Article Type: Primary Research Article
Date of preparation: January 13, 2020
Number of text pages: 39
Number of tables: 3
Number of figures: 3

Relations between mineralized N delivered by soil food web and soybean yield after long-term application of conservation tillage system in a black soil of Northeast China

Shixiu Zhang[a], Liang Chang[a], Neil B. McLaughlin[b], Shuyan Cui[c,d], Haitao Wu[a, *], Donghui Wu[a], Wenju Liang[c], Aizhen Liang[a, *]

[a] Key Laboratory of Mollisols Agroecology, Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences, Changchun 130012, China
[b] Ottawa Research and Development Centre, Agriculture and Agri-Food Canada, Ottawa, K1A 0C6, Canada
[c] Institute of Applied Ecology, Chinese Academy of Science, Shenyang 110016, China
[d] Liaoning Normal University, Liaoning 110036, China

[*] **Corresponding authors:**

Dr. Haitao Wu
Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences, Changchun 130012, China.
Tel.: +8643188542272; E-mail address: wuhaitao@iga.ac.cn
Dr. Aizhen Liang
Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences, Changchun 130012, China.
Tel.: +8643188542349; E-mail address: liangaizhen@iga.ac.cn

Abstract

Long-term (10 years) application of conservation tillage following conversion from conventional tillage (CT) can achieve a new stable and equalized soil environment, which is vital to reverse soil biodiversity declines and fulfil the goal of maintaining agroecosystem sustainability. However, in such situation, how the whole soil organism communities regulate nutrient cycle impacting crop growth is not well documented. Therefore, the relations between mineralized nitrogen (N) delivered by soil food web and soybean (*Glycine max* Merr.) yield were investigated after 14 years continuous application of CT, reduced tillage (RT) and no tillage (NT) in a Black soil (Typic Hapludoll) of Northeastern China. We hypothesized that soil mineralizable N pool would increase with the complexity of soil food web, and that the trophic groups involved in associating N mineralization with plant growth vary with soil depth in the conservation tillage practice. During the soybean growing season, soil organisms, including bacteria, fungi, nematodes, mites and collembolans, were extracted and identified monthly from 0-5 cm and 5-15 cm to estimate species richness and connectance of the food web, and to simulate the mineralized N using the food web energetic model approach. The species richness and connectance of the food web at both soil depths were significantly affected by tillage practices, and their values decreased in the order of NT > RT > CT. Similar trend was also revealed for the simulated N mineralization, that is, the mineralized N released either from the functional feeding guilds or from the energy pathways of the food web were greater in RT and NT than in CT at both soil depths. Multiple linear regression analysis showed that soil organisms involved in coupling the mineralized N with soybean yield were different at different soil depths, in which fungal and root pathways at 0-5 cm and bacterial pathway at 5-15 cm were the driving factors for the supply of mineralized N to soybean in NT and RT soils. These results support our hypothesis and suggest that long-term application of conservation tillage can form a complex soil food web and expand the potentially mineralizable N pool, which is a cornerstone for conservation tillage system to achieve the sustainable crop productivity.

Key words: conservation tillage, soil food web energetic model approach, organism biomass, energy pathways, soil N mineralization

1. Introduction

Nitrogen (N) is the most important growth-limiting nutrient for crops (Fageria et al., 2010). In order to achieve the maximum yield, N fertilizer is applied to crops all over the world; even legumes that fix N through symbiotic N-fixing microorganisms require additional chemical N application for maximum yield (La Menza et al., 2020). However, globally, the N recovery rate by crops is only at most 60% (Liu et al., 2010), which means that the rest of the fertilizer N is not available for the crop and is lost by leaching or nitrous oxide emission, resulting in undesirable environmental issues. Hence, crop production to a great extent relies on the N mineralization to meet the growth requirements (La Menza et al., 2020; Whalen et al., 2013).

The process of N mineralization mediated by soil organisms is closely related to the predation across multi-trophic groups, because soil organisms require carbon (C), N and other nutrients from the prey to support their metabolic activities, ultimately converting the organic N compounds into the form of mineral N (de Ruiter et al., 1993; Whalen et al., 2013). The N immobilized in the biomass of the lower trophic groups can be released by the predation of the higher trophic groups. Furthermore, the predators usually have a higher C:N ratio than their prey, which results in more N obtained than their nutritional requirements, and the excess N is excreted into the soil ammonium (NH_4^+) pool (de Ruiter et al., 1993; Whalen et al., 2013). It is estimated that the N amount released from the predation of soil organisms accounts for 30%-80% of the annual N mineralization under field conditions (Carrillo et al., 2016; de Ruiter et al., 1993; Holtkamp et al., 2011), and the value of this contribution varies with the complexity of soil food webs (Carrillo et al., 2016; de Ruiter et al., 1993; Holtkamp et al., 2011). Several studies (Bender et al., 2015; Thakur et al., 2014; Wagg et al., 2014) based on controlled (micro- or meso-cosm) experiments demonstrated that the potentially mineralizable N pool increases with the increase in complexity of the food web, which implies that a management practice that forms a complex soil food web is beneficial for improving N availability with less N fertilizer input.

Conservation tillage, including reduced tillage (RT) and no tillage (NT) with at least 30% mulch cover of the soil surface, is becoming a popular practice around the world to counteract the disadvantage of conventional tillage (CT, soil inversion by moldboard plowing) on soil health (Lal, 2004). The benefits of conservation tillage on soil properties and processes, especially on crop productivity, are not immediately apparent, but can only be achieved after a period of time (5-10 years) when soil environment reaches a new stable equilibrium (Six et al., 2004). In such stable situation, soil biodiversity and its spatial heterogeneity are strongly enhanced, thereby constructing a more complex network among soil organisms relative to CT (de Vries et al., 2013; D'Hose et al., 2018; van Capelle et al., 2012). For example, bacteria and bacterivorous fauna dominate the whole plow layer of CT, while conservation tillage is typically characterized by the fungi and fungivorous fauna near the surface and bacterial based communities at deeper soil depths (D'Hose et al., 2018; van Capelle et al., 2012). Moreover, the increase in the richness and density of predaceous fauna reorganize the topological structure of food web through modification of the prey-predator interactions (Bartley et al., 2019). As a result, our understanding of how the N mineralization mediated by the entire food web assemblages promotes crop yields after long-term conservation tillage is still limited.

Soybean (*Glycine max* Merr.) is a major crop produced in the black soil region of Northeast China and accounts for 50% of the total national soybean production (Liu et al., 2019). Monoculture cropping, moldboard plowing, ridging, seeding into ridges, and the removal of post-harvest residue is the typical practice in this region, which has caused serious land degradation threatening soil fertility and sustainability (Zhang et al., 2019). Consequently, a national project to promote the application of conservation tillage in the Northeast China was launched in 2020 (Ministry of Agriculture of China and Ministry of Financing of China, MoAC and MFC, 2020). Reliable information regarding the responses of soil properties and grain yield to the conversion from CT to conservation tillage is needed to help the farmers better adapt agronomic measures.

The objective of this study was to investigate the relations between N mineralization by the soil food web and soybean yield under long-term conservation tillage system. We hypothesized that (1) conservation tillage favors a greater release of mineralized N than CT as it forms a more complex soil food web, (2) the trophic groups of soil organisms associated with N mineralization and plant growth vary with soil depth in the conservation tillage system, given the strengthened heterogeneity of organisms along the soil profile.

To address these hypotheses, soil organisms, including bacteria, fungi, nematodes, mites and collembolans, were extracted monthly during the soybean growing season after a long-term (2001-2015) application of conventional tillage (CT), reduced tillage (RT) and no-tillage (NT) in a black soil of Northeast china. The amount of mineralized N delivered by all trophic groups in the food web was calculated using the food web energetic model approach. This approach has been applied to a range of natural and agricultural systems and has been proven very useful in simulating N mineralization and in understanding the ecological functions served by soil organisms (Barnes et al., 2014; Carrillo et al., 2016; Koltz et al., 2018; Pressler et al., 2017; Schwarz et al., 2017), although it cannot reflect the dynamics N flow in the same way as the isotope tracing technique.

