

1 **Lower functional redundancy in “narrow” than “broad” functions in global soil**
2 **metagenomics**

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Abstract

Understanding the relationship between soil microbial taxonomic compositions and functional profiles is essential for predicting ecosystem functions under various environmental disturbances. However, even though microbial communities are sensitive to disturbance, ecosystem functions remain relatively stable, as soil microbes are likely to be functionally redundant. Microbial functional redundancy may be more associated with “broad” functions carried out by a wide range of microbes, than with “narrow” functions specialized by specific microorganisms. Thus, a comprehensive study to evaluate how microbial taxonomic compositions correlate with “broad” and “narrow” functional profiles is necessary. Here, we evaluated soil metagenomes worldwide to assess whether functional and taxonomic diversities differ significantly between the five “broad” and the five “narrow” functions that we chose. Our results revealed that compared with the five “broad” functions, soil microbes capable of performing the five “narrow” functions were more taxonomically diverse, and thus their functional diversity was more dependent on taxonomic diversity, implying lower levels of functional redundancy in “narrow” functions. Co-occurrence networks indicated that microorganisms conducting “broad” functions were positively related, but microbes specializing “narrow” functions were interacting mostly negatively. Our study provides strong evidence to support our hypothesis that functional redundancy is significantly different between “broad” and “narrow” functions in soil microbes, as the association of functional diversity with taxonomy were greater in the five “narrow” rather than the five “broad” functions.

Keywords Functional redundancy, Soil metagenomics, Functional traits, Taxonomic compositions,

1. Introduction

Microbial communities often exhibit incredible taxonomic diversity, with one gram of soil harboring millions of microbial species (Gans et al., 2005). However, how such diversity governs microbial functional potential and ecosystem processes is largely unknown. Though microbial taxonomic composition is generally sensitive to disturbance and often **does** not rapidly recover (Allison and Martiny, 2008), it is unclear how changes in microbial community composition would regulate ecosystem functioning. Mechanistic understanding of microbial systems, including microbial taxonomic compositions and functional potential, is essential for predicting ecosystem functioning under various environmental disturbances (Torsvik and Øvreås, 2002; Wellington et al., 2003; McGill et al., 2006).

Though microbial community composition usually **shifts** in response to disturbance, ecosystem functions could remain relatively stable due to functional redundancy (Allison and Martiny, 2008). Microbial functional redundancy is an inevitable emergent property of microbial systems (Louca et al., 2018), as some metabolic functions can be performed by multiple species, which may thus be substitutable in certain ecosystem processes (Rosenfeld, 2002), implying that microbial taxonomy and function can be decoupled (Louca et al., 2016; Louca et al., 2017). **The concept of functional redundancy can be “strict redundancy” meaning that microorganisms sharing the exact same set of functions can easily substitute each other, or alternatively “partial redundancy” denoting that**

microbes have similarity in certain functions but still harbor difference in other functions, leading to partially dissimilar ecological requirements or environmental preference (Galand et al., 2018). In addition, microbial functional redundancy may be caused by more than just metabolic processes but other mechanical response to environmental disturbance, such as different foraging strategies, particles attachment and biofilm formation, nitrogen source usage, and resistance to antibiotics, which are difficult to be thoroughly evaluated in the current approach mostly focusing on metabolic redundancy (Louca et al., 2018).

Microbial functional redundancy has been mainly observed in “broad” ecosystem processes (Yin et al., 2000; Rousk et al., 2009; Banerjee et al., 2016), but is perhaps less significant in “narrow” functions specialized by certain microorganisms (Schimel, 1995; Balser et al., 2002). However, some studies simulating microbial diversity reduction and physiological processes challenged the hypothesis of microbial redundancy in soil microbes (Peter et al., 2011; Philippot et al., 2013; Delgado-Baquerizo et al., 2016).

Microbial functional redundancy is inevitable when a high-dimensional trait space is projected to a lower-dimensional function space of interest (Louca et al., 2018). Such apparent contradictory results suggest the degree of functional redundancy may arise from the definition of “redundancy” in different studies, our limitations in measuring the factors controlling niche space, and more importantly depending on the function of interest. Microbes conducting “broad” metabolic functions, such as carbon decomposition, are likely to distribute across most taxa (Crowther et al., 2019) and associate with high level of functional redundancy (Beier et al., 2017; Rivett and Bell, 2018). “Narrow” functions, such as nitrification or methanogenesis, may be restricted to a

few phylogenetic clades (Schimel and Gullledge, 1998), and are hypothesized to exhibit less redundancy than “broad” functions (Schimel, 1995; Rocca et al., 2015). Today, multifunctionality (Hector and Bagchi, 2007) has to be accounted for to avoid overestimating functional redundancy (Gamfeldt et al., 2008). By assessing multiple functions, the relationship between microbial diversity and ecosystem function can be better quantified in the soil (Bastida et al., 2016; Delgado-Baquerizo et al., 2016).

Nowadays, metagenomics have been increasingly used as a promising comparative tool (Tringe et al., 2005) to study the relationship between functional and taxonomic diversities (Fierer et al., 2012a; Fierer et al., 2012b; Fierer et al., 2013; Pan et al., 2014; Leff et al., 2015; Souza et al., 2015). The growing wealth of soil metagenome data thus poised well to aid in the generalization of global patterns of microbial attributes and standardizing frameworks for consistent representation of microbial community (Chen et al., 2021; Xu et al., 2021). However, a synthetic metagenomic analysis to assess how general microbial taxonomic and functional diversities differ between “broad” and “narrow” functions across the globe is still lacking.

