from the Web of Science database using search terms such as “soil  
123 metagenome”, “shotgun sequencing”, and “MG-RAST” to source the metagenomic data  
124 used in this study to their publications. We included soil metagenomes publicly available  
125 in the MG-RAST database that are generated using shotgun sequencing without  
126 amplification or that were directly deposited by peer-reviewed studies into the MG-  
127 RAST database. We then extracted data matrix of taxonomic and functional compositions  
128 of soil metagenomes from MG-RAST public server (<https://www.mg-rast.org/>) based on  
129 the Study ID and/or MG-RAST ID reported in the publications. Details of each soil  
130 metagenome extracted from publications and MG-RAST database was given in  
131 Supplementary Table S1.

132 The functional database that we used in this study, SEED Subsystems, is a  
133 categorization system which organizes gene functional categories into a hierarchy with  
134 three levels of resolution (Level 3, 2 and 1) (Overbeek et al., 2013). To download the  
135 taxonomic compositions to soil microbes to conduct “broad” and “narrow” functions, for  
136 each soil metagenome, in the ‘Analysis’ function of the MG-RAST server



137 (<https://www.mg-rast.org/mgmain.html?mgpage=analysis>), we loaded both SEED  
138 Subsystems (Level 3, 2 and 1) as functional profiles and RefSeq (Tatusova et al., 2013)  
139 databases (genus, family, order, class, and phylum levels) as taxonomic compositions  
140 (Chen et al., 2021b). The detailed protocols of MG-RAST server were followed to  
141 analyze the metagenomic functions (Meyer et al., 2008; Wilke et al., 2017). To obtain the  
142 taxonomic compositions of soil microbes that conduct the selected “broad” and “narrow”  
143 functions, we chose ‘RefSeq’ as source and ‘genus’ as level, and in ‘function filter’ we  
144 added the functional categories in Subsystems Level 1 that we are interested in, including  
145 five “broad” functions of AAD (Amino Acids and Derivatives), CHO (Carbohydrates),  
146 CBS (Clustering-based Subsystems), CVPGP (Cofactors, Vitamins, Prosthetic Groups,  
147 Pigments), and Protein (Protein Metabolism), of which the relative abundance was 5-  
148 13%. The functions of AAD, CHO, CBS, CVPGP, and Protein were the most abundant  
149 functional categories in Subsystems Level 1, which were used to represent broad-scale  
150 functions acquired by a large group of diverse soil microbes. Correspondingly, five  
151 “narrow” functions were chosen, namely N (Nitrogen Metabolism), P (Phosphorous  
152 Metabolism), K (Potassium Metabolism), S (Sulfur Metabolism), and Fe (Iron  
153 Acquisition and Metabolism), of which the relative abundance was 0.8-1.4%, as these are  
154 typical functional categories of specific nutrient cycling in Subsystems Level 1 and are  
155 only performed by certain groups of soil microbes. Total hits of taxonomic compositions  
156 of soil microbes conducting each function at Subsystems Level 1 were calculated as the  
157 sums of hits in different taxonomic categories at RefSeq genus level.

158 The comparative metagenomic analyses were performed using default settings  
159 (maximum e-value cutoff =  $1e^{-5}$ , minimum identity cutoff = 60%, and minimum



160 alignment length = 50) (Meyer et al., 2008). We then merged the taxonomic compositions  
161 of data matrix of each functions extracted from different studies together to generate new  
162 datasets of microbial taxonomic compositions annotated by the RefSeq database. The  
163 reason why we chose the Subsystems database for functional grouping rather than KEGG  
164 Orthology (KO) (Kanehisa et al., 2015), Clusters of Orthologous Groups (COG)  
165 (Galperin et al., 2014), and Non-supervised Orthologous Groups (NOG) (Huerta-Cepas et  
166 al., 2015) databases was that Subsystems had more diverse classification at Level 1,  
167 allowing us to conduct direct comparison between “broad” versus “narrow” functions.  
168 We chose RefSeq database rather than the traditional ribosomal RNA databases, such as  
169 RDP (Ribosomal Database Project) (Cole et al., 2008), Greengenes (DeSantis et al.,  
170 2006), or Silva LSU/SSU (Pruesse et al., 2007) databases, because taxonomic hits in the  
171 RefSeq database were over 1000-fold higher than the rRNA databases, rendering the  
172 resolution comparable to functional hits for comparison between “broad” and “narrow”  
173 functions. To increase the coverage of our datasets, soil metagenomes with/without  
174 assembly were both included.

175 The geographic coordinates of latitudes (LAT) and longitudes (LONG) of each soil  
176 metagenome were directly obtained from publications. Based on LAT and LONG,  
177 climate data of mean annual temperature (MAT) and precipitation (MAP) of study sites  
178 for each soil metagenome were extracted from the WorldClim dataset (Fick and Hijmans,  
179 2017) using the R package ‘raster’ (Hijmans et al., 2015). To examine how microbial  
180 taxonomic diversities of “broad” and “narrow” functions differ globally, soil  
181 metagenomic data was classified into seventeen climate zones based on the main





182 classification of Koeppen-Geiger Climatic Zones (Kottek et al., 2006) using the R  
183 package ‘kgc’ (Bryant et al., 2017).