2. Material and methods

2.1 Experimental design and soil sampling

This study was conducted at the Experimental Station (44°12'N, 125°33'E) of the Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences, in Dehui County, Jilin Province, China. The station is located in a continental temperate monsoon zone. The mean annual temperature is 4.4°C; the lowest mean monthly temperature occurs in January (-21 °C) and the highest in July (23 °C). The mean annual precipitation is 520 mm and > 70% occurs from June to August. The soil is classified as Black soil (Typic Hapludoll, USDA Soil Taxonomy) with a clay loam texture (the average soil texture is 36.0% clay, 24.5% silt and 39.5% sand).

The present experiment was conducted as part of an ongoing long-term tillage and crop rotation experiment. The long-term tillage experiment was established in the fall of 2001 and included conventional tillage (CT), reduced tillage (RT) and no tillage (NT) in a two year maize (*Zea mays* L.) - soybean (*Glycine max* Merr.) rotation system with residue return. Each treatment has four replicates and the plot area is 5.2 × 20 m. Crops are sown at the end of April or early May and harvested in October every year, and then fallowed for 6 months over the winter when the soil is frozen. The CT treatment consists of fall moldboard plowing (20 cm) followed by a secondary seedbed preparation in spring by disking (7.5-10 cm), harrowing and ridge-building. In RT, the ridges (16 cm in height and 75 cm in width) are rebuilt with a cultivator in June of each year, a modified lister and scrubber are used to form and press the ridge. The soil of NT has no disturbance except for planting using a no-till planter (KINZE-3000NT, Williamsburg, IA, USA). After harvest, the aboveground residues are returned to the soil surface in all treatments to prevent the water and wind erosion in winter and early spring (Zhang et al., 2019). For RT and NT plots, maize residue is cut into about 30 cm pieces leaving a 30-35 cm standing stubble; soybean residue is directly returned to the soil surface. Residues in CT plots are removed prior to, and manually replaced on the soil surface after fall moldboard plowing and then mixed with the plow layer by discing and cultivation in the following spring.

Starter fertilizer was applied with the planter at a rate of 89 kg N ha^{-1}, 51 kg P ha^{-1} and 51 kg K ha^{-1} for maize, and 40 kg N ha^{-1}, 49 kg P ha^{-1} and 53 kg K ha^{-1} for soybean. Additionally, 45 kg ha^{-1} of N was top dressed at the V–6 stage (6 leaves) of maize. The application rates of N, P and K were the same in all tillage treatments and the N application rate was reduced by about 30% compared to the local conventional application rate (187 and 60 kg N ha^{-1} for maize and soybean, respectively).

2.2 Soil sampling

Soils were sampled for the present experiment at the end of each month from April to September 2015 during the soybean growing season. The precipitation during the growing season was 365 mm in 2015, which was located in the range of 330-605 mm across the past 10 years (2004-2014) and had no typhoon attack in 2015 (data obtained from China Meteorological Data Service Center, http://data.cma.cn/en). This suggests that our samples in 2015 can yield reliable results to understand the relations between mineralized N delivered by soil food web and soybean yield after long-term application of conservation tillage system. All types of soil organisms, including microbes, nematodes and microarthropods, were determined monthly except nematodes, which were only determined in April, June and August due to the limitation of labor. The nematode populations for non-sampled months were estimated by linear interpolation between adjacent sampling dates.

Seven soil cores (2.5 cm in diameter) in each plot were randomly collected from a depth of 15 cm and each core was separated into 0-5 and 5-15 cm sections. Soil cores were combined to form a single composite sample for each plot and depth. Samples were immediately taken to the lab and stored at 4 °C. Soil bulk density for each plot was determined in the 0-5 and 5-15 cm depths using a slide-hammer probe with a 5 cm core diameter. The mean monthly bulk density is presented in supplementary Table S1 (hereafter 'S' is used as the abbreviation of supplementary. After plants had reached the physiological maturity, soybean yield in 2015 was determined by hand-harvesting 3 m lengths of 6 interior rows from each plot. Grain yield samples were dried to a constant weight at 75 °C in an oven, and then corrected to 13.5% grain moisture content.

2.3 Soil mineral N in the field condition

The content of soil mineral N, determined by summing NO_3^- and NH_4^+, in the field condition was measured within 12 hours after soil samples were collected each month. Mineral N was extracted by 1 M KCl (soil : KCl = 1:2 w/v) and determined by a continuous flow analyzer (SAN++, Skalar, Netherlands).

2.4 Soil organism extraction

Microbial community was determined using the phospholipid fatty acid analysis (PLFA) as described by Bossio et al. (1998). Lipids were extracted from 8 g of freeze-dried soil with a Bligh and Dyer solution (chloroform: methanol: citrate buffer = 1: 2: 0.8 (v: v: v)). Polar lipids were separated from neutral lipids and glycolipids in a solid phase extraction column (Supelco Inc., Bellefonte, PA, USA) and transformed into fatty acid methyl esters with a mild alkaline methanolysis. Samples were then dissolved in hexane and analyzed in an Agilent 6850 series Gas Chromatograph with MIDI peak identification software (Version 4.5; MIDI Inc., Newark, DE, USA). Fatty acids were grouped as bacteria (14:0, i14:0, a14:0, 15:0, i15:0, a15:0, 15:1ω6c, 16:0, i16:0, a16:0, 16:1ω7c, 16:1ω9c, i17:0, a17:0, 17:1ω8c, 17:1ω9c,18:1ω7c, 18:0, 20:0), saprophytic fungi (18:1ω9c and 18:2ω6c) and arbuscular mycorrhizal fungi (AMF) (16:1ω5c) (Bach et al., 2010; Dempsey et al., 2013). Microbial biomass was estimated using the following conversion factors of fatty acid concentrations (nmol): bacterial biomass, 363.6 nmol = 1 mg C; saprophytic fungal biomass, 11.8 nmol = 1 mg C; and AMF biomass, 1.047 nmol = 1 μg C (Tsiafouli et al., 2015).

Nematodes were extracted from a 50 g soil sample (fresh weight) using a modified cotton-wool filter method (Liang et al., 2009). At least 100 nematode specimens from each sample were selected randomly and identified to genus level (see Table S2 for the list of identified taxa and Table S4 for the abundance) using an Olympus BX51 microscope (OLYMPUS, Tokyo, Japan) according to Bongers (1994). Nematodes were assigned into four trophic groups: bacterivores, fungivores, plant-parasites and omnivores-predators (Ferris, 2010). Body length and maximum body diameter of nematodes were measured using an ocular micrometer to calculate the nematode fresh body mass (μg) (Andrássy, 1956). Nematode biomass was estimated by assuming that the dry weight of a nematode is 20% of the fresh weight, and the C in the body is 52% of the dry weight (Ferris, 2010).

Microarthropods were extracted from 200 mL fresh soil using modified high-gradient Tullgren funnels (Crossley and Blair, 1991) for 120 h at room temperature. Individuals were collected and stored in vials containing 95% ethanol for identification. Mites and collembolans were identified to species or morphospecies level (see Table S3 for the list of identified taxa and Table S4 for the abundance) according to Christiansen and Bellinger (1980-1981), Balogh and Balogh (1992), Bellinger et al. (2019), Pomorski (1998) and Niedbala (2002). Soil microarthropods were allocated into four different functional groups: fungivorous (oribatid) mites, predaceous mites, fungivorous collembolans and omnivorous collembolans. Individual body length and width were measured to estimate the dry weight based on regression equations from the literature (Douce, 1976; Hódar, 1996). Mite and collembolan biomass were estimated by assuming the C in the body as 50% of the dry weight (Berg, 2001).