Here, we constructed soil metagenomic datasets of taxonomic and functional diversities of five “broad” and five “narrow” functions across seventeen climate zones. We typically chose SEED Subsystems database (Overbeek et al., 2013) that has diverse classification at level 1, allowing us to conduct comparison between “broad” versus “narrow” functions. We selected five “narrow” functions, namely N (Nitrogen Metabolism), P (Phosphorous Metabolism), K (Potassium Metabolism), S (Sulfur Metabolism), and Fe (Iron Acquisition and Metabolism). These are typical functional categories of specific nutrient cycling in Subsystems Level 1 and are only performed by

certain groups of soil microbes (Schimel, 1995). The five “broad” functions selected were AAD (Amino Acids and Derivatives), CHO (Carbohydrates), CBS (Clustering-based Subsystems), CVPGP (Cofactors, Vitamins, Prosthetic Groups, Pigments), and Protein (Protein Metabolism), which are the most abundant functional categories in Subsystems level 1, and represent broad-scale functions acquired by a relatively larger group of diverse soil microbes (Balser et al., 2002). We further constructed the pairwise similarity of function and taxonomy based on the relative abundance of functional and taxonomic compositions, respectively, for the five “broad” and the five “narrow” functions. We hypothesized that the taxonomic similarity of soil microbes would be more linearly correlated to the functional similarity for the five “narrow” functions in comparison to the five “broad” functions. Therefore, using these global soil metagenomes, our objective was to test whether the taxonomic compositions of soil microbes that conduct the five “narrow” functions are more dependent on the functional compositions, leading to a lower level of functional redundancy in the “narrow” functions than the “broad” functions.

2. Materials and Methods

2.1. Data collection

To ensure that the quality and completeness of the metagenomes analyzed were of standard, we carefully selected soil metagenomes in MG-RAST server that have been published in peer-reviewed journals. We searched peer-reviewed publications from 2012 to 2018 from the Web of Science database using search terms such as “soil metagenome”, “shotgun sequencing”, and “MG-RAST” to source the metagenomic data

used in this study to their publications. We included soil metagenomes publicly available in the MG-RAST database that are generated using shotgun sequencing without amplification or that were directly deposited by peer-reviewed studies into the MG-RAST database. We then extracted data matrix of taxonomic and functional compositions of soil metagenomes from MG-RAST public server (<https://www.mg-rast.org/>) based on the Study ID and/or MG-RAST ID reported in the publications. Details of each soil metagenome extracted from publications and MG-RAST database was given in Supplementary Table S1.

The functional database that we used in this study, SEED Subsystems, is a categorization system which organizes gene functional categories into a hierarchy with three levels of resolution (Level 3, 2 and 1) (Overbeek et al., 2013). To download the taxonomic compositions to soil microbes to conduct “broad” and “narrow” functions, for each soil metagenome, in the ‘Analysis’ function of the MG-RAST server (<https://www.mg-rast.org/mgmain.html?mgpage=analysis>), we loaded both SEED Subsystems (Level 3, 2 and 1) as functional profiles and RefSeq (Tatusova et al., 2013) databases (genus, family, order, class, and phylum levels) as taxonomic compositions (Chen et al., 2021). The detailed protocols of MG-RAST server were followed to analyze the metagenomic functions (Meyer et al., 2008; Wilke et al., 2017). To obtain the taxonomic compositions of soil microbes that conduct the selected “broad” and “narrow” functions, we chose ‘RefSeq’ as source and ‘genus’ as level, and in ‘function filter’ we added the functional categories in Subsystems Level 1 that we are interested in, including five “broad” functions of AAD (Amino Acids and Derivatives), CHO (Carbohydrates), CBS (Clustering-based Subsystems), CVPGP (Cofactors, Vitamins, Prosthetic Groups,

Pigments), and Protein (Protein Metabolism), of which the relative abundance was 5-13%. The functions of AAD, CHO, CBS, CVPGP, and Protein were the most abundant functional categories in Subsystems Level 1, which were used to represent broad-scale functions acquired by a large group of diverse soil microbes. Correspondingly, five “narrow” functions were chosen, namely N (Nitrogen Metabolism), P (Phosphorous Metabolism), K (Potassium Metabolism), S (Sulfur Metabolism), and Fe (Iron Acquisition and Metabolism), of which the relative abundance was 0.8-1.4%, as these are typical functional categories of specific nutrient cycling in Subsystems Level 1 and are only performed by certain groups of soil microbes. The genus level was used as the taxonomic classification level across different datasets. Following default setting in MG-RAST, if the species were classified into the higher classification levels than genus but failed to be identified at the genus level, they were classified into “unclassified” groups. Across different studies, there were 2.16 ± 0.85 % of sequences belonging to the “unclassified” groups, showing that most taxonomic groups could be classified into the genus level. Total hits of taxonomic compositions of soil microbes conducting each function at Subsystems Level 1 were calculated as the sums of hits in different taxonomic categories at RefSeq genus level.

The comparative metagenomic analyses were performed using default settings (maximum e-value cutoff = $1e^{-5}$, minimum identity cutoff = 60%, and minimum alignment length = 50) (Meyer et al., 2008). We then merged the taxonomic compositions of data matrix of each functions extracted from different studies together to generate new datasets of microbial taxonomic compositions annotated by the RefSeq database. The reason why we chose the Subsystems database for functional grouping rather than KEGG

Orthology (KO) (Kanehisa et al., 2015), Clusters of Orthologous Groups (COG) (Galperin et al., 2014), and Non-supervised Orthologous Groups (NOG) (Huerta-Cepas et al., 2015) databases was that Subsystems had more diverse classification at Level 1, allowing us to conduct direct comparison between “broad” versus “narrow” functions. We chose RefSeq database rather than the traditional ribosomal RNA databases, such as RDP (Ribosomal Database Project) (Cole et al., 2008), Greengenes (DeSantis et al., 2006), or Silva LSU/SSU (Pruesse et al., 2007) databases, because taxonomic hits in the RefSeq database were over 1000-fold higher than the rRNA databases, rendering the resolution comparable to functional hits for comparison between “broad” and “narrow” functions. To increase the coverage of our datasets, soil metagenomes with/without assembly were both included.

The geographic coordinates of latitudes (LAT) and longitudes (LONG) of each soil metagenome were directly obtained from publications. Based on LAT and LONG, climate data of mean annual temperature (MAT) and precipitation (MAP) of study sites for each soil metagenome were extracted from the WorldClim dataset (Fick and Hijmans, 2017) using the R package ‘raster’ (Hijmans et al., 2015). To examine how microbial taxonomic diversities of “broad” and “narrow” functions differ globally, soil metagenomic data was classified into seventeen climate zones based on the main classification of Koeppen-Geiger Climatic Zones (Kottek et al., 2006) using the R package ‘kgc’ (Bryant et al., 2017).