184

## 185 **2.2. Statistical Analyses**

186 To minimize bias caused by different sequencing depths and read lengths among studies,  
187 we standardize the hits of each taxonomic (or functional) category in each data to relative  
188 abundance by dividing them by the total number of hits. To calculate the pairwise  
189 similarity of taxonomy based on the relative taxonomic abundance at genus level of  
190 microbes conducting the five “broad” and five “narrow” functions, we calculated Bray-  
191 Curtis similarity following log transformation of the compositional taxonomic data by  
192 constructing pairwise Bray-Curtis similarity matrix between each pair of two samples for  
193 each functional categories at Subsystems database at Level 1, which were further  
194 transformed to lists of pairwise Bray-Curtis similarities ordered by sample pair names in  
195 PRIMER 7 (Plymouth Routines in Multivariate Ecological Research Statistical Software,  
196 v7.0.13, PRIMER-E Ltd, UK) (Clarke and Gorley, 2015). To calculate the pairwise  
197 similarity of function, based on the functional abundance at function gene level within  
198 each of the five “broad” and five “narrow” functions, we calculated Bray-Curtis  
199 similarity following log transformation of the compositional functional data by  
200 constructing pairwise Bray-Curtis similarity matrix between each pair of two samples for  
201 each functional categories at Subsystems database at Level 1, which were further  
202 transformed to lists of pairwise Bray-Curtis similarities ordered by sample pair names in  
203 PRIMER 7. To examine the relationship between functional and taxonomic diversities,  
204 Pearson’s correlations were constructed between the transformed lists of pairwise Bray-



205 Curtis similarity of soil metagenomes annotated using Subsystems database at Level 3  
206 (Function) and the RefSeq database at genus level (Taxonomy). The approaches for  
207 processing the relative abundance of compositional data follow the requirements (Gloor  
208 et al., 2017). To analyze the taxonomic composition structures of soil metagenomes  
209 annotated using the RefSeq database at genus level (Taxonomy) of the five “broad” and  
210 five “narrow” functions, PCoA (principal coordinates analysis) and PERMANOVA  
211 (Permutational multivariate analysis of variance) were conducted using the pairwise  
212 Bray-Curtis similarity matrix in PRIMER 7.

213 To compare microbial taxonomic compositions among the five “broad” and the five  
214 “narrow” functions, one-factor PERMANOVA was conducted using the main test and pair-  
215 wise test in PRIMER 7 with *P* values and Sq. root reported. Pearson’s correlations were  
216 constructed to assess the relationships between functional and taxonomic diversities in  
217 the “broad” and “narrow” functions with adjusted P-Square reported. A RELATE  
218 analysis was also performed to evaluate the relatedness among “broad” and “narrow”  
219 functions by calculating a Spearman’s Rho correlation coefficient in PRIMER 7. To  
220 examine the relative abundance of dominant microbial at phylum and class level (mean >  
221 1%) among the five “broad” and five “narrow” functions, heatmaps were constructed  
222 using HeatMapper (Babicki et al., 2016). One-way analysis of variance (ANOVA) with *P*  
223 values adjusted by Bonferroni-correction for multiple comparisons was conducted using  
224 SPSS 22.0 software (Chicago, IL, USA) to evaluate the differences in the relative  
225 abundance of dominant taxonomic compositions (mean > 1%) among climate zones after  
226 the normality of residues and homogeneity of variance were checked using Shapiro-Wilk  
227 and Levene test, respectively. The significance level was set at  $\alpha=0.05$  unless otherwise



228 stated. To calculate the statistical difference between the relative abundance of dominant  
229 microbial taxonomic groups (mean > 1%) in the “broad” and “narrow” functions, LEfSe  
230 (linear discriminant analysis effect size) method was used  
231 (<http://huttenhower.sph.harvard.edu/lefse/>) (Segata et al., 2011). Venn’s diagrams were  
232 constructed to visualize the amount of dominant microbial taxonomic groups at genus  
233 levels or network nodes shared between the five “broad” and the five “narrow” functions  
234 using InteractiVenn (Heberle et al., 2015).

235 To find out potential interactions of microbial taxonomic compositions between  
236 “broad” and “narrow” functions across the globe, co-occurrence network analysis was  
237 performed using the Molecular Ecological Network Analyses Pipeline  
238 (<http://ieg4.rccc.ou.edu/MENA/>) (Zhou et al., 2011;Deng et al., 2012). To make the  
239 minimum observed value close to but no less than 1 as required by the pipeline, the data  
240 of relative abundance were multiplied by  $10^6$ , which would not change the correlation  
241 coefficients. The data matrix of transformed data matrix was uploaded to construct the  
242 network with default settings, including (1) keeping only the species present in more than  
243 a half of all samples; (2) only filling with 0.01 in blanks with paired valid values; (3)  
244 taking logarithm with recommended similarity matrix of Pearson’s correlation  
245 coefficient; and (4) calculation ordered to decrease the cutoff from top using regress  
246 poisson distribution only. A default cutoff value (similarity threshold,  $S_T$ ) for the  
247 similarity matrix was used to assign a link between the pair of species. After that, the  
248 global network properties, the individual nodes' centrality, and the module separation and  
249 modularity were analyzed based on default settings using greedy modularity  
250 optimization. Network files were exported and visualized using Cytoscape software