2.5 Modeling N mineralization by the food web

The first step in modeling the N mineralization by the food web was to construct a soil food web using the published feeding relationships (Fig. S1). All determined species of soil micro-flora and -fauna were grouped into six functional feeding guilds based on the trophic resources they exploit (Burns, 1989): bacteria, fungi, herbivorous feeders, bacterivorous feeders, fungivorous feeders, and predaceous feeders. For omnivorous/predaceous species, we constructed every possible predator-prey interaction. Omnivorous-predaceous nematodes were assumed to feed on all other nematode groups (Yeates et al., 1993). Omnivorous collembolans, which mainly feed on bacteria, fungi, plant and microfauna (de Vries et al., 2013), were proportionally assigned to bacterivorous, fungivorous, herbivorous and predaceous collembolans according to the assumption that their diet consists of 25% bacteria, 25% fungi, 25% plant and 25% other microfauna. Taking into account the changes in abundance of soil organisms over time, the biomass during the soybean growing season was estimated by summing the monthly biomass. The biomass unit of each functional guild was converted from mg C g^{-1} to mg C m^{-2} based the soil bulk density (g cm^{-3}) and thickness of the soil layer (cm). Furthermore, the complexity of soil food web was measured by the species richness (the number of taxa detected in the sample) and the connectance (expressed as the ratio of the number of actual links to the total number of possible feeding links) indices (Zhang et al., 2015).

The N mineralization was simulated using soil food web energetic model based on a mass-balance assumption that the energy flowing into the biomass of a group is equal to the energy flowing out through natural death and predation (Barnes et al., 2014; de Ruiter et al., 1993). Following equations were used to simulate the N mineralization delivered by soil organisms according to de Ruiter et al. (1993):

$$F_{ij} = \frac{w_{ij}B_i}{\sum_{k=1}^{n} w_{kj}B_k} \quad (1)$$

$$F = \frac{d_jB_j + P_j}{e_{ass} \times e_{prod}} \times F_{ij} \quad (2)$$

$$N_{min} = e_{ass} \times \left(\frac{1}{C:N_i} - \frac{e_{prod}}{C:N_j}\right) \times F \quad (3)$$

where, in equation 1, F_{ij} is the feeding preference of predator (j) on prey (i), which was calculated based on the density independent feeding preference of j on i (w_{ij}, dimensionless; listed in Table S5), n is the total number of potential prey types (k = 1, 2, 3… n), and B is the biomass of prey (mg C m^{-2}). In equation 2, F is the feeding rate of predator on prey (mg C m^{-2} yr^{-1}); d_j is the natural death rate of j (yr^{-1}); B_j is the biomass of j (mg C m^{-2}); P_j is the energy loss of j due to the predation (mg C m^{-2} yr^{-1}); e_{ass} and e_{prod} are the assimilation efficiency and production efficiency of j, respectively. In equation 3, N_{min} is the N mineralization mediated by the predation of j on i (mg N m^{-2} yr^{-1}); C:N_i and C:N_j is the body C:N ratio of prey (i) and predator (j), respectively. The parameters of d, e_{ass}, e_{prod}, C:N of soil organisms were taken from the literature and were presented in Table S6. It is impossible to measure and confirm each parameter value under field conditions, therefore these parameters were cited from the relevant studies that were also conducted on an agricultural system, and updated according to the latest reports.

The simulation of N mineralization was started with the top predators, which are considered to have no energy loss from the predation, and then proceeded to the lower trophic groups. Based on the specific primary actors that drive energy flow from the basal resource to the food web, the soil food web was further categorized into three energy pathways: 1) fungal pathway, in which the energy flux is driven by fungi and then flows to fungivores and their predators; 2) bacterial pathway, in which the energy flux is driven by bacteria and then flows to bacterivores and their predators; 3) root pathway, in which energy flux is driven by herbivores and then flows to their predators. The N mineralization was first estimated at the functional guild level by summing up contribution of all species within a functional guild, and then estimated at the level of each energy pathway (bacterial, fungal and root pathways), and then for the whole food web.

2.6 Statistical analyses

In our initial model, the omnivorous collembolans were assigned into bacterivores, fungivores, herbivores and predators in equal portions to model the mineral N flux within the soil food web. To assess the impact of this assumption affecting mineral N flux, a sensitivity analysis was performed by re-assigning omnivorous collembolans into fungivores and herbivores (50% each) according to Barnes et al. (2014). This acted as a null model with the least diet preferences and the difference from the initial model was expressed as a percentage.

Data were checked for normality and for homogeneity of variances prior to statistical analysis. If necessary, the data were ln(x + 1) transformed to meet the assumptions of ANOVA. Two-way analysis of variance was performed to test the effect of tillage, soil depth and their interaction on the biomass of each feeding guild, and the N mineralization of soil food webs. When their interaction was significant, multiple comparisons were performed based on post hoc test to determine if tillage effects were significant in each soil depth. Tukey's honestly significant difference test was used for means comparisons and a difference at the $P < 0.05$ level was considered statistically significant.

Forward stepwise multiple linear regression (MLR) was used to identify which energy pathways closely link the release of mineralized N to soybean yield at each soil depth. In stepwise regression, only one independent variable is considered at a time and another variable is added to the model at each step until no significant (P-value was set at 0.05) improvement in the percentage of explained variance is obtained. Prior to MLR, all parameters were min-max normalized to accurately preserve all relations of data value and prevent potential bias from the domination of variables with large numeric ranges over those with small numeric ranges. Min-max normalization subtracted the minimum value of an attribute from each value of the attribute and then divided the difference by the range of the attribute. The normalized data lay in the range [0, 1] (Jayalakshmi and Santhakumaran, 2011). All statistical analyses were performed in R software (R 3.4.0, R Development Core Team 2017) with the package '*car*' for ANOVAs and the package '*stats*' for MLR analyses.

3. Results

3.1 Soil mineral N and soybean yield

Tillage effect on the soil mineral N over the whole soybean growing season varied with soil depths. At 0-5 cm, the amount of soil mineral N was higher (P = 0.001) in RT than in CT, while an opposite trend was observed at 5-15 cm, with a lower ($P < 0.001$) amount in RT and NT than in CT. For the entire soil layer (0-15 cm), NT significantly (P = 0.027) decreased the amount of soil mineral N relative to CT. There was no statistical significance (P = 0.065) for differences in soybean yield among tillage treatments (Table 1) although there was a general trend of NT > RT > CT.

3.2 Metrics of soil food web

Tillage significantly influenced the complexity of soil food web, as indicated by the indices of species richness and connectance (Fig. 1). Compared with CT, the whole food web richness (P = 0.035) and connectance (P = 0.001) were significantly increased in NT at both soil depths, while only connectance (P = 0.045) was significantly increased in RT.

Compared to CT, NT and RT significantly increased the biomass of the whole food web by 33-56% at 0-5 cm and by 28-42% at 5-15 cm (Table 2). A similar trend was also found for the functional feeding guilds of bacteria, fungi, herbivores, bacterivores, fungivores and predators at both soil depths, with a higher biomass in RT and NT than that in CT ($P < 0.05$; Table 2).

3.3 Mineralization N modeled by soil food web

To visualize the mineralization N within the food web, an N flux network was constructed for different tillage systems at both soil depths (Fig. 2). When the omnivorous collembolans proportionally assigned into bacterivorous, fungivorous, herbivorous and predaceous collembolans were replaced with fungivorous and herbivorous collembolans in equal portions, there was a slight decrease in the total amount of mineralized N within the food web throughout all tillage systems (loss of 0.3% at 0-5 cm and 2% at 5-15 cm; Fig. 3). When considering functional feeding guilds, a sharp decrease in the mineralized N from bacterivores, fungivores and herbivores to the top predators was observed for all tillage systems (decreasing 22-83% at 0-5 cm and 2-24% at 5-15 cm), although there was an increase in the mineralized N from roots to herbivores and from fungi to fungivores.

Across the entire soybean growing season, RT and NT significantly ($P < 0.001$) increased the total amount of mineralized N within the food web by 33-41% at 0-5 cm and 28-38% at 5-15 cm relative to CT, and the maximum increase was observed in NT at both soil depths (Fig. 2 and Table S7). Furthermore, the mineralized N delivered by the energy pathways also varied with tillage systems (Fig. 2). Compared to CT, RT and NT significantly ($P < 0.001$) increased the amount of mineralized N in the bacterial and fungal pathways at both soil depths, while only NT significantly ($P = 0.001$) increased the amount of mineralized N in the root pathway at 0-5 cm (Table S7). The similar tillage effect pattern was also observed for the components within these energy pathways (Fig. 2 and Table S7). Compared with CT, NT significantly ($P < 0.05$) increased the mineralized N released from each component in the fungal and bacterial pathways, while RT only significantly ($P < 0.05$) increased the mineralized

N from basal resource to the primary decomposers and then to the intermediate microbial feeding fauna (fungivores and bacterivores) at both soil depths. For the components in the root pathway at both soil depths, a greater quantity of mineralized N from basal resource to herbivores was released in RT and NT than in CT ($P < 0.001$), and the mineralized N from herbivores to predators revealed no significant ($P > 0.05$) difference among CT, RT and NT.