2.2. Statistical Analyses

206 To minimize bias caused by different sequencing depths and read lengths among studies,
 207 we standardize the hits of each taxonomic (or functional) category in each data to relative
 208 abundance by dividing them by the total number of hits. To calculate the pairwise
 209 similarity of taxonomy based on the relative taxonomic abundance at genus level of
 210 microbes conducting the five “broad” and five “narrow” functions, we calculated Bray-
 211 Curtis similarity following log transformation of the compositional taxonomic data by
 212 constructing pairwise Bray-Curtis similarity matrix between each pair of two samples for
 213 each functional categories at Subsystems database at Level 1, which were further
 214 transformed to lists of pairwise Bray-Curtis similarities ordered by sample pair names in
 215 PRIMER 7 (Plymouth Routines in Multivariate Ecological Research Statistical Software,
 216 v7.0.13, PRIMER-E Ltd, UK) (Clarke and Gorley, 2015). To calculate the pairwise
 217 similarity of function, based on the functional abundance at function gene level within
 218 each of the five “broad” and five “narrow” functions, we calculated Bray-Curtis
 219 similarity following log transformation of the compositional functional data by
 220 constructing pairwise Bray-Curtis similarity matrix between each pair of two samples for
 221 each functional categories at Subsystems database at Level 1, which were further
 222 transformed to lists of pairwise Bray-Curtis similarities ordered by sample pair names in
 223 PRIMER 7. To examine the relationship between functional and taxonomic diversities,
 224 Pearson’s correlations were constructed between the transformed lists of pairwise Bray-
 225 Curtis similarity of soil metagenomes annotated using Subsystems database at Level 3
 226 (Function) and the RefSeq database at genus level (Taxonomy). The approaches for
 227 processing the relative abundance of compositional data follow the requirements (Gloor
 228 et al., 2017). To analyze the taxonomic composition structures of soil metagenomes

annotated using the RefSeq database at genus level (Taxonomy) of the five “broad” and five “narrow” functions, PCoA (principal coordinates analysis) and PERMANOVA (Permutational multivariate analysis of variance) were conducted using the pairwise Bray-Curtis similarity matrix in PRIMER 7.

To compare microbial taxonomic compositions among the five “broad” and the five “narrow” functions, one-factor PERMANOVA was conducted using the main test and pair-wise test in PRIMER 7 with *P* values and Sq. root reported. Pearson’s correlations were constructed to assess the relationships between functional and taxonomic diversities in the “broad” and “narrow” functions with adjusted P-Square reported. A RELATE analysis was also performed to evaluate the relatedness among “broad” and “narrow” functions by calculating a Spearman’s Rho correlation coefficient in PRIMER 7. To examine the relative abundance of dominant microbial at phylum and class level (mean > 1%) among the five “broad” and five “narrow” functions, heatmaps were constructed using HeatMapper (Babicki et al., 2016). One-way analysis of variance (ANOVA) with *P* values adjusted by Bonferroni-correction for multiple comparisons was conducted using SPSS 22.0 software (Chicago, IL, USA) to evaluate the differences in the relative abundance of dominant taxonomic compositions (mean > 1%) among climate zones after the normality of residues and homogeneity of variance were checked using Shapiro-Wilk and Levene test, respectively. The significance level was set at $\alpha=0.05$ unless otherwise stated. To calculate the statistical difference between the relative abundance of dominant microbial taxonomic groups (mean > 1%) in the “broad” and “narrow” functions, LEfSe (linear discriminant analysis effect size) method was used (<http://huttenhower.sph.harvard.edu/lefse/>) (Segata et al., 2011). Venn’s diagrams were

constructed to visualize the amount of dominant microbial taxonomic groups at genus levels or network nodes shared between the five “broad” and the five “narrow” functions using InteractiVenn (Heberle et al., 2015).

To find out potential interactions of microbial taxonomic compositions between “broad” and “narrow” functions across the globe, co-occurrence network analysis was performed using the Molecular Ecological Network Analyses Pipeline (<http://ieg4.rccc.ou.edu/MENA/>) (Zhou et al., 2011; Deng et al., 2012). To make the minimum observed value close to but no less than 1 as required by the pipeline, the data of relative abundance were multiplied by 10^6 , which would not change the correlation coefficients. The data matrix of transformed data matrix was uploaded to construct the network with default settings, including (1) keeping only the species present in more than a half of all samples; (2) only filling with 0.01 in blanks with paired valid values; (3) taking logarithm with recommended similarity matrix of Pearson’s correlation coefficient; and (4) calculation ordered to decrease the cutoff from top using regress poisson distribution only. A default cutoff value (similarity threshold, S_r) for the similarity matrix was used to assign a link between the pair of species. After that, the global network properties, the individual nodes' centrality, and the module separation and modularity were analyzed based on default settings using greedy modularity optimization. Network files were exported and visualized using Cytoscape software (Shannon et al., 2003). The scatter plots of within-module connectivity (z_i) and among-module connectivity (P_i) were constructed to show the network node distribution of module-based topological roles of taxonomic compositions for the “broad” and “narrow” functions. The threshold values of Z_i and P_i for categorizing were 2.5 and 0.62

respectively (Guimerà and Nunes Amaral, 2005; Olesen et al., 2006; Guimerà et al., 2007).

An overview of data acquisition, transformation, and analysis processes in this study was given in Supplementary Fig. 1.

