











#### 28 ABSTRACT

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30 In Egypt, the total area of agricultural land is 3.36 million acres, which is 3.8% of the entire 31 territory of the country. One of the main obstacles to agricultural production in Egypt is soil salinization and degradation. Therefore, saline-sodic soil reclamation in arid regions is highly relevant. 32 33 This study aimed to use Azospirillum inoculation with eco-friendly organic wastes for free remediation of saline-sodic soils. In this work nine treatments included two levels of spent grain (S1 and S2), two 34 levels of compost (C1 and C2), a mix of both sources (C1S1), one level of Azospirillum (A1), a mix of 35 both sources with Azospirillum (A1S1 and A1C1) and an untreated control. The treatments were 36 previously incubated with soil at field capacity for five months under laboratory conditions at 28° C. 37 38 The most relevant chemical and biological parameters were analysed every month for five months. Results indicate that Azospirillum inoculation with spent grain increased soil organic carbon (SOC), 39 dehydrogenase and urease enzymes, micro-nutrients (Fe<sup>2+</sup>, Zn<sup>2+</sup>, Mn<sup>2+</sup>, Cu<sup>2+</sup> and B<sup>+</sup>), and macro-40 nutrients (N, P and K); while decreased exchangeable sodium percentage (ESP), pH and EC, by 75 %, 41 42 12%, and 43 % respectively, compared to initial conditions. The significant variation was observed in 43 chemical and biological properties among all treatments in the order of S2>A1S1>A1>C2>C1S1>A1C1>C1>control. In conclusion, the addition of Azospirillum with spent 44 grain is highly recommended for amelioration of the saline-sodic soil and is more effective compared 45 46 with compost to remediate saline-sodic soils.

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48 Keywords: Soil incubation, Azospirillum, amelioration, dehydrogenase, spent grain, saline-sodic

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## INTRODUCTION

51 One of the main obstacles to agricultural production in Egypt is soil salinization, which lead 52 to land degradation (Rashad et al., 2010). Soil salinization and degradation subsequent to salinization 53 is a key environmental impediment that severely influences agricultural productivity and sustainability in arid and semi-arid regions (Qadir et al., 2008). Globally salt-affected soils account for 54 approximately 46 million hectares (20%) of agricultural land (Ahmad et al., 2013). More than 60 % of 55 56 salt-affected soils classified as saline-sodic soils, the saline-sodic soils possess  $EC > 4 dSm^{-1}$ , pH >8.2 and exchangeable sodium percentage (ESP) >15 % (Rath and Rousk, 2015). The remediation of saline-57 sodic soils requires removal of Na<sup>+</sup> ions from the exchange areas and leaching of the substituted Na<sup>+</sup> 58 out of the root zone in percolating water (Sastre-Conde et al., 2015; Zhang et al., 2019). The chemical 59 amendments as desulfurization gypsum open a source of  $Ca^{2+}$  to substitute the exchangeable sodium 60 from the cation exchange areas (Luo et al., 2018), which plays a very important role in the ameliorant 61 62 of saline-sodic soils. The worldwide expansion of saline-sodic soils is increasing due to





mismanagement and using low quality of irrigation water. The soil called saline-sodic due to its high 63 64 ESP, cultivation of saline-sodic soils faces many challenges, such as poor soil structure, low soil stability aggregates, soil crusting, low hydraulic conductivity, low water infiltration, low water holding 65 capacity, bulk density is high, alkalinity, high Na<sup>+</sup> concentrations in soil solution, low soil fertility 66 (Dodd et al., 2013; Rath and Rousk, 2015), low biological activity, nutrient deficiencies, microbial 67 community and soil fertility (Zhang et al., 2019). Effects of salinity, including low agricultural 68 productivity, low economic returns and soil erosion (Hu and Schmidhalter, 2005). The salt effects are 69 70 the results of complex interactions between the physiological, biochemical and morphological 71 processes, including plant growth, nutrients, seed germination, and water absorption (Shrivastava and Kumar, 2015; Zhang et al., 2019). Soil salinity imposes ionic toxicity, osmotic pressure and nutrients 72 73 (N, Ca, K, P, Fe, and Zn) deficiency and oxidative stress on plants, thus limiting the absorption of 74 water from the soil. Soil salinity significantly reduces the absorption of phosphorus (P) due to 75 phosphate ions are deposited with calcium ions (Bano and Fatima, 2009). Some elements, such as Na<sup>+</sup>, 76 Cl<sup>-</sup>, and B<sup>+</sup>, have specific toxic effects to plants. Excessive accumulation of Na<sup>+</sup> in cell walls can quickly lead to osmotic pressure and cell death (Munns, 2002). Soil organic amendments can improve 77 soil physical and chemical properties and microbial activity in saline-sodic (Luo et al., 2018; Zhen et 78 79 al., 2014)