3.4 Relation between mineralized N in the food web and soybean yield

At 0-5 cm, 83.6% of the variation in soybean yield was explained by the mineralized N released from fungal and root pathways (Table 3). Their relative contributions to the soybean yield decreased in the order of fungal pathway (0.557) > plant pathway (0.550), which means that when the min-max normalized N mineralization in the fungal pathway and plant pathway increases by one unit, the min-max normalized soybean production would correspondingly increase by 0.557 and 0.550 units respectively. At 5-15 cm, only the mineralized N delivered by bacterial pathway significantly affected soybean yield and accounted for 37.3% of the yield variance. Soybean yield would increase by 0.656 units when the bacterial pathway increases by one unit.

4. Discussion

Soil N availability, which is generally linear with the crop yield, highly depends on the pool of soil mineral N and mineralizable N regulated by soil organisms

(Fageria et al., 2010; Whalen et al., 2013). In this study, after long-term application of conservation tillage system, soybean yield in RT and NT was equal to or even higher than that in CT. This variation pattern is counter to the empirically observed soil mineral N, that is, the mineral N pool in either the lower soil layer (5-15 cm) or the entire layer (0-15 cm) was lower in RT and NT than in CT (Table 1). However, the simulation of N mineralization within the food web (Fig. 1 and Table S7) showed that the mineralizable N pool was decreased in the order of NT > RT > CT at both soil depths. These findings indicate that the potential mineralizable N pool has a greater responsibility than the soil mineral N pool over the growing season for the soybean yield in RT and NT relative to CT; a detailed discussion is presented below.

4.1 Performance of modeling N mineralization within the food web

A source of uncertainty in the simulation of mineralization N was the feeding preference assignment of omnivorous collembolans, which were allocated into equal portions of bacterivores, fungivores, herbivores and predaceous. The robustness of this assumption was tested using a sensitivity analysis by re-assigning the omnivorous collembolans into fungivores and herbivores (50% each), which resulted in at the most 3% loss in the mineralized N of the whole food web. However, when considering the functional feeding guilds, there was a dramatic decline in mineral N from bacterivores, fungivores and herbivores to the top predators (Fig. 2). These results indicate that the disparity between these two models highly depends on the feeding guilds, and accordingly, the assignment of species into the functional guilds should be done with caution.

To our knowledge, there is no literature to date that has comprehensively identified the feeding habits of collembolans, because they consume a wide spectrum of resources, including plant roots or litter, different types of soil microorganisms and metazoan soil fauna (Potapov et al., 2016). Additionally, collembolans can shift their diet from one food resource to another when choices are available (Chahartaghi et al., 2005; Endlweber et al., 2009). This inherently complex feeding nature of collembolans makes it difficult to correctly assign them to specific feeding guilds without using isotope tracer techniques. In this study, the diet of collembolans may change throughout the crop year according to the availability of basal food resources of growing plants and crop residue, and organisms higher up in the food web in the different tillage systems. Therefore, except for those specific species that feed on fungi, other species classified as omnivorous collembolans (Table S3) are reasonably treated as generalists. In summary, our presented model is robust in calculating the mineral N flux within the food webs under different tillage systems over the soybean growing season.

4.2 Tillage effects on the N mineralization within the food web

Consistent with our first hypothesis, the results showed that, as the structure of soil food web became more complex after the conversion from conventional tillage to conservation tillage, mineralized N released either from the functional feeding guilds or from the energy pathways of the food web was greater in RT and NT than in CT at both soil depths. Our result is in agreement with the reports of Bender et al. (2015), Carrillo et al. (2016) and de Vries et al. (2013) that farming practices favoring rich and abundant soil organisms can increase N availability. This may be due to the increase in the number of different kinds of species, leading soil organisms to release more N when they consume basal resources to create their own biomass (Holtkamp et al., 2011; Koltz et al., 2018). The higher biomass of the food web revealed in RT and NT than in CT (Table 2) further supports our results as more N would be released with the build-up of biomass. Additionally, the strengthened connectance between functional guilds (Fig. 1) also contributes to the increase in the amount of simulated N mineralization in RT and NT, because the tight interlinkage within trophic levels in the food web stimulates the N release from predation (Bender et al., 2015; Carrillo et al., 2016; Wagg et al., 2014). Therefore, after 14 years continuous application of conservation tillage, a large variety of organisms and complex interlinks among them expand the potentially mineralizable N pool.

4.3 Relations between N mineralization within the food web and soybean yield

Although RT and NT improved the amount of mineralized N within the food web over the soybean growing season, the multiple linear regression analysis showed that soil organisms involved in coupling the mineralized N with soybean yield were different along with the soil profile (Table 3). We found that the mineralized N released from fungal and root pathways was strongly related to the soybean yield at the surface of 0-5 cm, while at the 5-15 cm depth, only the mineralized N released from bacterial pathway significantly contributed to the yield. These results strongly support our second hypothesis that the trophic groups of soil organisms associated N mineralization with plant growth vary with soil depth.

Fungal and bacterial pathways have been considered as two very important parallel pathways in mediating N mineralization rate, and their relative importance varies with the changes in soil environment resulting from changes in management practice (de Vries et al., 2013; Kou et al., 2020; Wardle et al., 2004). In this study, there was an obvious spatial difference in the distribution of fungal and bacterial pathways, in which the fungal pathway at 0-5 cm and the bacterial pathway at 5-15 cm were the driving factors in relating N mineralization to the soybean yield. This vertical distribution pattern of fungal and bacterial pathways is not surprising, as many studies (D'Hose et al., 2018; Sun et al., 2016; van Capelle et al., 2012) have reported that fungal and bacterial communities, which are the primary decomposers of fungal and bacterial pathways, exhibit the same spatial pattern within the plow layer under conservation tillage system. Residues under conservation tillage were placed on the soil surface instead of being mixed with the soil, resulting in large soil pores (as indicated by lower soil bulk density in 0-5 cm; Table S1) and longer distance for soil microbes to gain access to nutrients in the upper soil. These environmental conditions are recognized as more suitable for the growth of fungal communities (Moore et al., 2005), thereby promoting energy transfer through fungi-based pathway.

Fungal and bacterial pathways differ in N process rate as their components have different metabolic strategies (de Vries et al., 2013; Wardle et al., 2004). In contrast to the "slower turnover rate" of the fungal pathway which favors N retention in the soil by immobilizing N in the biomass and organism-processed compounds, the bacterial pathway supports a faster N turnover rate, releasing more mineral N from biosynthesis into the soil solution (de Vries et al., 2013; Wardle et al., 2004; Whalen et al., 2013). In the present study, we found that the N mineralization from the bottom bacteria to the intermediate bacterial feeders, and then to the top predaceous feeders was greatly enhanced in NT soils, suggesting a tight interlinkage and effective energy transfer across trophic levels in the bacterial pathway. These features of the food web have been recognized to play a prominent role in promoting N turnover among immobilized and mobile forms (de Vries et al., 2013; Pressler et al., 2017; Wagg et al., 2014). Therefore, the enhanced N mineralization of bacterial-channel is expected to stimulate N mineralization and release more mineral N that can be readily absorbed by plants. This may partially explain why the severe shortage of soil mineral N empirically observed at 5-15 cm in NT soils (Table 1) during the growing season did not result in a compromise of soybean yield relative to CT.