3. Results and Discussion

3.1. Microbial taxonomic compositions

This study included 845 soil metagenomes across seventeen climate zones around the world extracted from 56 MG-RAST studies published in 51 peer-reviewed papers. They resulted in 356090 pairwise comparisons of Bray-curtis similarity in functional (Subsystems L3) and taxonomic (RefSeq genus) diversities for the five “broad” and five “narrow” functions, which were analyzed to find out whether the correlations of function and taxonomy were greater in the five “narrow” functions. Overall, for the five “narrow” functions, the positive correlations of the pairwise similarity of taxonomy and function between either two samples ($r^2 = 0.36-0.49$) were greater than those for the five “broad” functions ($r^2 = 0.23-0.29$) (Fig. 1). This suggests that rare phylotypes could be more associated with narrow ecosystem processes than broad-scale functions, supporting the notion that the abundance of particular specialists could influence narrow functional measures (Peter et al., 2011; Rivett and Bell, 2018), leading to a lower degree of functional redundancy associated with “narrow” functions, such as the nutrient cycling examined in this study.

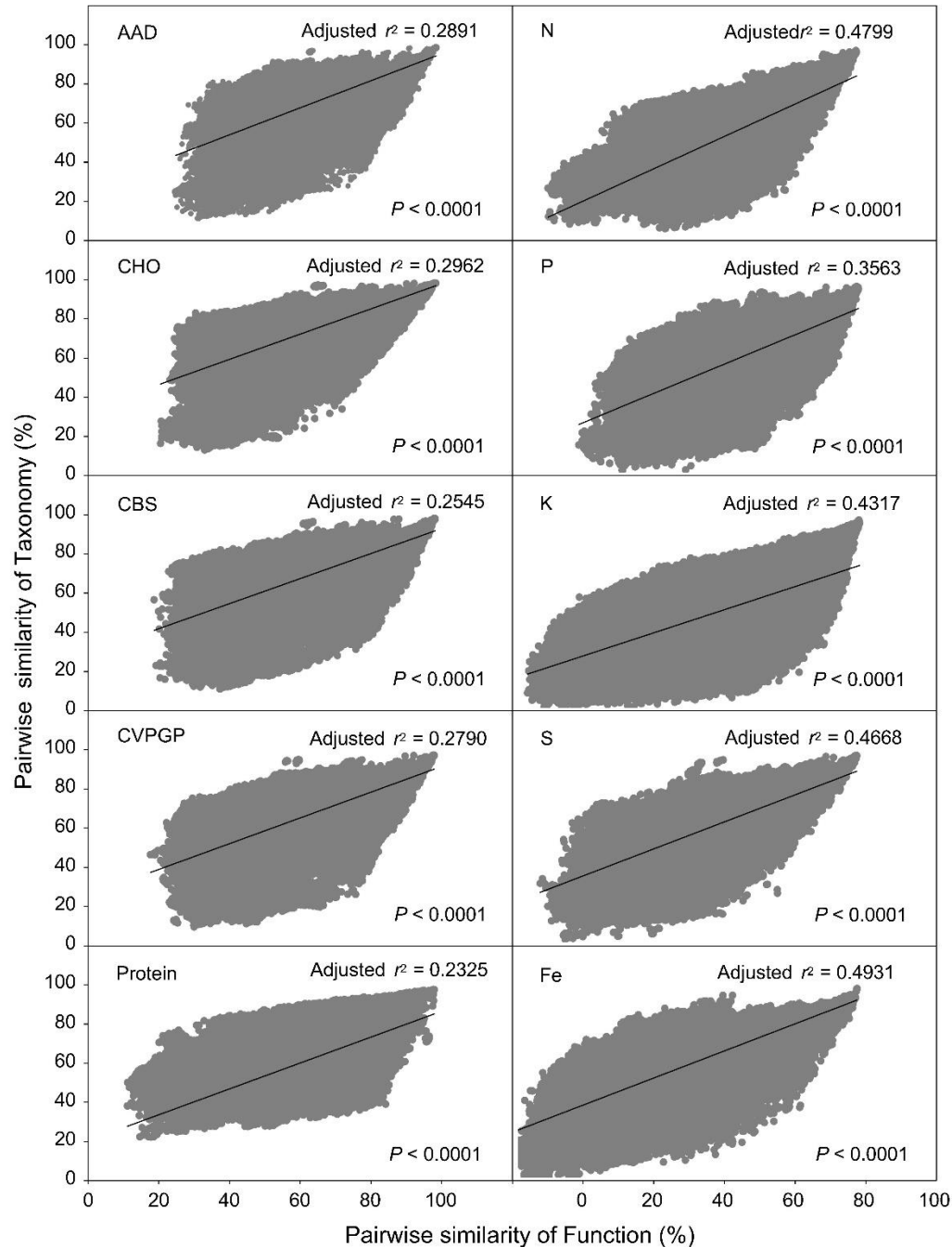


Fig. 1. Relations between functional and taxonomic beta-diversities for “broad” and “narrow” functions. Pearson’s correlations between pairwise Bray-curtis similarity of microbial taxonomic and functional compositions for “broad” and “narrow” functions annotated using Subsystems at function level (Function) and RefSeq at genus level (Taxonomy). Correlation adjusted r -squared and P values are given. “Broad” functions include AAD (Amino Acids and Derivatives), CHO (Carbohydrates), CBS (Clustering-

based Subsystems), CVPGP (Cofactors, Vitamins, Prosthetic Groups, Pigments), and Protein (Protein Metabolism). “Narrow” functions include N (Nitrogen Metabolism), P (Phosphorous Metabolism), K (Potassium Metabolism), S (Sulfur Metabolism), and Fe (Iron Acquisition and Metabolism).

Several soil metagenomic studies have reported a linear relationship between functional and taxonomic diversities (Fierer et al., 2012b;Fierer et al., 2013;Leff et al., 2015), indicating a somewhat dependency of microbial functional profiles on taxonomic compositions. This dependency, however, does not necessarily imply an absence of microbial functional redundancy. In fact, those studies all showed lower variation of beta-diversity of metagenomic functions than taxonomy (Fierer et al., 2012b;Fierer et al., 2013;Pan et al., 2014;Souza et al., 2015) or higher similarity in composition of functional profiles than taxonomic composition (Leff et al., 2015). Those findings support that microbial functions are relatively more stable than taxonomy responding to ecological and environmental perturbations. In this study, the five “broad” and the five “narrow” functions had relative abundance of 5-13% and 0.8-1.4%, respectively. Thus, the five “broad” functions are more abundant than the five “narrow” functions. In addition, the numbers of genes within the categories of the five “broad” functions were also greater than those of the “narrow” functions. As the diversities of the microbes conducting the five “broad” functions were also greater than those conducting the “narrow” functions, we calculated the relationship between the diversities of taxonomy and of function, and compared these relationships between the five “broad” and the five “narrow” functions. Our study further evidenced a lower extent of functional redundancy in the five “narrow” functions compared to the five “broad” functions despite the linear correlations found in our study.