80 Application of agro-industrial organic wastes as soil amendments such as the Brewers' spent 81 grain, 85 % of the total by-products generated and compost with bacteria inoculation to reclamation saline soil. Organic additives have been used from the residues of the beryl industry, these wastes are 82 inexpensive, and their accumulation causes severe damage to the environment (Hafez et al., 2019; 83 84 Mussatto, 2014; Nassar et al., 2014). Spent grain has a negative value for environmental quality due to 85 its accumulation as industrial waste. It has been recently applied as an environment-friendly organic 86 waste to the soil, as well as composting of agricultural biomass, to improve physical and chemical properties of calcareous and sandy soils (Hafez et al., 2019; Rashad et al., 2016). Spent grain is acidic, 87 88 rich in macro and micronutrients, amino acids, and vitamins as well as cellulose, hemicelluloses, lipid, protein, arabinoxylan, ash, and lignin (Mussatto and Roberto, 2006). Therefore, it is a good candidate 89 90 for recycling in agriculture. The environmental systems using a low level of external input have 91 become more important all over the world in the last few years from natural resources, and reduction 92 of environmental systems degradation (Mader, 2002)

93 In this context, organic wastes incubation with bacteria encourages the use of agricultural 94 practices and alternative technologies, according to economic situation, social and environmental 95 conditions of the region the system is in particular because it refers to fertilization, which is usually





96 done by application of organic matter with *Azospirillum* bacteria such as spent grain, compost, plant
97 residues, or manure, a lot of which are produced within the same farm (Hafez *et al.*, 2019).

98 On the other hand, application of eco-friendly organic wastes and bacteria to the saline-sodic soil not only generates a better food situation, but moreover, positively affects other properties such as 99 soil particle assembly, water holding capacity and increase biological activity (Hafez et al., 2019; 100 Hayat et al., 2010; Scherer et al., 2011), contribute to generating high production, even with low 101 application or none of the fertilizer. In the aspect incubation system for organic wastes with micro-102 organisms are found in soil (Zhuang et al., 2007) incubated decreased soil salinity and increased the 103 content of nitrogen, phosphorus, and potassium (Rashad et al., 2016; Rojas-Tapias et al., 2012). The 104 application of biofertilizers, plant growth promoters and organic amendments enhancing soil fertility, 105 106 microbial activities and salt-affected. In addition, plant growth-promoting bacteria (PGPB) have been recently applied as a biofertilizer in soil to increase nitrogen fixation and phosphate availability and 107 108 reduce soil salinity (Hafez et al., 2019; Rojas-Tapias et al., 2012; Zhen et al., 2014; Shrivastava and Kumar, 2015; Zarea et al., 2012). 109

110 One of the PGPB is the Azospirillum brasilense, this is free-living bacteria capable of affecting the growth and seed yield of numerous plant species and many agronomic and ecological 111 significance. Plants treated with PGPB producing bacteria display increased resistance to salinity stress 112 due to improved soil structure (Karagöz et al., 2012; Zhuang et al., 2007). PGPB can also bind to 113 cations including Na<sup>+</sup> thus making it unavailable to plants under saline conditions. (Zhao et al., 2018). 114 Egypt, is located in the arid region therefore, the study of salinity in such dry land area has become of 115 great interest. We hypothesized that the applied spent grain with Azospirillum inoculation, for tow 116 117 period 60 and 150 days of incubation, would reclaimed saline-sodic soil while increasing soil fertility and reduce the ESP. The objectives of this work were to (i) the effect of eco-friendly organic wastes 118 doses with Azospirillum inoculation on some chemical and biological properties of saline-sodic soil in 119 the North-Western part of Egypt on reduce soil salinity, (ii) increase the content of beneficial microbes 120 121 and activity of enzymes, and (iii) increase soil fertility in regards to the SOC improve the availability of macro and micro-nutrients in saline-sodic soil after 60 and 150 days of incubation. 122