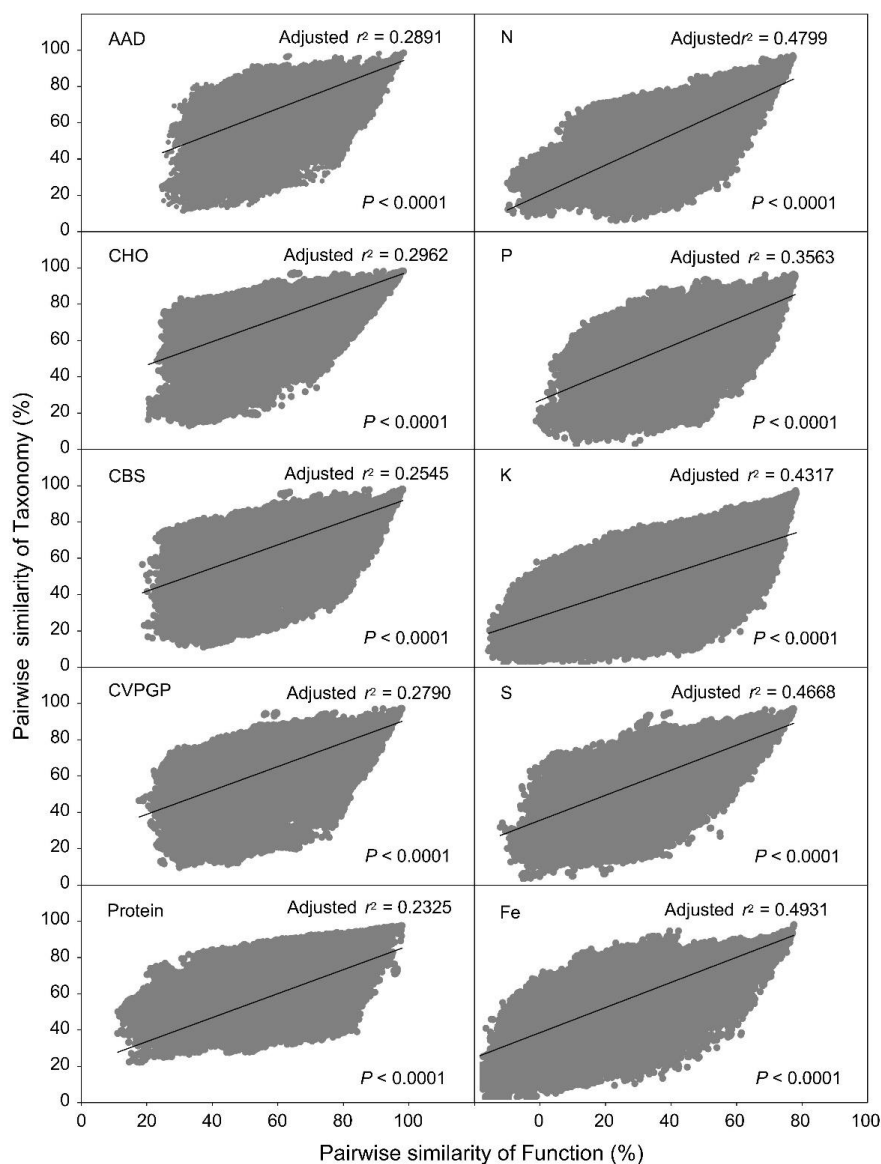
251 (Shannon et al., 2003). The scatter plots of within-module connectivity ( $z_i$ ) and among-  
252 module connectivity ( $P_i$ ) were constructed to show the network node distribution of  
253 module-based topological roles of taxonomic compositions for the “broad” and “narrow”  
254 functions. The threshold values of  $Z_i$  and  $P_i$  for categorizing were 2.5 and 0.62  
255 respectively.

256

### 257 **3. Results and Discussion**

#### 258 **3.1. Microbial taxonomic compositions**

259 This study included 845 soil metagenomes across seventeen climate zones around the  
260 world extracted from 56 MG-RAST studies published in 51 peer-reviewed papers. They  
261 resulted in 356090 pairwise comparisons of Bray-curtis similarity in functional  
262 (Subsystems L3) and taxonomic (RefSeq genus) diversities for the five “broad” and five  
263 “narrow” functions, which were analyzed to find out whether the correlations of function  
264 and taxonomy were greater in the five “narrow” functions. Overall, for the five “narrow”  
265 functions, the positive correlations of the pairwise similarity of taxonomy and function  
266 between either two samples ( $r^2 = 0.36-0.49$ ) were greater than those for the five “broad”  
267 functions ( $r^2 = 0.23-0.29$ ) (Fig. 1). This suggests that rare phylotypes could be more  
268 associated with narrow ecosystem processes than broad-scale functions, supporting the  
269 notion that the abundance of particular specialists could influence narrow functional  
270 measures (Peter et al., 2011; Rivett and Bell, 2018), leading to a lower degree of  
271 functional redundancy associated with “narrow” functions, such as the nutrient cycling  
272 examined in this study.