To compare similarity ranges of these two compositions related to the five “broad” functions versus the five “narrow” functions, the boxplots were constructed based on the pairwise similarity of function and taxonomy. For the functional compositions at specific function gene levels, the average similarity of the five “broad” functional diversity (58%) was comparable to that of the five “narrow” functions (56%) (Fig. 2a). However, the pairwise similarity of the five “narrow” functions had larger variation, in which Fe function had the lowest similarity of 36% and N function had the highest similarity of 69%. On the contrary, the taxonomic similarity of the five “broad” functions were consistently greater (63-69%) than those of the five “narrow” functions (50-59%). The PERMANOVA pairwise test was conducted to find out the difference between taxonomic similarity of microbes conducting the five “broad” and the five “narrow” functions based on the relative abundance. Our results indicated that the microbial taxonomic compositions of the five “broad” functions were more phylogenetically different from those of the five “narrow” functions (13-22%) than from each other (8-13%) (Supplementary Table 2). The RELATE test was also conducted to evaluate the relationship of the taxonomic compositions of microbes conducting the five “broad” and the five “narrow” functions. Our results confirmed that the microbial taxonomic compositions of the five “broad” functions were more correlated with each other (0.97-0.99) than those of the five “narrow” functions (0.77-0.94) (Supplementary Table 3). When the microbial taxonomic compositions of the ten functional categories were combined in PCoA analysis, the resulting scatter plot showed that the five “broad” functions were grouped closely together and separated from the five “narrow” functions (Fig. 2b). Grouping of the ten functions generally explain up to 18.0% of the community

difference, in which the five “narrow” functions were more distinct from each other.
 These evidences together suggest that the taxonomic composition of soil microbes
 conducting the five “broad” functions were more conserved in taxonomy than those
 conducting the five “narrow” functions. However, it should be noted that the current
 analysis had some limitations as the metagenomics datasets consisted of sequencing data
 that are phylogenetically classified and assigned based on **certain** taxonomic and
 functional databases. Thus, our results may to some extent depend on the databases
 chosen, of which the classification and assignment **may** contain potential bias. Future
 studies should continue to test this hypothesis using regional samples and individual
 datasets.

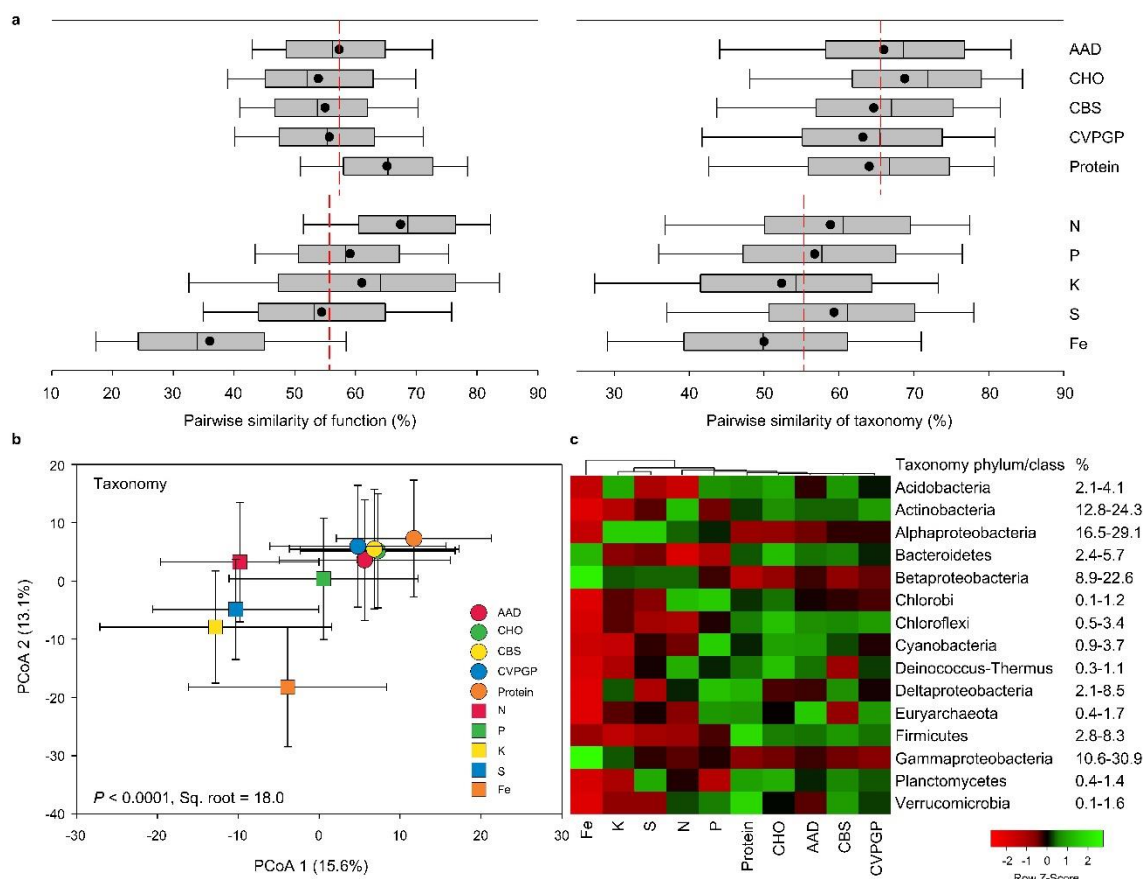


Fig. 2. Functional and taxonomic diversities for “broad” versus “narrow” functions.