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#### 2-MATERIALS AND METHODS

125 2.1. Site Information

The study sites are located at the farm of El-Bangar village 28, El-Hamam City, Matrouh Governorate, North-Western part of Egypt (30°45'44.47"N 29°25'26.54"E). The soil samples are saline-sodic typically mixed with rocks. Due to its inadequate soil, water regime, salinity in arid regions can occur when the water table is between two and three meters from the surface of the soil. The use and potential effect of soil enhancer materials are important and current raise. The study site





has a desert climate arid, hot summer and semi-cool and semi-wet winters. The climate is classified as BWh by the Köppen-Geiger system (Climate-Data.org, 2019). The average air temperature in El-Hamam is 19.2 °C. In a year, Annual precipitation is 124 mm. The temperatures are higher on average in August, at around 27.5 °C. In January, is the coldest month the average temperature is 12.7 °C. It is the lowest average temperature of the whole year (Fig 1). The soil samples were granules with diameters less than 2- mm, and used after the sieving process. The samples of the airdried soil were used for the physical and chemical parameters of the saline-sodic soil are shown in (Table 1).

139 (Table.1) Initial physical and chemical parameters of saline-sodic soil used for the study

$\frac{\text{pH} (1:2.5 \text{ w; w})}{\text{EC}_c (dS m^1)} \qquad \qquad$		Soil parameter	Saline-sodic soil
$EC_{e} (dS m^{-1}) = 5.43 \pm 0.10$ Total N (gkg <sup>-1</sup> ) 0.02 ± 0.001 Available P (mgkg <sup>-1</sup> ) 1.20 ± 0.07 Available K' (mgkg <sup>-1</sup> ) 7.81 ± 0.44 Total CaCO <sub>3</sub> (%) 18.6 ± 1.35 CEC (Cmol <sup>+</sup> kg <sup>-1</sup> ) 7.56 ± 0.24 Organic Matter (g kg <sup>-1</sup> ) 1.03 ± 0.01 Total DOC (%) 0.012 ± 0.003 ESP (%) 53.1 ± 1.93 C/N Ratio 49.14 ± 2.1 Sand (%) 31.2 ± 0.11 Silt (%) 23.5 ± 0.21 Clay (%) 45.3 ± 0.18 Texture Clay Loam Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> 0.08 ± 0.01 Zn <sup>2+</sup> Nd <sup>+</sup> Mn <sup>2+</sup> 0.11 ± 0.020 Cu <sup>2+</sup> 0.003 ± 0.01 B <sup>+</sup> Nd <sup>+</sup> Cl <sup>-</sup> 192 ± 0.166 Data correspond to means of four replicates ± standard deviation. Nd <sup>+</sup> = not detected		pH (1:2.5 w: w)	$8.84 \pm 0.05$
Total N (gkg <sup>-1</sup> ) 0.02 ± 0.001 Available P (mgkg <sup>-1</sup> ) 1.20 ± 0.07 Available K* (mgkg <sup>-1</sup> ) 78.1 ± 0.44 Total CaCO <sub>3</sub> (%) 18.6 ± 1.35 CEC (CmO <sup>1</sup> kg <sup>-1</sup> ) 7.56 ± 0.24 Organic Matter (g kg <sup>-1</sup> ) 1.03 ± 0.01 Total DOC (%) 0.012 ± 0.003 ESP (%) 53.1 ± 1.93 C/N Ratio 49.14 ± 2.1 Sand (%) 31.2 ± 0.11 Silt (%) 23.5 ± 0.21 Clay (%) 45.3 ± 0.18 Texture Clay Loam Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> 0.08 ± 0.01 Zn <sup>2+</sup> Nd <sup>*</sup> Mn <sup>2+</sup> 0.11 ± 0.020 Cu <sup>2+</sup> 0.003 ± 0.01 B <sup>+</sup> Nd <sup>*</sup> Cl <sup>-</sup> 192 ± 0.166 Data correspond to means of four replicates ± standard deviation. Nd <sup>*</sup> = not detected		$EC_e$ (dS m <sup>-1</sup> )	$5.43 \pm 0.10$