Root pathway has been considered to have a very minor effect on N mineralization (Holtkamp et al., 2011; Pressler et al., 2017). In this study, the amount of mineralized N in root pathway was indeed the least among different energy pathways across tillage systems (Fig. 2). However, to our surprise, the multiple linear regression analysis showed that there was a positive association between the mineralized N in root pathway at 0-5 cm and soybean yield. This may be primarily due to the significant increase of mineralized N delivered by herbivores in root pathway under RT and NT (Fig. 2 and Table S6). Verschoor (2002) reported that the N mineralization of herbivores accounted for 10% of total N mineralization in a grassland system, and attributed these beneficial effects of herbivores to the activity of soil microbes that was stimulated by the increase in root exudates after infection by herbivores. In our study, most groups classified into herbivores are the facultative feeders. For example, herbivorous collembolans can switch their diet from plant roots to decaying litter (Endlweber et al., 2009). Therefore, we propose that the positive role of herbivores at 0-5 cm in RT and NT soil may partly be due to their manipulation of surface residues by fragmenting and mixing. Therefore, the surface area of litter in contact with soil microbes would be increased, which is beneficial for N mineralization (Soong et al., 2016).

5. Conclusion

Combining the experimental data and the soil food web energetic model approach, our results suggest that, after long-term (14 years) application, conservation tillage has a larger potentially mineralizable N pool as the soil food web becomes more complex relative to conventional tillage. Furthermore, soil organisms involved in associating mineralized N with soybean yield are different along with soil profile, in which the fungal and root pathways at 0-5 cm and the bacterial pathway at 5-15 cm are the key driving factors for the supply of mineralized N to plants. Given that our finding is based on simulations and assumptions of steady state soil biological communities resulting from a long duration of conservation tillage, more studies using isotope tracing technique across different durations of management practice are needed to gain insight into how the soil food web processes energy and nutrients to maintain agroecosystem service and sustainability.

Acknowledgments: This research was supported by the National Natural Science Foundation of China (No. 41401272 and 41430857), the Foundation of Excellent Young Talents in Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences (DLSYQ15001), the Jilin Province Science and Technology Development Plan Project (20190201116JC), and the Key Research Program of Frontier Sciences of Chinese Academy of Sciences (QYZDB-SSW-DQC035).

Date accessibility: all data are included in the manuscript and its supporting information.

Author contribution: S.X.Z, H.T.W and A.Z.L designed research; S.X.Z, S.Y.C and L.C performed research; W.J.L and W.D.H guided species classification; S.X.Z analyzed data; and S.X.Z, N.B.M, H.T.W and A.Z.L wrote this paper.

Competing interests: The authors declare that they have no known competing financial interests or personal relations that could have appeared to influence the work reported in this paper.

References

Andrássy, I.: Die rauminhalst- und gewichtsbestimmung der fadenwürmer, (Nematoden). Acta Zoologica Hungarica, 2(1), 1–15, 1956.

Bach, E.M., Baer, S.G., Meyer, C.K. and Six, J.: Soil texture affects soil microbial and structural recovery during grassland restoration, Soil Biology & Biochemistry, 42, 2182–2191, doi: 10.1016/j.soilbio.2010.08.014, 2010.

Balogh, J. and Balogh, P. (Eds.): The oribatid mites genera of the world, The Hungarian Natural Museum Press, Budapest, 1992.

Barnes, A.D., Jochum, M., Mumme, S., Haneda, N.F., Farajallah, A., Widarto, T.H. and Brose, U.: Consequences of tropical land use for multitrophic biodiversity and ecosystem functioning, Nature Communication, 5, 5351, doi: 10.1038/ncomms6351, 2014.

Bartley, T.J., McCann, K.S., Bieg, C., Cazelles, K., Granados, M., Guzzo, M.M., MacDougall, A.S., Tunney, T.D. and McMeans, B.C.: Food web rewiring in a changing world, Nature Ecology & Evolution, 3, 345–354, doi: 10.1038/s41559-018-0772-3, 2019.

Bellinger, P.F., Christiansen, K.A. and Janssens, F: Checklist of the Collembola of the World, Available at: http://www.collembola.org, 2019.

Bender, S.F. and van der Heijden, M.G.A: Soil biota enhance agricultural sustainability by improving crop yield, nutrient uptake and reducing nitrogen leaching losses, Journal of Applied Ecology, 52, 228–239, doi: 10.1111/1365-2664.12351, 2015.

Berg, M., de Ruiter, P., Didden, W. Janssen, M., Schouten, T. and Verhoef, H.: Community food web, decomposition and nitrogen mineralisation in a stratified Scots pine forest soil, Oikos, 94, 130–142, doi: 10.1034/j.1600-0706.2001.09121.x, 2001.

Bongers, T. (Eds): De Nematoden van Nederland. Vormgeving en technische realisatie, Uitgeverij Pirola, Schoorl, Netherlands, 1994.

Bossio, D.A., Scow, K.M., Gunapala, N. and Graham, K.J.: Determinants of soil microbial communities: effects of agricultural management, season, and soil type on phospholipid fatty acid profiles, Microbial Ecology, 36, 1–12, doi:10.1007/s002489900087, 1998.

Burns, T.P.: Lindeman’s contribution and the trophic structure of ecosystems, Ecology, 70(5), 1355–1362, doi:10.2307/1938195, 1989.

Carrillo, Y., Ball, B.A. and Molina, M.: Stoichiometric linkages between plant litter, trophic interactions and nitrogen mineralization across the litter - soil interface, Soil Biology & Biochemistry, 92, 102–110, doi: 10.1016/j.soilbio.2015.10.001, 2016.

Chahartaghi, M., Langel, R., Scheu, S. and Ruess, L.: Feeding guilds in Collembola based on nitrogen stable isotope ratios, Soil Biology & Biochemistry, 37, 1718–1725, doi: 10.1016/j.soilbio.2005.02.006, 2005.

Christiansen, K. and Bellinger, P. (Eds.): The collembola of north America north of the Rio Grande, Grinnell College, Grinnell Iowa, 1–1322, 1980–1981.

Crossley, D.A. and Blair, J.M.: A high-efficiency, low-technology tullgren-type extractor for soil microarthropods, Agriculture, Ecosystems & Environment, 34, 187–192, doi: 10.1016/0167-8809(91)90104-6, 1991.

de Ruiter, P.C., van Veen, J.A., Moore, J.C. Brussaard, M.L. and Hunt, H.W.: Calculation of nitrogen mineralization in soil food webs, Plant & Soil, 157, 263–273, doi: 10.1007/BF00011055, 1993.

de Vries, F.T., Thébault, E., Liiri, M. Birkhofer, K., Tsiafouli, M.A., Bjørnlund, L., Jørgensen, H.B., Brady, M.V., Christensen, S., de Ruiter, P. C., d'Hertefeldt, T., Frouz, J., Hedlund, K., Hemerik, L., Gera Hol, W.H., Hotes, S., Mortimer, S.R., Setälä, H., Sgardelis, S.P., Uteseny, K., van der Putten, W.H., Wolters, V. and Bardgett, R.D.: Soil food web properties explain ecosystem services across European land use systems, Proceedings of the National Academy of Sciences, 110, 14296–14301, doi: 10.1073/pnas.1305198110, 2013.

Dempsey, M.A., Fisk, M.C., Yavitt, J.B. Fahey, T.J. and Balser, T.C.: Exotic earthworms alter soil microbial community composition and function, Soil Biology & Biochemistry, 67, 263–270, doi: 10.1016/j.soilbio.2013.09.009, 2013.

D'Hose, T., Molendijk, L., Van Vooren, L., van den Berg, W., Hoek, H., Runia, W., van Evert, F., ter Berge, H., Spiegel, H., Sandèn, T., Grignani, C. and Ruysschaert, G.: Response of soil biota to non-inversion tillage and organic amendments: An analysis on European multiyear field experiments, Pedobiologia, 66, 18–28, doi: 10.1016/j.pedobi.2017.12.003, 2018.

Douce, G.K.: Biomass of soil mites (Acari) in Arctic coastal tundra. Oikos, 27, 324–330, 1976.

Endlweber, K., Ruess, L. and Scheu, S.: Collembola switch diet in presence of plant roots thereby functioning as herbivores, Soil Biology & Biochemistry, 41, 1151–1154, doi: 10.1016/j.soilbio.2009.02.022, 2009.