273

274 **Fig. 1. Relations between functional and taxonomic beta-diversities for “broad” and**  
275 **“narrow” functions.** Pearson’s correlations between pairwise Bray-curtis similarity of  
276 microbial taxonomic and functional compositions for “broad” and “narrow” functions  
277 annotated using Subsystems at function level (Function) and RefSeq at genus level  
278 (Taxonomy). Correlation adjusted  $r$ -squared and  $P$  values are given. “Broad” functions  
279 include AAD (Amino Acids and Derivatives), CHO (Carbohydrates), CBS (Clustering-



280 based Subsystems), CVPGP (Cofactors, Vitamins, Prosthetic Groups, Pigments), and  
281 Protein (Protein Metabolism). “Narrow” functions include N (Nitrogen Metabolism), P  
282 (Phosphorous Metabolism), K (Potassium Metabolism), S (Sulfur Metabolism), and Fe  
283 (Iron Acquisition and Metabolism).

284

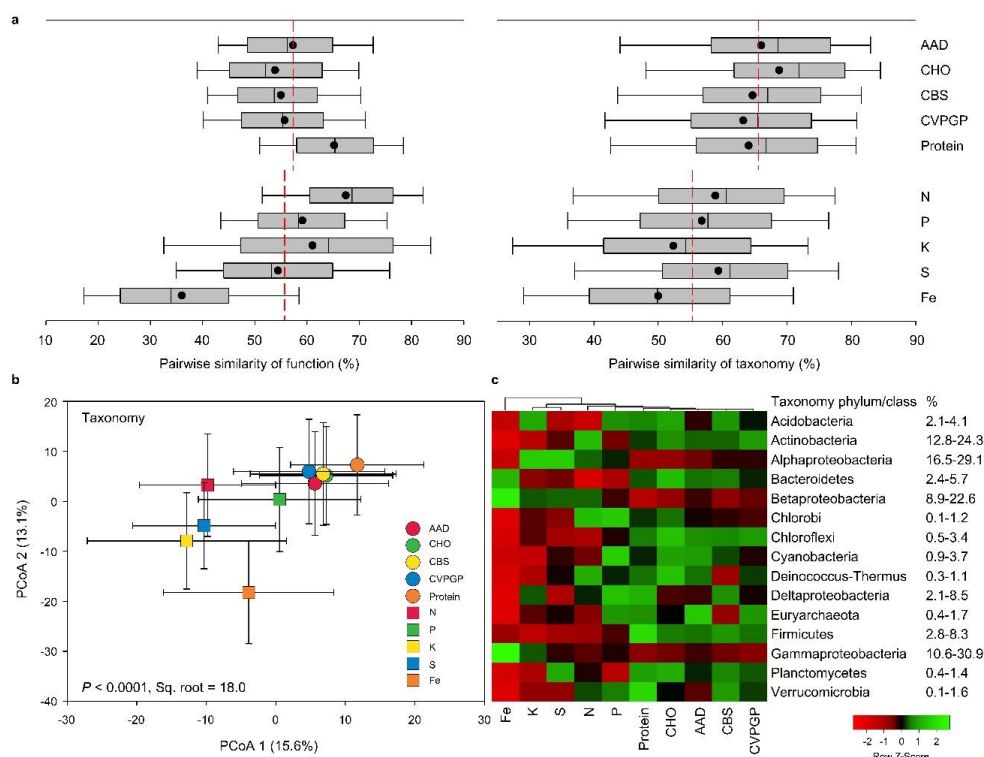
285       Several soil metagenomic studies have reported a linear relationship between  
286 functional and taxonomic diversities (Fierer et al., 2012b;Fierer et al., 2013;Leff et al.,  
287 2015), indicating a somewhat dependency of microbial functional profiles on taxonomic  
288 compositions. This dependency, however, does not necessarily imply an absence of  
289 microbial functional redundancy. In fact, those studies all showed lower variation of beta-  
290 diversity of metagenomic functions than taxonomy (Fierer et al., 2012b;Fierer et al.,  
291 2013;Pan et al., 2014;Souza et al., 2015) or higher similarity in composition of functional  
292 profiles than taxonomic composition (Leff et al., 2015). Those findings support that  
293 microbial functions are relatively more stable than taxonomy responding to ecological  
294 and environmental perturbations. In this study, the five “broad” and the five “narrow”  
295 functions had relative abundance of 5-13% and 0.8-1.4%, respectively. Thus, the five  
296 “broad” functions are more abundant than the five “narrow” functions. In addition, the  
297 numbers of genes within the categories of the five “broad” functions were also greater  
298 than those of the “narrow” functions. As the diversities of the microbes conducting the  
299 five “broad” functions were also greater than those conducting the “narrow” functions,  
300 we calculated the relationship between the diversities of taxonomy and of function, and  
301 compared these relationships between the five “broad” and the five “narrow” functions.  
302 Our study further evidenced a lower extent of functional redundancy in the five “narrow”  
303 functions compared to the five “broad” functions despite the linear correlations found in  
304 our study.