Available P (mgkg <sup>-1</sup> ) Available K <sup>+</sup> (mgkg <sup>-1</sup> ) Total CaCO <sub>3</sub> (%) CEC (Cmol <sup>+</sup> kg <sup>-1</sup> ) Organic Matter (g kg <sup>-1</sup> ) Total DOC (%) ESP (%) C/N Ratio Sand (%) Sand (%) Sand (%) Texture Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Mn <sup>2+</sup> Clay (%) Cir T = 19.2 °C P = 124 mm Micronutrients DTPA extractible $\pm \pm -1.66$ T = 19.2 °C P = 124 mm Micronutrients DTPA extractible (mgkg <sup>-1</sup> ) F * C T = 19.2 °C P = 124 mm Micronutrients DTPA extractible (mgkg <sup>-1</sup> ) F * C Sand Cir Sand Cir Cir Cir Cir Cir Cir Cir Cir		Total N (gkg <sup>-1</sup> )	$0.02 \pm 0.001$
Available K <sup>+</sup> (mgkg <sup>-1</sup> ) Total CaCO <sub>3</sub> (%) CEC (Cmol <sup>+</sup> kg <sup>-1</sup> ) Soil Organic Carbon (g kg <sup>-1</sup> ) Total DOC (%) ESP (%) C/N Ratio Sand (%) Silt (%) Clay (%) Fe <sup>2+</sup> Clay (%) Texture Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Clay Loan Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Nd <sup>*</sup> Mn <sup>2+</sup> Clay Loan B <sup>+</sup> Clay Loan B <sup>+</sup> Clay Loan Clay Loan Te 19.2 °C P = 124 mm 60 F °C Clay Loan F °C T = 19.2 °C P = 124 mm Clay Loan F °C Clay Loan F °C Clay Loan Clay Loa		Available P (mgkg <sup>-1</sup> )	$1.20\pm0.07$
Total CaCO <sub>3</sub> (%) Total CaCO <sub>3</sub> (%) CEC (Cm0 <sup>1</sup> kg <sup>-1</sup> ) Soil Organic Matter (g kg <sup>-1</sup> ) Total DOC (%) Collar (%) Soll DOC (%) C/N Ratio Sand (%) Sand (%) Texture Clay (%) Fe <sup>2+</sup> Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Nd* Mn <sup>2+</sup> Cl <sup>2</sup> Cl <sup>2</sup> Ma* Mn <sup>2+</sup> Cl <sup>2</sup> Cl <sup>2</sup> T = 19.2 °C P = 124 mm Micronutrients down of the collar for the col		Available K <sup>+</sup> (mgkg <sup>-1</sup> )	78.1 ±0.44
CEC (Cmol <sup>+</sup> kg <sup>-1</sup> ) Organic Matter (g kg <sup>-1</sup> ) Soil Organic Carbon (g kg <sup>-1</sup> ) Total DOC (%) ESP (%) C/N Ratio Matter (g kg <sup>-1</sup> ) Total DOC (%) ESP (%) C/N Ratio Sand (%) Sand (%) Sand (%) Sand (%) Sand (%) Clay (%) Texture Clay (%) Fe <sup>2+</sup> Clay Loam Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> O.08 ± 0.01 Zn <sup>2+</sup> Mn <sup>2+</sup> O.03 ± 0.01 B <sup>+</sup> Cl <sup>-</sup> Data correspond to means of four replicates ± standard deviation. Nd <sup>+</sup> = not detected T= 19.2 °C P= 124 mm 60 63 20 Fe <sup>2+</sup> Sand deviation. Nd <sup>+</sup> = not detected Fe <sup>2+</sup> Sand deviation. Nd <sup>+</sup> = not detected		Total CaCO <sub>3</sub> (%)	$18.6 \pm 1.35$