a, Box plots and mean values of pairwise Bray-curtis similarity of microbial functional and taxonomic diversities for “broad” versus “narrow” functions. **b**, PCoA (Principal coordinates analysis) showing beta-diversity of microbial taxonomic diversity for “broad” and “narrow” functions annotated using RefSeq at genus level (Taxonomy). The error bars represent the standard deviation of data ranges. Variations (by percentage) explained by the two principal coordinate dimensions are given in parentheses. *P* values and sq. root of PERMANOVA are also given. **c**, Heatmaps showing relative abundance of dominant microbial taxonomic composition (mean > 0.5%) for “broad” and “narrow” functions annotated using RefSeq at phylum/class levels (Taxonomy). “Broad” functions include AAD (Amino Acids and Derivatives), CHO (Carbohydrates), CBS (Clustering-based Subsystems), CVPGP (Cofactors, Vitamins, Prosthetic Groups, Pigments), and Protein (Protein Metabolism); “Narrow” functions include N (Nitrogen Metabolism), P (Phosphorous Metabolism), K (Potassium Metabolism), S (Sulfur Metabolism), and Fe (Iron Acquisition and Metabolism).

To investigate how microbial taxonomic diversities differ globally, the taxonomic compositions of soil microbes conducting the five “broad” and the five “narrow” functions were analyzed among the seventeen climate zones based on the PCoA analysis. Across climate zones, microbial taxonomic compositions of the five “narrow” functions (sq. root = 15.2-18.8) were more distinct than the five “broad” functions (sq. root = 13.4-15.1) based on the PERMANOVA analysis (Supplementary Fig. 2). This suggests that microorganisms relating to “broad” functions were similar to each other in taxonomy, because “broad” functions are more broadly distributed across most taxa, but soil microbes performing “narrow” functions were more phylogenetically diverse due to the specialty of “narrow” functions. Thus, though microbial metabolic functions can be strongly coupled to elemental cycles and certain environmental factors, the decoupling of

microbial taxonomic and functional profiles is still inevitable when a low-dimensional functional space is projected to a high-dimensional taxonomic space (Louca et al., 2018), especially for “broad” functions. Moreover, certain environmental factors may have significant effects on the coupling of taxonomy and function due to their already existent selective pressure, such as the extreme environment of ice cap, and thus future research can focus on comparison of relationship between function and taxonomy among terrestrial ecosystems of different selective pressure levels.

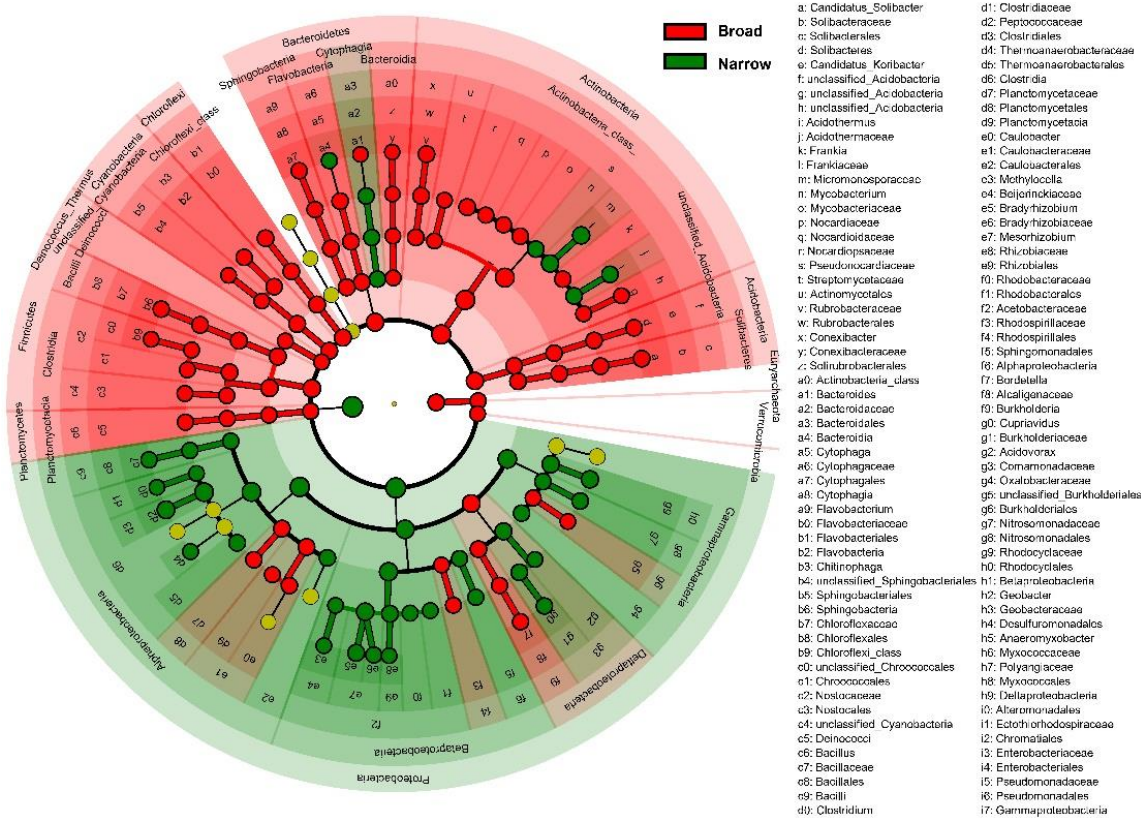


Fig. 3. Difference of taxonomic compositions between “broad” and “narrow functions”. LefSe (linear discriminant analysis effect size) results showing the significant differences in the relative abundance of dominant microbial taxonomic groups (mean > 0.5%) between “broad” (red) versus “narrow” (green) functions annotated using RefSeq (Taxonomy). From the center outward, each circle represents the level of domain,

phylum, class, order, family, and genus, respectively. The taxonomic groups with significant differences are labeled by colors.