305           The boxplots were constructed based on the pairwise similarity of function and  
306 taxonomy to compare similarity ranges of these two compositions related to the five  
307 “broad” functions versus the five “narrow” functions. For the functional compositions at  
308 specific function gene levels, the average similarity of the five “broad” functional  
309 diversity (58%) was comparable to that of the five “narrow” functions (56%) (Fig. 2a).  
310 However, the pairwise similarity of the five “narrow” functions had larger variation, in  
311 which Fe function had the lowest similarity of 36% and N function had the highest  
312 similarity of 69%. On the contrary, the taxonomic similarity of the five “broad” functions  
313 were consistently greater (63-69%) than those of the five “narrow” functions (50-59%).  
314 The PERMANOVA pairwise test was conducted to find out the difference between  
315 taxonomic similarity of microbes conducting the five “broad” and the five “narrow”  
316 functions based on the relative abundance. Our results indicated that the microbial  
317 taxonomic compositions of the five “broad” functions were more phylogenetically  
318 different from those of the five “narrow” functions (13-22%) than from each other (8-  
319 13%) (Supplementary Table 2). The RELATE test was also conducted to evaluate the  
320 relationship of the taxonomic compositions of microbes conducting the five “broad” and  
321 the five “narrow” functions. Our results confirmed that the microbial taxonomic  
322 compositions of the five “broad” functions were more correlated with each other (0.97-  
323 0.99) than those of the five “narrow” functions (0.77-0.94) (Supplementary Table 3).  
324 When the microbial taxonomic compositions of the ten functional categories were  
325 combined in PCoA analysis, the resulting scatter plot showed that the five “broad”  
326 functions were grouped closely together and separated from the five “narrow” functions  
327 (Fig. 2b). Grouping of the ten functions generally explain up to 18.0% of the community



328 difference, in which the five “narrow” functions were more distinct from each other.  
 329 These evidences together suggest that the taxonomic composition of soil microbes  
 330 conducting the five “broad” functions were more conserved in taxonomy than those  
 331 conducting the five “narrow” functions. However, it should be noted that the current  
 332 analysis had some limitations as the metagenomics datasets consisted of sequencing data  
 333 that are phylogenetically classified and assigned based on certain the taxonomic and  
 334 functional databases. Thus, our results may to some extent depend on the databases  
 335 chosen, of which the classification and assignment may not contain potential bias. Future  
 336 studies should continue to test this hypothesis using regional samples and individual  
 337 datasets.



338





339 **Fig. 2. Functional and taxonomic diversities for “broad” versus “narrow” functions.**

340 **a**, Box plots and mean values of pairwise Bray-curtis similarity of microbial functional  
341 and taxonomic diversities for “broad” versus “narrow” functions. **b**, PCoA (Principal  
342 coordinates analysis) showing beta-diversity of microbial taxonomic diversity for  
343 “broad” and “narrow” functions annotated using RefSeq at genus level (Taxonomy). The  
344 error bars represent the standard deviation of data ranges. Variations (by percentage)  
345 explained by the two principal coordinate dimensions are given in parentheses. *P* values  
346 and sq. root of PERMANOVA are also given. **c**, Heatmaps showing relative abundance  
347 of dominant microbial taxonomic composition (mean > 0.5%) for “broad” and “narrow”  
348 functions annotated using RefSeq at phylum/class levels (Taxonomy). “Broad” functions  
349 include AAD (Amino Acids and Derivatives), CHO (Carbohydrates), CBS (Clustering-  
350 based Subsystems), CVPGP (Cofactors, Vitamins, Prosthetic Groups, Pigments), and  
351 Protein (Protein Metabolism); “Narrow” functions include N (Nitrogen Metabolism), P  
352 (Phosphorous Metabolism), K (Potassium Metabolism), S (Sulfur Metabolism), and Fe  
353 (Iron Acquisition and Metabolism).