Fageria, N.K., Baligar, V.C. and Jones, C.A.: Growth and mineral nutrition of field crops, 3rd edn. CRC Press, Boca Raton, FL, 2010.

Ferris, H.: Form and function: Metabolic footprints of nematodes in the soil food web, European Journal of Soil Biology, 46, 97–104, doi: 10.1016/j.ejsobi.2010.01.003, 2010.

Hódar, J.A.: The use of regression equations for estimation of arthropod biomass in ecological studies, Acta Œcologica, 17, 421–433, 1996.

Holtkamp, R., van der Wal, A., Kardol, P. van der Putten, W.H., de Ruiter, P.C. and Dekker, S.C.: Modelling C and N mineralisation in soil food webs during secondary succession on ex-arable land, Soil Biology & Biochemistry, 43, 251–260, doi: 10.1016/j.soilbio.2010.10.004, 2011.

Jayalakshmi, T., and Santhakumaran, A.: Statistical normalization and back propagation for classification, International Journal of Computer Theory and Engineering, 3(1), 1793–8201, 2011.

Koltz, A.M., Asmus, A., Gough, L., Pressler, Y. and Moore, J.C.: The detritus-based microbial-invertebrate food web contributes disproportionately to carbon and nitrogen cycling in the Arctic, Polar Biology, 41, 1531–1545, doi: 10.1007/s00300-017-2201-5, 2018.

Kou, X.C., Ma, N.N., Zhang, X.K., Xie, H.T., Zhang, X.D., Wu, Z.F., Liang, W.J., Li, Q. and Ferris, H.: Frequency of stover mulching but not amount regulates the decomposition pathways of soil micro-food webs in a no-tillage system, Soil Biology & Biochemistry, 144, 107789, doi: 10.1016/j.soilbio.2020.107789, 2020.

La Menza, N.C., Monzon, J.P., Lindquist, J.L., Arkebauer, T.J., Knops, J.M.H., Unkovich, M., Specht, J.E. and Grassini, P.: Insufficient nitrogen supply from symbiotic fixation reduces seasonal crop growth and nitrogen mobilization to seed in highly productive soybean crops, Plant, Cell & Environment, 43, 1958-1972, doi: 10.1111/pce.13804, 2020.

Lal, R.: Soil carbon sequestration to mitigate climate change, Geoderma, 123, 1–22, doi: 10.1016/j.geoderma.2004.01.032, 2004.

Liang, W.J., Lou, Y.L., Li, Q., Zhong, S., Zhang, X.K. and Wang, J.K.: Nematode faunal response to long-term application of nitrogen fertilizer and organic manure in Northeast China, Soil Biology & Biochemistry, 41, 883–890, doi: 10.1016/j.soilbio.2008.06.018, 2009.

Liu, J.G., You, L.Z., Amini, M., Obersteiner, M., Herrero, M., Zehnder, A.J.B. and Yang, H.: A high-resolution assessment on global nitrogen flows in cropland, Proceedings of the National Academy of Sciences of the United States of America, 107(17), 8035–8040, doi: 10.1073/pnas.0913658107, 2010.

Liu, J.J., Yao, Q., Li, Y.S., Zhang, W., Mi, G., Chen, X.L., Yu, Z.H. and Wang, G.H.: Continuous cropping of soybean alters the bulk and rhizospheric soil fungal communities in a Mollisol of Northeast PR China, Land Degradation & Development, 30, 1725–1738, doi: 10.1002/ldr.3378, 2019.

Moore, J.C., McCann, K. and de Ruiter, P.C.: Modeling trophic pathways, nutrient cycling, and dynamic stability in soils, Pedobiologia, 49, 499–510, doi: 2005,

Niedbala, W. (Ed.): Ptyctimous Mites (Acari, Oribatida) of the Nearctic Region, Monographs of the Upper Silesian Museum, 2002.

Pomorski, R.J.: Onychiurinae of Poland (Collembola: Onychiuridae), Polskie Towarzystwo Taksonomiczne, Genus (Supplement): 1–201, 1998.

Potapov, A.A., Semenina, E.E., Korotkevich, A.Y., Kuznetsova, N.A. and Tiunov, A.V.: Connecting taxonomy and ecology: Trophic niches of collembolans as related to taxonomic identity and life forms, Soil Biology & Biochemistry, 101, 20–31, doi: 10.1016/j.soilbio.2016.07.002, 2016.

Pressler, Y., Foster, E.J., Moore, J.C. and Cotrufo, M.F.: Coupled biochar amendment and limited irrigation strategies do not affect a degraded soil food web in a maize agroecosystem, compared to the native grassland, Global Change Biology Bioenergy, 9, 1344–1355, doi: 10.1111/gcbb.12429, 2017.

Schwarz, B., Barnes, A.D., Thakur, M.P., Brose, U., Ciobanu, M., Reich, P.B., Rich, R.L., Rosenbaum, B., Stefanski, A and Eisenhauer, N.: Warming alters energetic structure and function but not resilience of soil food webs, Nature Climate Change, 7, 895–900, doi: 10.1038/s41558-017-0002-z, 2017.

Six, J., Ogle, S.M., Jay breidt, F., Conant, R.T., Mosier, A.R. and Paustian, K.: The potential to mitigate global warming with no-tillage management is only realized when practised in the long term, Global Change Biology, 10, 155–160, doi: 10.1111/j.1529-8817.2003.00730.x, 2004.

Soong, J.L. and Nielsen, U.N.: The role of microarthropods in emerging models of soil organic matter, Soil Biology & Biochemistry, 102, 37–39, doi: , 2016.

Sun, B.J., Jia, S.X., Zhang S.X., McLaughlin N.B., Zhang, X.P., Liang A.Z., Chen, X.W., Wei, S.C. and Liu, S.Y.: Tillage, seasonal and depths effects on soil microbial properties in black soil of Northeast China, Soil & Tillage Research, 155, 421–428, doi: 10.1016/j.still.2015.09.014, 2016.

Thakur, M.P., van Groenigen, J.M., Kuiper, I. and de Deyn, G.B.: Interactions between microbial-feeding and predatory soil fauna trigger N_2O emissions, Soil Biology & Biochemistry, 70, 256–262, doi: 10.1016/j.soilbio.2013.12.020, 2014.

Tsiafouli, M.A., Thébault, E., Sgardelis, S.P. de Ruiter, P.C., van der Putten, W.H., Birkhofer, K., Hemerik, L., de Vries, F.T., Bardgett, R.D., Brady, M.V., Bjornlund, L., Jørgensen, H.B., Christensen, S., D' Hertefeldt, T., Hotes, S., Gera Hol, W.H., Frouz, J., Liiri, M., Mortimer, S.R., Setälä, H., Tzanopoulos, J., Uteseny, K., Pižl, V., Stary, J., Wolters, V. and Hedlund, K.: Intensive agriculture reduces soil biodiversity across Europe, Global Change Biology, 21, 973–985. doi: 10.1111/gcb.12752, 2015.

van Capelle, C., Schrader, S. and Brunotte, J.: Tillage-induced changes in the functional diversity of soil biota - a review with a focus on German data, European Journal of Soil Biology, 50, 165–181, doi: 10.1016/j.ejsobi.2012.02.005, 2012.

Verschoor, B.C.: Carbon and nitrogen budgets of plant-feeding nematodes in grasslands of different productivity, Applied Soil Ecology, 20, 15–25, doi: 10.1016/S0929-1393(02)00010-0, 2002.

Wagg, C., Bender, S.F., Widmer, F. and van der Heijden, M.G.A.: Soil biodiversity and soil community composition determine ecosystem multifunctionality, Proceedings of the National Academy of Sciences, 111, 5266–5270, doi: 10.1073/pnas.1320054111, 2014.

Wardle, D.A., Bargett, R.D., Klironomos, J.N., Setälä, van der Putten, W.H. and Wall, D.H.: Ecological linkages between aboveground and belowground biota, Science, 304, 1629–1633, doi: 10.1126/science.1094875, 2004.