Organic Matter (g kg <sup>-1</sup> ) Soil Organic Carbon (g kg <sup>-1</sup> ) Total DOC (%) ESP (%) C/N Ratio Micromutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Mn <sup>2+</sup> Mn <sup>2+</sup> Cl <sup>-</sup> Data correspond to means of four replicates $\pm$ standard deviation. Nd <sup>+</sup> = not detected T = 19.2 °C P = 124 mm 68 20 Organic Matter (g kg <sup>-1</sup> ) 1.78 $\pm 0.21$ 1.03 $\pm 0.01$ Model A = 1.93 0.01 $\pm 1.93$ C/N Ratio 49.14 $\pm 2.1$ Sand (%) 31.2 $\pm 0.11$ 23.5 $\pm 0.21$ Clay Loam Micromutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> 0.08 $\pm 0.01$ Zn <sup>2+</sup> 0.003 $\pm 0.01$ B <sup>+</sup> Cl <sup>-</sup> 192 $\pm 0.166$ Data correspond to means of four replicates $\pm$ standard deviation. Nd <sup>+</sup> = not detected		CEC (Cmol <sup>+</sup> kg <sup>-1</sup> )	$7.56\pm0.24$
Soil Organic Carbon (g kg <sup>-1</sup> ) Total DOC (%) ESP (%) C/N Ratio $49.14 \pm 2.1$ Sand (%) Silt (%) Clay (%) Texture Clay (%) Fe <sup>2+</sup> $2n^{2+}$ 2n		Organic Matter (g kg <sup>-1</sup> )	$1.78 \pm 0.21$
Total DOC (%) ESP (%) C/N Ratio Sand (%) Silt (%) Clay (%) Texture Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Mn <sup>2+</sup> Cl <sup>2</sup> Cl <sup>2+</sup> Cl <sup>2</sup> Cl <sup>2</sup>		Soil Organic Carbon (g kg <sup>-1</sup> )	$1.03 \pm 0.01$
ESP (%) C/N Ratio Sand (%) Silt (%) Clay (%) Texture Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Mn <sup>2+</sup> Clay Loam Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Mn <sup>2+</sup> Clay Loam Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Mn <sup>2+</sup> Clay Loam Nd* Mn <sup>2+</sup> Clay Loam Nd* Mn <sup>2+</sup> Clay Loam Nd* Mn <sup>2+</sup> Clay Loam Nd* Mn <sup>2+</sup> Clay Loam Nd* Clay Loam Nd* Nd* Clay Loam Nd* Nd* Clay Loam Nd* Nd* Nd* Nd* Nd* Nd* Nd* Nd*		Total DOC (%)	$0.012 \pm 0.003$
C/N Ratio Sand (%) Silt (%) Clay (%) Texture Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Mn <sup>2+</sup> Clu <sup>2+</sup> Clu <sup>2+</sup> Clu <sup>2+</sup> Data correspond to means of four replicates $\pm$ standard deviation. Nd*= not detected T= 19.2 °C P= 124 mm 60 F °C T= 19.2 °C P= 124 mm 60 A 20 F °C 6 30 6 30 7		ESP (%)	$53.1 \pm 1.93$
Sand (%) Silt (%) Clay (%) Texture Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Zn <sup>2+</sup> Mn <sup>2+</sup> Cl <sup>2</sup> Cl <sup>2</sup> Data correspond to means of four replicates $\pm$ standard deviation. Nd*= not detected T= 19.2 °C P= 124 mm 60 F °C 6 30 6 30 7 °C 6 30 6 30 7 °C 8 20 7 °C 8 °C 8 °C 8 20 7 °C 8 °C 8 °C 8 °C 8 20 7 °C 8 °C 8 °C 8 20 7 °C 8 °C 8 20 7 °C 8 °C 8 20 7 °C 8 °C 8 20 7 °C 9 °C		C/N Ratio	$49.14 \pm 2.1$
Silt (%) Clay (%) Texture Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> Zn <sup>2+</sup> Mn <sup>2+</sup> Cl <sup>2+</sup> Cl <sup>2+</sup> Double to construct the form of the		Sand (%)	$31.2 \pm 0.11$
Clay (%) Texture Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> $Zn^{2+}$ $Mn^{2+}$ $Cl^{2+}$ $Cl^{2+}$ $Cl^{2+}$ $Cl^{2+}$ $Cl^{2+}$ $D.08 \pm 0.01$ $Nd^*$ $0.11 \pm 0.020$ $Cu^{2+}$ $0.003 \pm 0.01$ $B^+$ $Cl^{-}$ $192 \pm 0.166$ Data correspond to means of four replicates $\pm$ standard deviation. Nd*= not detected *F *C $T= 19.2 \text{ °C}$ $P= 124 \text{ mm}$ 60  6  30  6 8  20  6  30  6		Silt (%)	$23.5 \pm 0.21$