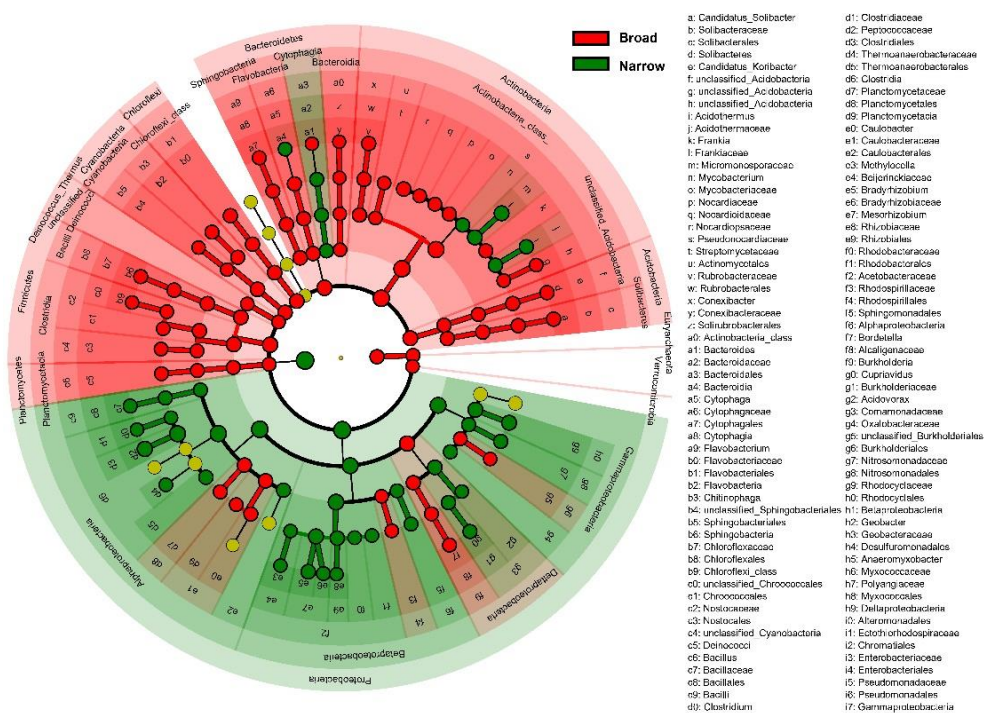
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355 To investigate how microbial taxonomic diversities differ globally, the taxonomic  
356 compositions of soil microbes conducting the five “broad” and the five “narrow”  
357 functions were analyzed among the seventeen climate zones based on the PCoA analysis.  
358 Across climate zones, microbial taxonomic compositions of the five “narrow” functions  
359 (sq. root = 15.2-18.8) were more distinct than the five “broad” functions (sq. root = 13.4-  
360 15.1) based on the PERMANOVA analysis (Supplementary Fig. 1). This suggests that  
361 microorganisms relating to “broad” functions were similar to each other in taxonomy,  
362 because “broad” functions are more broadly distributed across most taxa, but soil  
363 microbes performing “narrow” functions were more phylogenetically diverse due to the  
364 specialty of “narrow” functions. Thus, though microbial metabolic functions can be  
365 strongly coupled to elemental cycles and certain environmental factors, the decoupling of



366 microbial taxonomic and functional profiles is still inevitable when a low-dimensional  
 367 functional space is projected to a high-dimensional taxonomic space (Louca et al., 2018),  
 368 especially for “broad” functions.

369 The taxonomic compositions of microbes conducting the five “broad” functions were  
 370 more abundant in most major phyla, such as Acidobacteria, Actinobacteria,  
 371 Bacteroidetes, and Firmicutes, while the relative abundance of the taxonomic  
 372 composition of microbes conducting the five “narrow” functions were greater in  
 373 Proteobacteria, especially Alphaproteobacteria and Betaproteobacteria (Fig. 2c). Other  
 374 studies also found that some bacteria conducting N cycling, such as ammonia-oxidizers  
 375 and rhizobia for N fixation, mainly belong to Alphaproteobacteria or Betaproteobacteria  
 376 (Stephen et al., 1996; Moulin et al., 2001).



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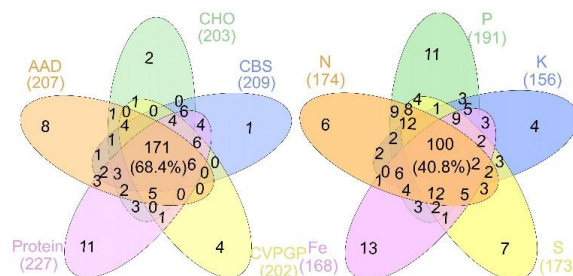
378 **Fig. 3. Taxonomic compositions shared among “broad” and “narrow” functions.**  
379 Venn’s diagrams showing dominant microbial taxonomic groups (mean > 0.1%)  
380 annotated using RefSeq at genus levels (Taxonomy) shared among “broad” and “narrow”  
381 functions.

382

383 LefSe analysis was used to show the dominant microbial groups at the taxonomic  
384 levels of domain, phylum, class, order, family, and genus that were statistically different  
385 between the five “broad” and the five “narrow” functions based on their relative  
386 abundances. In particular, among the Proteobacteria conducting the five “narrow”  
387 functions, *Bacillaceae* from Bacilli, *Clostridium*, *Peptococcaceae*, and  
388 *Thermoanaerobacteraceae* from Clostridia, *Methylocella*, *Bradyrhizobium*,  
389 *Bradyrhizobiaceae*, and *Rhizobiaceae* from Rhodospirillaceae, and *Cupriavidus* from  
390 Comamonadaceae had higher relative abundance than the others (Fig. 3). The Venn’s  
391 diagrams indicated that the taxonomic compositions of soil microbes performing the  
392 “broad” functions shared 68% dominant genera among the five functional categories,  
393 while the proportion was reduced to only 41% for the five “narrow” functions (Fig. 4).  
394 However, it should be stated that all the analyses performed in our study were based on  
395 relative abundance data that is compositional, so it is difficult to directly compare  
396 taxonomic diversities among samples and/or datasets. Despite the differences in the  
397 identification protocol and quantification of soil metagenomes, we deem the effects of  
398 these differences to be trivial for our analyses as we intended to understand the general  
399 patterns of microbial taxonomic and functional linkages, rather than simply compare soil  
400 community structures across samples. By uncovering universal patterns of these  
401 relationships within the microbial community, we can then further establish a potential



402 linkage framework to account for the microbial contributions to major biogeochemical  
403 cycles.