Whalen, J.K., Kernecker, M.L., Thomas, B.W., Sachdeva, V. and Ngosong, C.: Soil food web controls on nitrogen mineralization are influenced by agricultural practices in humid temperate climates, CAB Reviews, 8, 1–18, doi:10.1079/PAVSNNR20138023, 2013.

Yeates, G.W., Bongers, T., De Goede, R.G.M., Freckman, D.W. and Georgieva, S.S.: Feeding habits in soil nematode families and genera - An outline for soil ecologists, The Journal of Nematology, 25, 315–331, 1993.

Zhang S.X., Li, Q., Lü, Y., Sun, X.M., Jia, S.X., Zhang, X.P. and Liang, W.J.: Conservation tillage positively influences the microflora and microfauna in the black soil of Northeast China, Soil & Tillage Research 149, 46–52, doi: 10.1016/j.still.2015.01.001, 2015.

Zhang, Y., Li, X., Gregorich, E.G., McLaughlin, N.B., Zhang, X.P., Guo, F., Gao, Y. and Liang, A.Z.: Evaluating storage and pool size of soil organic carbon in degraded soils: Tillage effects when crop residue is returned, Soil & Tillage Research, 192, 215–221, doi: 10.1016/j.still.2019.05.013, 2019.

Table 1 Effects of tillage systems on soybean yield and the cumulative mineral N concentrations (means (SD)) during the soybean growing season.

	CT	RT	NT
Soybean yield* (kg ha^{-1})	1242 (96) a	1324 (189) a	1570 (221) a
Soil mineral N (g m^{-2})			
0-5 cm	15.27 (1.44) b	20.09 (2.90) a	17.90 (1.46) ab
5-15 cm	28.10 (1.05) a	21.33 (1.79) b	20.06 (2.14) b
0-15 cm	21.68 (0.65) a	20.71 (1.86) ab	18.98 (0.67) b

CT, conventional tillage; RT, ridge tillage; NT, no tillage. Same lowercase letter in the same row indicates no significant difference among tillage systems ($P > 0.05$).

Table 2 Cumulative soil biotic biomass (expressed as mg C m^{-2}) over the soybean growing season under different tillage practices (means (SD)).

	0-5 cm			5-15 cm			ANOVA		
	CT	RT	NT	CT	RT	NT	Tillage (T)	Depth (D)	T × D
Total biomass	22727	30186	35498	27761	35598	39289	< 0.001	< 0.001	ns
	(1837)	(1849)	(5955)	(3725)	(4893)	(2385)			
Bacteria	6077	7367	8452	9000	11393	12780	< 0.001	< 0.001	ns
	(499)	(363)	(1408)	(1362)	(1324)	(733)			
Fungi	16386	22375	26646	18558	23938	26168	< 0.001	ns	ns
	(1309)	(1639)	(7661)	(2409)	(3622)	(1769)			
Herbivorous feeders	67	90	95	73	87	110	0.017	ns	ns
	(5)	(23)	(13)	(4)	(25)	(18)			
Bacterivorous feeders	78 b	168 a	128 a	56 b	93 a	112 a	< 0.001	< 0.001	0.045
	(15)	(17)	(30)	(14)	(13)	(11)			
Fungivorous feeders	58	98	99	34	57	55	0.023	0.002	ns
	(15)	(12)	(19)	(12)	(16)	(24)			
Predaceous feeders	60	88	78	96	123	176	0.002	< 0.001	ns
	(14)	(14)	(8)	(15)	(22)	(49)			

CT, conventional tillage; RT, ridge tillage; NT, no tillage; ns indicate no significant difference ($P > 0.05$). Means for the different tillage systems at the same depth and followed by the same lowercase letter are not significantly different ($P > 0.05$).

Table 3 Relations between modeled mineral N delivered by different energy channels and soybean yield based on multiple linear regression. Data were min-max normalized and are dimensionless.

Soil depth (cm)	Variable	Beta standardized coefficient	T value	Adjusted R^2	F value of the regression
0–5	Fungal channel	0.557	2.886*	0.836	19.737**
	Plant channel	0.550	2.437*		
5–15	Bacterial channel	0.656	2.745*	0.373	7.555*

* and ** indicate significant at 0.05 and 0.01, respectively.

Figure legends

Fig. 1 Changes in soil food web complexity (indicated by richness (A) and connectance (B)) under different tillage systems. CT, conventional tillage; RT, ridge tillage; NT, no tillage.

Fig.2 Modeled mineral N flux (mg N m^{-2} $year^{-1}$) through the fungal, bacterial and root pathways in the food webs of different tillage systems during the soybean growing season. Fungal, bacterial and root pathways are the energy fluxes driven on the base of fungal, bacterial and herbivorous communities. Vertical width of bars represents the amount of mineral N transferred between the trophic levels. CT, conventional tillage; RT, ridge tillage; NT, no tillage.

Fig. 3 Differences (%) in the modeled mineral N delivered by the food web between Model 1 that assigned the omnivorous collembolans into bacterivorous, fungivorous, herbivorous and predaceous collembolans in equal portions and Model 2 that assigned the omnivorous collembolans into fungivorous and herbivorous collembolans in equal portions. CT, conventional tillage; RT, ridge tillage; NT, no tillage.

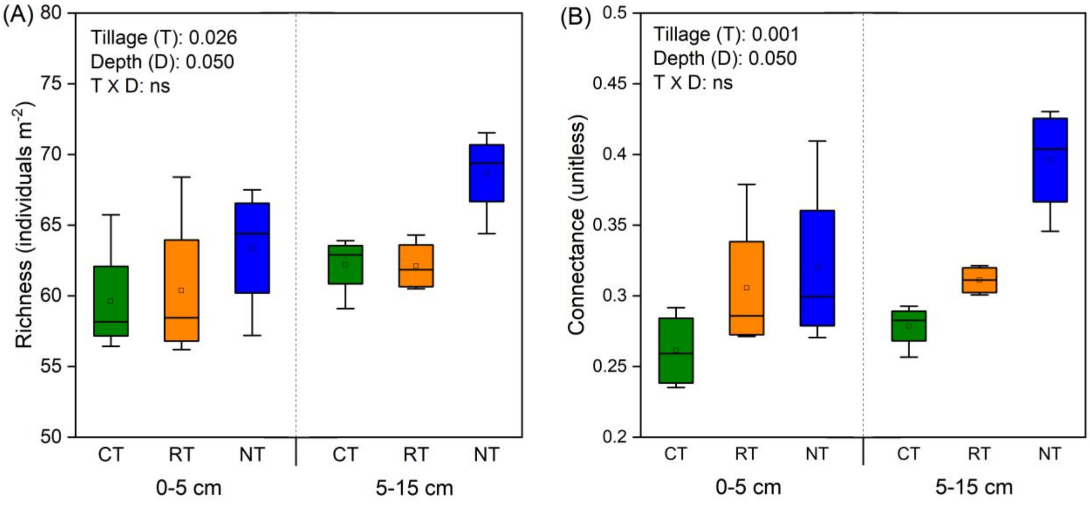
(A)
Tillage (T): 0.026
Depth (D): 0.050
T X D: ns
Richness (individuals m^{-2})
80
75
70
65
60
55
50
CT
RT
NT
CT
RT
NT
0-5 cm
5-15 cm
(B)
Tillage (T): 0.001
Depth (D): 0.050
T X D: ns
Connectance (unitless)
0.5
0.45
0.4
0.35
0.3
0.25
0.2
CT
RT
NT
CT
RT
NT
0-5 cm
5-15 cm