Texture Clay Loam Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> $Zn^{2+}$ $Mn^{2+}$ $Cl^{2+}$ $Cl^{2+}$ $D.08 \pm 0.01$ $Nd^*$ $0.11 \pm 0.020$ $Cu^{2+}$ $D.003 \pm 0.01$ $B^+$ $Cl^-$ $192 \pm 0.166$ Data correspond to means of four replicates $\pm$ standard deviation. Nd*= not detected *F *C T= 19.2 °C P= 124 mm 60 6 8 20 6 8 20 6 8 20 7 7 7 7 7 7 7 7		Clay (%)	$45.3 \pm 0.18$
Micronutrients DTPA Extractible (mgkg <sup>-1</sup> ) Fe <sup>2+</sup> $Zn^{2+}$ $Mn^{2+}$ $Cu^{2+}$ $Data correspond to means of four replicates \pm standard deviation. Nd*= not detected*F *C T= 19.2 °C P= 124 mm 60 6 30 6 30 6 30 6 30 6 30 6 30 6 3$		Texture	Clay Loam
$Fe^{2+} = 0.08 \pm 0.01$ $Nd^{*} = 0.11 \pm 0.020$ $Cu^{2+} = 0.003 \pm 0.01$ $B^{+} = 0.01 \pm 0.020$ $Cu^{2+} = 0.003 \pm 0.01$ $B^{+} = 0.01 \pm 0.020$ $Data correspond to means of four replicates \pm standard deviation. Nd^{*} = not detected$ $F = C = 0 = 124 \text{ mm} = 0 = 0 = 0 = 0 = 0 = 0 = 0 = 0 = 0 = $		Micronutrients DTPA Extractible (mgkg <sup>-1</sup> )	)
$Zn^{2+}$ $Mn^{2+}$ $Cu^{2+}$ $B^{+}$ $Cl^{-}$ $I = 19.2 \circ C$ $P = 124 \text{ mm}$ $F$ $Cl^{-}$ $Refine the second se$		$\mathrm{Fe}^{2+}$	$0.08 \pm 0.01$
$Mn^{2+} \qquad 0.11 \pm 0.020$ $Cu^{2+} \qquad 0.003 \pm 0.01$ $B^{+} \qquad Nd^{*}$ $Cl^{-} \qquad 192 \pm 0.166$ Data correspond to means of four replicates $\pm$ standard deviation. Nd*= not detected $F = C \qquad T = 19.2 \text{ °C} \qquad P = 124 \text{ mm} \qquad 60 \qquad 6 \qquad 30 \qquad 6 $		$\mathrm{Zn}^{2+}$	Nd*
$Cu^{2+} \qquad 0.003 \pm 0.01$ $B^{+} \qquad Nd^{*}$ $Cl^{-} \qquad 192 \pm 0.166$ Data correspond to means of four replicates $\pm$ standard deviation. Nd*= not detected $F = C \qquad T = 19.2 \text{ °C} \qquad P = 124 \text{ mm} \qquad F = C \qquad 60 \qquad 6 \qquad 30 \qquad 6 \qquad 6 \qquad 6 \qquad 30 \qquad 6 \qquad $		$Mn^{2+}$	$0.11 \pm 0.020$
$B^{+} \\Cl^{-} \\Data correspond to means of four replicates \pm standard deviation. Nd*= not detected$ $F^{+} C^{+} \\Be 30 \\68 20 \\Cl^{-} \\Data correspond to means of four replicates \pm standard deviation. Nd*= not detected$		$\mathrm{Cu}^{2+}$	$0.003 \pm 0.01$
Cl <sup>-</sup> 192 $\pm$ 0.166 Data correspond to means of four replicates $\pm$ standard deviation. Nd <sup>*</sup> = not detected <sup>o</sup> F <sup>o</sup> C T= 19.2 <sup>o</sup> C P= 124 mm <sup>mm</sup> F <sup>o</sup> C 6 30 68 20 40 8 20 8 2		$\mathbf{B}^+$	Nd*
Data correspond to means of four replicates $\pm$ standard deviation. Nd*= not detected <sup>o</sup> F <sup>o</sup> C T= 19.2 <sup>o</sup> C P= 124 mm <sup>mm</sup> F <sup>o</sup> C 6 30 68 20 40 8 20 8 2		Cl	$192 \pm 0.166$
F = C $F = 19.2 \circ C$ $P = 124 \text{ mm}$ F = C 60 6 6 30 40 8 20 40 8 20		Data correspond to means of four replicates ± standard	d deviation. Nd*= not detected
$\begin{array}{c} \mathbf{F} & \mathbf{C} \\ 86 & 30 \\ 68 & 20 \end{array} \xrightarrow{T= 19.2 \circ C} \mathbf{P} = 124 \text{ mm}} \begin{array}{c} mm \\ 60 \\ 60 \\ 6 \\ 8 \\ 20 \end{array} \xrightarrow{F} \begin{array}{c} \circ C \\ 60 \\ 6 \\ 8 \\ 20 \end{array}$			
	۰F	<sup>o</sup> C T= 19.2 °C P= 124 mm F	°C
	86	30 - 60 6	30 -
	68	20 - 40 8	20
	50		10