404

405 **Fig. 4. Difference of taxonomic compositions between “broad” and “narrow**  
406 **functions”.** LEfSe (linear discriminant analysis effect size) results showing the  
407 significant differences in the relative abundance of dominant microbial taxonomic groups  
408 (mean > 0.5%) between “broad” (red) versus “narrow” (green) functions annotated using  
409 RefSeq (Taxonomy). From the center outward, each circle represents the level of domain,  
410 phylum, class, order, family, and genus, respectively. The taxonomic groups with  
411 significant differences are labeled by colors.

412

413 Because of functional redundancy of soil microbes, understanding what types of  
414 functions that have more significant association with microbial taxonomy can be critical  
415 for accurate prediction of microbial metabolic activity and flexibility across space and  
416 time. As microbial taxonomic composition and diversity plays critical role in maintaining  
417 ecosystem function (Allison and Martiny, 2008), our results suggest that taxonomic  
418 information alone provides limited utility in predicting basic metabolic capabilities, but  
419 may be capable of forecasting biogeochemical transformations or changes in the rate of  
420 biogeochemical process at ecosystem level (Hall et al., 2018). Investigating functional  
421 redundancy with respect to functions associated with elemental cycles provides useful  
422 information for guiding the development of explicit microbial biogeochemical prediction,



423 and further delving into major pathways of C and N cycles will be a fruitful approach for  
424 scrutinizing microbes' functional potentials.

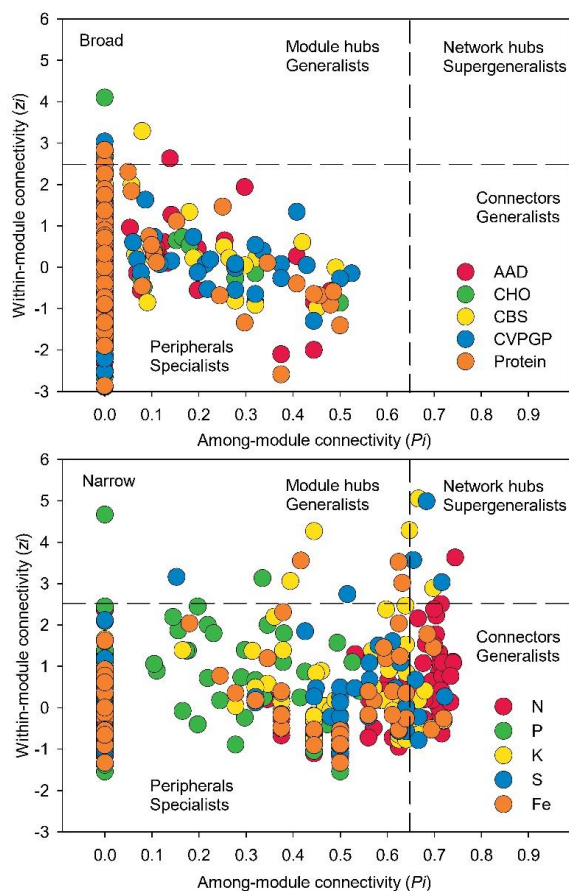
425 **Table 1. Summary of key properties of co-occurrence networks for the five “broad”**  
426 **and the five “narrow” functions.**

Network Indexes	Total nodes	Total links (positive%)	Average connectivity	Average clustering coefficient	Average geodesic distance	Modularity (modules numbers)
		1472				
AAD	225	(100%)	13.084	0.663	2.873	0.695 (11)
CHO	207	1155 (99%)	11.159	0.615	3.805	0.672 (10)
CBS	246	1622 (99%)	13.187	0.663	2.859	0.671 (11)
CVPGP	201	1293 (99%)	12.866	0.65	3.303	0.697 (9)
Protein	285	1651 (99%)	11.586	0.638	2.992	0.749 (14)
N	101	519 (12%)	10.277	0.349	1.903	0.184 (5)
P	160	449 (4%)	5.612	0.299	3.298	0.615 (10)
K	143	364 (67%)	5.091	0.08	2.676	0.429 (6)
S	132	264 (15%)	4	0.09	2.563	0.486 (12)
Fe	95	215 (11%)	4.526	0.071	2.601	0.435 (6)

427

### 428 3.2. Microbial taxonomic co-occurrence networks

429 Co-occurrence networks of taxonomic compositions were generated to identify potential  
430 interaction patterns of microbial groups that conduct the five “broad” and the five  
431 “narrow” functions across the globe. Network graphs with submodule structures  
432 indicated distinct topology of taxonomic networks between the “broad” and “narrow”  
433 functions (Table 1, Supplementary Fig. 2 and Supplementary Fig. 3). Compared to the  
434 “narrow” functions, the “broad” functions harbored larger and more complex networks  
435 with more nodes (201-285 vs. 95-160) and links (1293-1651 vs. 215-519), with higher  
436 average connectivity (11.2-13.2 vs. 4.0-10.3) and average clustering coefficient (0.64-  
437 0.66 vs. 0.07-0.35). The “broad” function network had 99-100% positive links, while the  
438 “narrow” function had 33-96% negative links.