Fig. 1

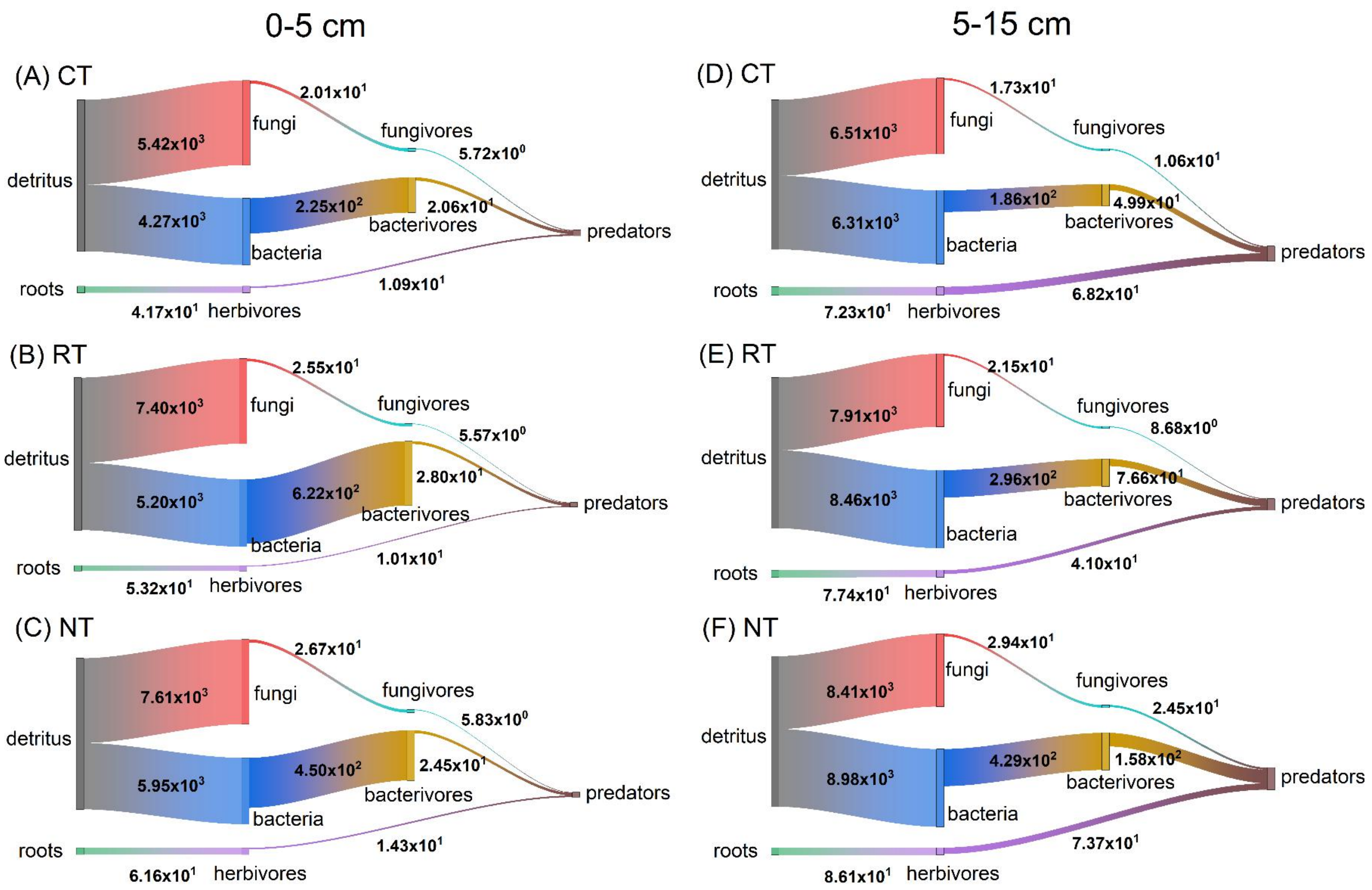
0-5 cm
5-15 cm
(A) CT
detritus
5.42x10^3
fungi
2.01x10^1
fungivores
5.72x10^0
4.27x10^3
bacteria
2.25x10^2
bacterivores
2.06x10^1
predators
roots
4.17x10^1
herbivores
1.09x10^1
(D) CT
detritus
6.51x10^3
fungi
1.73x10^1
fungivores
1.06x10^1
6.31x10^3
bacteria
1.86x10^2
bacterivores
4.99x10^1
predators
roots
7.23x10^1
herbivores
6.82x10^1
(B) RT
detritus
7.40x10^3
fungi
2.55x10^1
fungivores
5.57x10^0
5.20x10^3
bacteria
6.22x10^2
bacterivores
2.80x10^1
predators
roots
5.32x10^1
herbivores
1.01x10^1
(E) RT
detritus
7.91x10^3
fungi
2.15x10^1
fungivores
8.68x10^0
8.46x10^3
bacteria
2.96x10^2
bacterivores
7.66x10^1
predators
roots
7.74x10^1
herbivores
4.10x10^1
(C) NT
detritus
7.61x10^3
fungi
2.67x10^1
fungivores
5.83x10^0
5.95x10^3
bacteria
4.50x10^2
bacterivores
2.45x10^1
predators
roots
6.16x10^1
herbivores
1.43x10^1
(F) NT
detritus
8.41x10^3
fungi
2.94x10^1
fungivores
2.45x10^1
8.98x10^3
bacteria
4.29x10^2
bacterivores
1.58x10^2
predators
roots
8.61x10^1
herbivores
7.37x10^1

Fig. 2

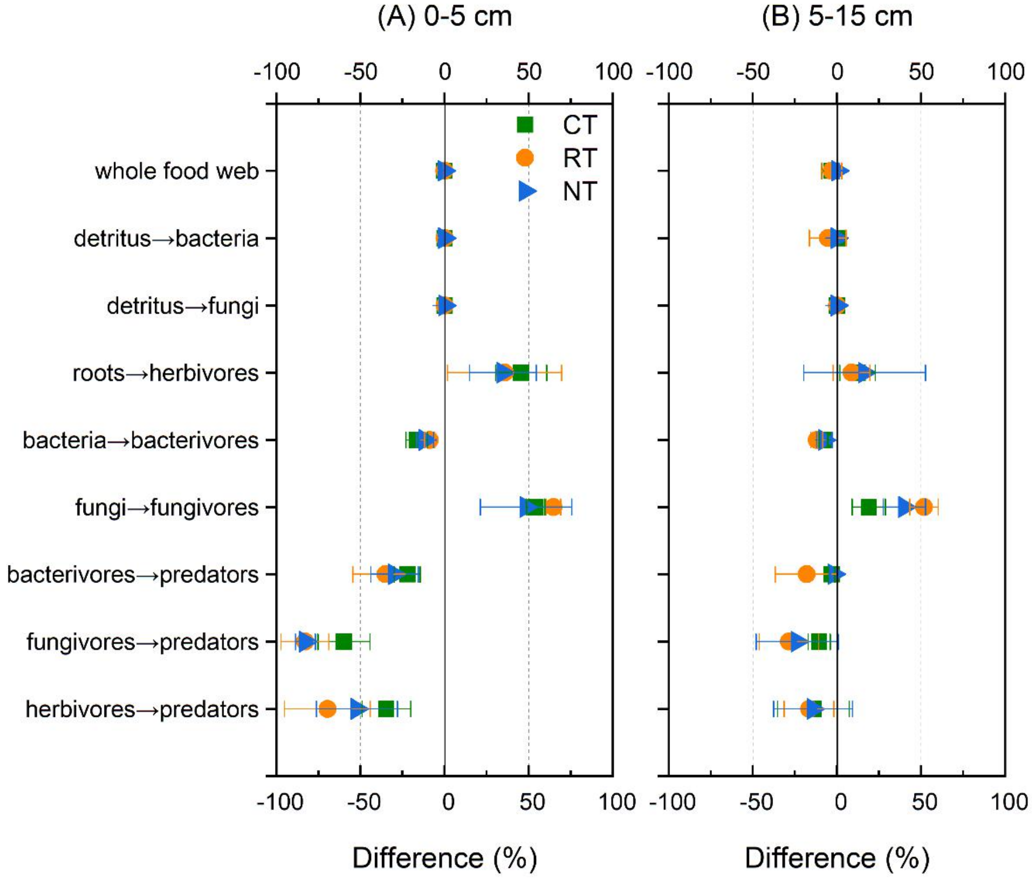
(A) 0-5 cm
(B) 5-15 cm
-100
-50
0
50
100
CT
RT
NT
whole food web
detritus→bacteria
detritus→fungi
roots→herbivores
bacteria→bacterivores
fungi→fungivores
bacterivores→predators
fungivores→predators
herbivores→predators
Difference (%)
Difference (%)

Fig. 3