<sup>(</sup>Fig. 1). Annual precipitation and temperature study area of soil (Climate-Data.org, 2019). https://en.climate-data.org/africa/egypt/alexandria-governorate/alexandria-515/.





#### 153 2.2. Soil Treatments and Azospirillum preparation

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155 Three types of treatments (Azospirillum, compost and spent grain) were used. The 156 Azospirillum bacteria, it was from Faculty of Agriculture, Ain Shams, Cairo Governorate, Egypt. The 157 compost consisted of plant and animal wastes from the national factory for the production of 158 compost, Alexandria, Governorate, Egypt. The spent grain, a by-product from beer industry, was 159 obtained from Al Ahram Beverages Company, Abu Hammad, Al Sharkia Governorate, Egypt 160 (Schema, 1). The characteristics of the treatments were determined according to the standard procedures shown in (Table 2). Azospirillum brasilense (Sp245) was cultured; growth and 161 inoculation were performed as described by (Pii et al., 2015b). Briefly, the Azospirillum bacterium 162 was grown for 4 days in LB medium (10g L<sup>-1</sup> triptone, 5g L<sup>-1</sup> NaCl, 10g L<sup>-1</sup> yeast extract) with 163 164 continuous shaking at 30°C. Bacteria were then centrifuged for 15 min at 4500 x g and washed four times with sterile saline-sodic solution (NaCl 0.85% w/v) after that the Azospirillum bacteria became 165 ready for inoculation with the soil. 166

### 167 2.3. Experimental design and treatments management

168 The experiment was carried out during the summer of 2017 laboratory of the Department of Land and Water Technologies, City of Scientific Research and Technological Applications (SRTA-169 City), and analyses were done between the (SRTA-City) and laboratory of the Department of Soil 170 Science and Soil Ecology, Saint Petersburg State University (SPBSU). The incubation experiment 171 was nine treatments included two levels of spent grain, were referred to 12.8 gkg<sup>-1</sup> soil (S1) and 25.5 172 gkg<sup>-1</sup> soil (S2); two levels of compost, were referred to 23.5 gkg<sup>-1</sup> soil (C1), and 47 gkg<sup>-1</sup> (C2); one 173 level of Azospirillum brasilense inoculation (A1) was 5 % by weight of the soil; a mix of both 174 sources spent grain and compost (C1S1); a mix of both sources with Azospirillum (A1C1) and 175 176 (A1S1); all treatments were used and compared to the control without amendments. All treatments 177 were incorporated with bottle 400g of saline-sodic soil. To let added treatments to affect soil properties, all bottle treatments were incubated under laboratory conditions, without plants, for five 178 months at 28° C. The bottles were distributed in a randomized complete block design with four 179 replications of each treatment. All bottles were watered with tab deionized-water once to field 180 181 capacity of the soil. The most relevant chemical and biological parameters were analyzed every 182 month for five months.







(Schema 1). Schematic beer brewing process and product eco-friendly organic wastes "spent grain"
description of extraction method by (Lordan *et al.*, 2019).

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## (Table, 2). Organic wastes characteristics (oven-dry weight basis).

Soil parameter	Compost	Spent grain
pH (1:5 w:w)	$7.20\pm0.01$	$4.16\pm0.03$
EC (dS/m, 1:5 w:w)	$5.81 \pm 0.21$	$1.45 \pm 0.21$
Organic Matter (g kg <sup>-1</sup> )	332 ±1.23	$750 \pm 0.57$
Total N (%)	$2.10\pm0.32$	$3.12\pm0.68$
Total P (%)	$1.03 \pm 0.52$	$1.86 \pm 0.54$
Total K (%)	$0.57 \pm 0.01$	$1.74 \pm 0.63$
C: N ratio	$9.16 \pm 0.35$	$13.9 \pm 0.12$
Organic carbon (%)	$19.25\pm0.12$	$43.5\pm0.94$
$Fe^{2+}$ (mgkg <sup>-1</sup> )	$960 \pm 2.97$	$1130\pm3.87$
$Zn^{2+}$ (mgkg <sup>-1</sup> )	$220 \pm 5.34$	$368 \pm 2.34$
$Mn^{2+}$ (mgkg <sup>-1</sup> )	$100 \pm 2.14$	$210\pm1.98$
$Cu^+$ (mgkg <sup>-1</sup> )	$61 \pm 1.21$	98 ± 1.54