439

440 **Fig. 5. Network information of taxonomic compositions for “broad” and “narrow”**

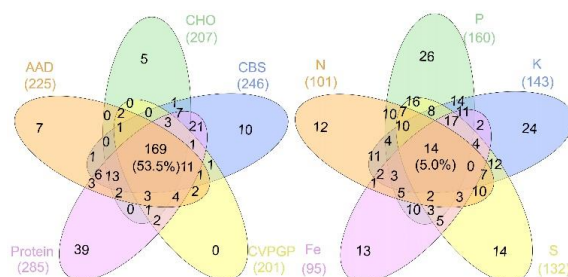
441 **functions.** Node distribution of module-based topological roles of taxonomic  
442 compositions for “broad” and “narrow” functions determined by the scatter plot of  
443 within-module connectivity ( $z_i$ ) and among-module connectivity ( $P_i$ ). The threshold  
444 values of  $Z_i$  and  $P_i$  for categorizing were 2.5 and 0.62 respectively.

445

446 In addition, network modularity was greater in the “broad” functions, indicating that  
447 significant correlations between taxonomic compositions of microbes that conduct the  
448 five “broad” functions are mainly within similar taxonomic groups. No node could be  
449 classified as connectors in the five “broad” function networks (Fig. 5), reaffirming that the



450 “broad” function networks had links mainly within modules of similar species. In the co-  
451 occurrence network of taxonomic composition of the “narrow” functions, 13% of the  
452 nodes were identified as connectors linking several modules (high  $P_i$ ) connectors, while  
453 3% were identified as module hubs that connected other nodes within their own modules  
454 (high  $Z_i$ ), indicated by the  $Z_i$ - $P_i$  plot (Olesen et al., 2007;Deng et al., 2012). Thus,  
455 significantly less nodes were identified as module hubs in the co-occurrence network of  
456 the taxonomic composition of the “broad” functions, indicating less correlations found  
457 among different modules. This is expected given that module was comprised of genera  
458 that were mainly from the same phylogenetic groups. This difference was consistent with  
459 the Venn’s diagrams showing significantly more nodes (54%) shared among the five  
460 functional categories representing the “broad” functions, while only 5% of the nodes  
461 were overlaid among the five “narrow” function networks (Fig. 6). Environmental  
462 conditions likely determine the microbial taxonomic composition, and microbial  
463 phylotypes sharing similar habitat preferences tend to co-occur (Delgado-Baquerizo et  
464 al., 2018;Ram íez-Flandes et al., 2019). We emphasize that this analysis is a combination  
465 of snapshots of microbial communities compared across space, thus environmental  
466 conditions (at the same geographic location) may vary, and the levels of functional  
467 redundancy may change depending on the mechanisms selecting specific functions and  
468 the phylogenetic distribution of those functions (Louca et al., 2018).



469



470 **Fig. 6. Taxonomic network nodes shared among “broad” and “narrow” functions.**  
471 Venn’s diagrams showing the microbial taxonomic network nodes shared among “broad”  
472 and “narrow” functions.

473

### 474 **3.3. Conclusion**

475 By analyzing and generalizing microbial taxonomic and functional profiles, we provide  
476 strong evidence that the degree of soil microbial functional redundancy differ  
477 significantly between “broad” and “narrow” functions across the global. The level of  
478 functional redundancy varies depending on the functions of interest. Here, by contrasting  
479 the five “broad” metabolic functions and the five “narrow” functions that are important  
480 for elemental cycles, we found lower levels of functional redundancy associated with the  
481 five “narrow” functions of biogeochemical cycling, despite the fact that even for the five  
482 “narrow” functions, there is still a high level of functional redundancy in the soil  
483 communities. Although there is a caveat concerning direct comparison of metagenomic  
484 data, the present study demonstrated the use of comparative metagenome and co-  
485 occurrence network analysis in generalizing patterns of microbial characteristics  
486 regulating biogeochemical cycling of major elements. With the increasing advancement  
487 of sequencing techniques and data coverage, future sequencing efforts will likely increase  
488 our confidence in comparative metagenomes and provide time-series information to  
489 further identify to what extent microbial functional redundancy regulates dynamic  
490 ecological fluxes across space and time.

491

### 492 **Author Contributions**





493 Huaihai Chen conceived the study, performed the data analysis, interpreted the results,  
494 and drafted the manuscript. CL, YY, CWS, and Hao Chen secured the research funding.  
495 KM, YH, YY, and Hao Chen critically assessed and interpreted the findings. All authors  
496 discussed results, commented on, edited, revised, and approved the manuscript.

497

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505

#### 506 **Data Availability Statement**

507 The data that support the findings of this study are available from the corresponding  
508 author upon request. All metagenomic data used in this study are publicly assessable in  
509 the MG-RAST server with study and MG-RAST ID reported in supplementary files.

510

#### 511 **Competing Interests**

512 The authors declare no competing interests.

513

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