The taxonomic compositions of microbes conducting the five “broad” functions were more abundant in most major phyla, such as Acidobacteria, Actinobacteria, Bacteroidetes, and Firmicutes, while the relative abundance of the taxonomic composition of microbes conducting the five “narrow” functions were greater in Proteobacteria, especially Alphaproteobacteria and Betaproteobacteria (Fig. 2c). Other studies also found that some bacteria conducting N cycling, such as ammonia-oxidizers and rhizobia for N fixation, mainly belong to Alphaproteobacteria or Betaproteobacteria (Stephen et al., 1996; Moulin et al., 2001).

To find out the dominant microbial groups that were statistically different between the five “broad” and the five “narrow” functions, LEfSe analysis was conducted based on the relative abundances at the taxonomic levels of domain, phylum, class, order, family, and genus. In particular, among the Proteobacteria conducting the five “narrow” functions, *Bacillaceae* from Bacilli, *Clostridium*, *Peptococcaceae*, and *Thermoanaerobacteraceae* from Clostridia, *Methylocella*, *Bradyrhizobium*, *Bradyrhizobiaceae*, and *Rhizobiaceae* from Rhodospirillaceae, and *Cupriavidus* from Comamonadaceae had higher relative abundance than the others (Fig. 3). The Venn’s diagrams indicated that the taxonomic compositions of soil microbes performing the “broad” functions shared 68% dominant genera among the five functional categories, while the proportion was reduced to only 41% for the five “narrow” functions (Fig. 4). However, it should be stated that all the analyses performed in our study were based on relative abundance data that is compositional, so it is difficult to directly compare

taxonomic diversities among samples and/or datasets. Despite the differences in the identification protocol and quantification of soil metagenomes, we deem the effects of these differences to be trivial for our analyses as we intended to understand the general patterns of microbial taxonomic and functional linkages, rather than simply compare soil community structures across samples. By uncovering universal patterns of these relationships within the microbial community, we can then further establish a potential linkage framework to account for the microbial contributions to major biogeochemical cycles.

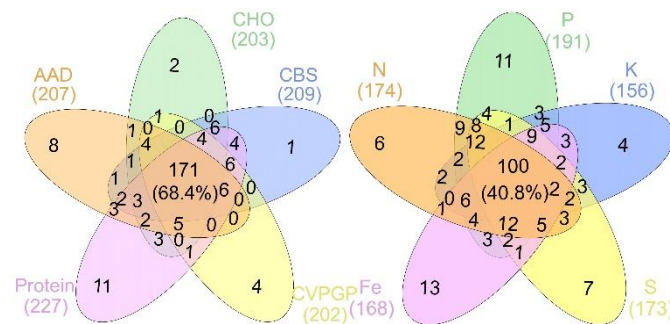


Fig. 4. Taxonomic compositions shared among “broad” and “narrow” functions. Venn’s diagrams showing dominant microbial taxonomic groups (mean > 0.1%) annotated using RefSeq at genus levels (Taxonomy) shared among “broad” and “narrow” functions.

Because of functional redundancy of soil microbes, understanding what types of functions that have more significant association with microbial taxonomy can be critical for accurate prediction of microbial metabolic activity and flexibility across space and time. As microbial taxonomic composition and diversity plays critical role in maintaining ecosystem function (Allison and Martiny, 2008), our results suggest that taxonomic information alone provides limited utility in predicting basic metabolic capabilities, but may be capable of forecasting biogeochemical transformations or changes in the rate of

biogeochemical process at ecosystem level (Hall et al., 2018). Investigating functional redundancy with respect to functions associated with elemental cycles provides useful information for guiding the development of explicit microbial biogeochemical prediction, and further delving into major pathways of C and N cycles will be a fruitful approach for scrutinizing microbes' functional potentials.

Table 1. Summary of key properties of co-occurrence networks for the five “broad” and the five “narrow” functions.

Network Indexes	Total nodes	Total links (positive%)	Average connectivity	Average clustering coefficient	Average geodesic distance	Modularity (modules numbers)
		1472				
AAD	225	(100%)	13.084	0.663	2.873	0.695 (11)
CHO	207	1155 (99%)	11.159	0.615	3.805	0.672 (10)
CBS	246	1622 (99%)	13.187	0.663	2.859	0.671 (11)
CVPGP	201	1293 (99%)	12.866	0.65	3.303	0.697 (9)
Protein	285	1651 (99%)	11.586	0.638	2.992	0.749 (14)
N	101	519 (12%)	10.277	0.349	1.903	0.184 (5)
P	160	449 (4%)	5.612	0.299	3.298	0.615 (10)
K	143	364 (67%)	5.091	0.08	2.676	0.429 (6)
S	132	264 (15%)	4	0.09	2.563	0.486 (12)
Fe	95	215 (11%)	4.526	0.071	2.601	0.435 (6)

3.2. Microbial taxonomic co-occurrence networks

To identify potential interaction patterns of microbial groups that conduct the five “broad” and the five “narrow” functions, the co-occurrence networks of taxonomic compositions were generated based on the taxonomic composition at the genus level across the globe. Network graphs with submodule structures indicated distinct topology of taxonomic networks between the “broad” and “narrow” functions (Table 1, Supplementary Fig. 3 and Supplementary Fig. 4). Compared to the “narrow” functions, the “broad” functions harbored larger and more complex networks with more nodes (201-285 vs. 95-160) and links (1293-1651 vs. 215-519), with higher average connectivity

(11.2-13.2 vs. 4.0-10.3) and average clustering coefficient (0.64-0.66 vs. 0.07-0.35). The “broad” function network had 99-100% positive links, while the “narrow” function had 33-96% negative links. These significant difference of network properties between “broad” and “narrow” functions suggests that taxonomic composition of “narrow” functions had both facilitative and inhibitive interactions, while taxonomic compositions of the “broad” function are all cooperative (Faust and Raes, 2012). Thus, soil microbes with “broad” functions tended to respond to the environment in a similar way, indicating functional sharing and association, while distinct microorganisms to conduct “narrow” functions competitively interact with each other, reflecting regulatory or suppression relationships (Delgado-Baquerizo et al., 2018).

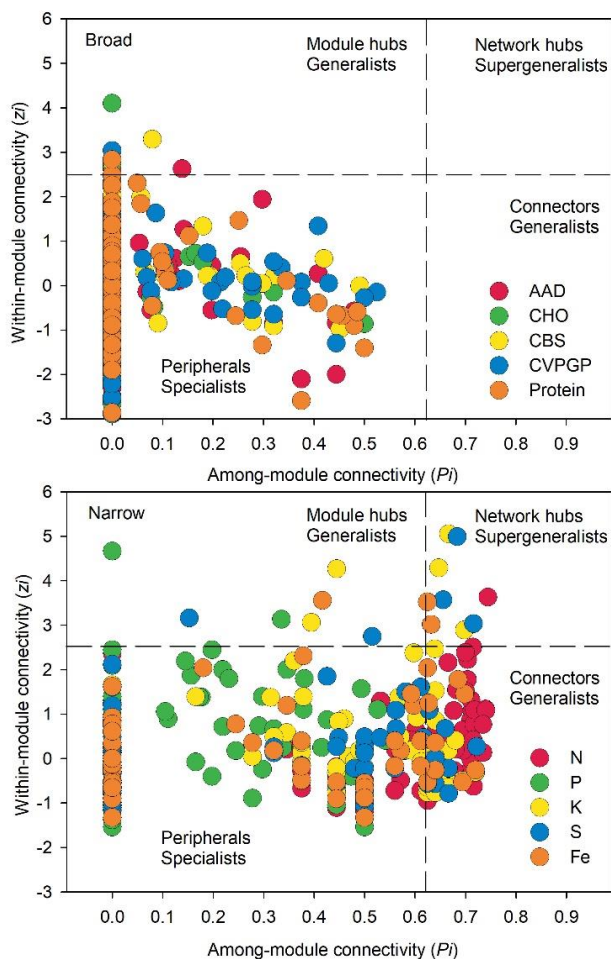


Fig. 5. Network information of taxonomic compositions for “broad” and “narrow” functions. Node distribution of module-based topological roles of taxonomic compositions for “broad” and “narrow” functions determined by the scatter plot of within-module connectivity (Z_i) and among-module connectivity (P_i). The threshold values of Z_i and P_i for categorizing were 2.5 and 0.62 respectively.

In addition, network modularity was greater in the “broad” functions, indicating that significant correlations between taxonomic compositions of microbes that conduct the five “broad” functions are mainly within similar taxonomic groups. No node could be classified as connectors in the five “broad” function networks (Fig. 5), reaffirming that the “broad” function networks had links mainly within modules of similar species. In the co-occurrence network of taxonomic composition of the “narrow” functions, 13% of the nodes were identified as connectors linking several modules (high P_i) connectors, while 3% were identified as module hubs that connected other nodes within their own modules (high Z_i), indicated by the Z_i - P_i plot (Olesen et al., 2007; Deng et al., 2012). Thus, significantly less nodes were identified as module hubs in the co-occurrence network of the taxonomic composition of the “broad” functions, indicating less correlations found among different modules. This is expected given that module was comprised of genera that were mainly from the same phylogenetic groups. This difference was consistent with the Venn’s diagrams showing significantly more nodes (54%) shared among the five functional categories representing the “broad” functions, while only 5% of the nodes were overlaid among the five “narrow” function networks (Fig. 6). Environmental conditions likely determine the microbial taxonomic composition, and microbial phylotypes sharing similar habitat preferences tend to co-occur (Delgado-Baquerizo et al., 2018; Ramírez-Flandes et al., 2019). We emphasize that this analysis is a combination

of snapshots of microbial communities compared across space, thus environmental conditions (at the same geographic location) may vary, and the levels of functional redundancy may change depending on the mechanisms selecting specific functions and the phylogenetic distribution of those functions (Louca et al., 2018).

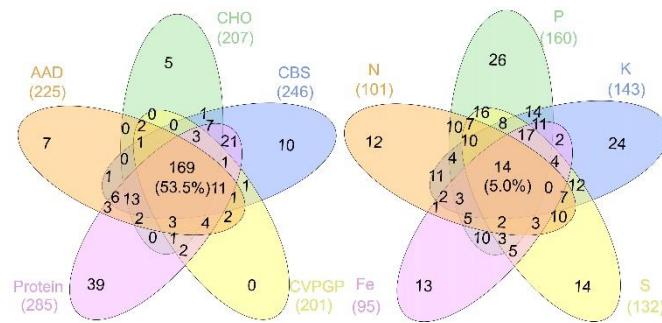


Fig. 6. Taxonomic network nodes shared among “broad” and “narrow” functions.

Venn’s diagrams showing the microbial taxonomic network nodes shared among “broad” and “narrow” functions.

3.3. Conclusion

By analyzing and generalizing microbial taxonomic and functional profiles, we provide strong evidence that the degree of soil microbial functional redundancy differs significantly between “broad” and “narrow” functions across the global. The level of functional redundancy varies depending on the functions of interest. Here, by contrasting the five “broad” metabolic functions and the five “narrow” functions that are important for elemental cycles, we found lower levels of functional redundancy associated with the five “narrow” functions of biogeochemical cycling, despite the fact that even for the five “narrow” functions, there is still a high level of functional redundancy in the soil communities. Although there is a caveat concerning direct comparison of metagenomic data, the present study demonstrated the use of comparative metagenome and co-

occurrence network analysis in generalizing patterns of microbial characteristics
regulating biogeochemical cycling of major elements. With the increasing advancement
of sequencing techniques and data coverage, future sequencing efforts will likely increase
our confidence in comparative metagenomes and provide time-series information to
further identify to what extent microbial functional redundancy regulates dynamic
ecological fluxes across space and time.

Author Contributions

Huaihai Chen conceived the study, performed the data analysis, interpreted the results,
and drafted the manuscript. JL, CWS, and Hao Chen secured the research funding. KM,
YH, QF, YQ, and Hao Chen critically assessed and interpreted the findings. All authors
discussed results, commented on, edited, revised, and approved the manuscript.

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Data Availability Statement

The data that support the findings of this study are available from the corresponding author upon request. All metagenomic data used in this study are publicly assessable in the MG-RAST server with study and MG-RAST ID reported in supplementary files.

Competing Interests

The authors declare no competing interests.

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