190

Data correspond to means of four replicates ± standard deviation

191

192 3.4. Sampling

# 193 3.4.1. Soil physicochemical analyses

The total thirty-six samples referred to nine treatments  $\times$  four replications, soil samples it was taken from each bottle after the removal of visible roots and fresh litter material, the composite samples were sieved (2 mm) and then stored at room temperature for less than three months until chemical analyses were performed. Samples were air-dried and ready for measured. The soil pH was measured potentiometrically in a soil : water suspension 1:2.5 w/v (Page, *et al.*, 1982). The electrical





199 conductivity (EC<sub>e</sub>) was measured in saturated paste extracts using an EC meter (Corwin and Yemoto, 200 2017). The total nitrogen (N), available phosphorus (P), and available potassium (K) were measured 201 using (Anderson, *et al.*, 1982). Micronutrients (Fe<sup>2+</sup>, Zn<sup>2+</sup>, Mn<sup>2+</sup>, Cu<sup>2+</sup>, and B<sup>+</sup>) were extracted by 202 DTPA solution as explained by (Soltanpour and Schwab, 1977) then measured with inductively 203 coupled plasma (ICP) atomic emission spectroscopy. Exchangeable sodium percentage (ESP) of soil 204 was determined using the relationship ESP = (exchangeable sodium (Ex Na<sup>+</sup>) / cation exchange 205 capacity (CEC))\*100. Extractable sodium was extracted with 1 M ammonium acetate solution 206 (Normandin *et al.*, 1998). Soil CEC was estimated following the Bower saturation method as outlined 207 by (Richards, 1954).

208

## 209 3.4.2 Biological analyses

210 Dehydrogenase activity was determined by triphenyltetrazolium chloride (TTC) as described by 211 (Casida *et al.*, 1964). Soil urease (S-UE) was determined using methods previously published by 212 (Kandeler and Gerber, 1988). Soil organic carbon (SOC) concentration was determined by 213 oxidization with  $K_2Cr_2O_7$  (Ouyang *et al.*, 2013).

214

#### 215 Statistical analysis

Statistical analysis was carried out using of the SPSS v.16 software (Visauta, 2007). Data were submitted to a normality test before the analysis of variance. When statistical significance was found ( $P \le 0.05$ ), a comparison of the means was carried out by using the Tukey test. Furthermore, a Pearson correlation analysis was carried out to observe the degree of association between some of the studied variables.

- 221
- 222

# **RESULTS AND DISCUSSION**

223 Several factors could control eco-friendly organic wastes decomposition. In the present study, organic types, decomposition condition, and soil properties are the dominant factors that 224 225 control the decomposition processes to reclamation saline-sodic soil. The concentrations of nutrients in the incubated soils were determined as an indication for soil fertility. The chemical and biological 226 227 properties are discussed and presented separately. Chemical properties, including soil acidity (pH), electrical conductivity in saturated paste extract (ECe), exchangeable sodium percent (ESP), cation 228 exchange capacity (CEC), exchangeable sodium (Ex-Na<sup>+</sup>), soil organic carbon (SOC), humic 229 substance (HS), total acidity (TA), available phosphorus (P), available potassium ( $K^+$ ), total nitrogen 230





- 231 (N), micro-nutrients (Fe<sup>2+</sup>, Zn<sup>2+</sup>, Mn<sup>2+</sup>, Cu<sup>2+</sup>, and B<sup>+</sup>), dehydrogenase and urease enzymes in saline-232 sodic soil after 60 and, 150 days of incubation.
- 233 3.1. Chemical properties changed as a soil reclamation indicator after the incubation period
- The soil chemical properties including, pH, ESP, EX-Na<sup>+</sup>, EC<sub>e</sub> and CEC after 60 and 150
- 235 days of incubation, were significantly changed with increasing eco-friendly organic wastes (Table 3).
- 236 Soil soluble and exchangeable salts content statistically significantly decreased with increasing eco-
- 237 friendly organic wastes and *Azospirillum* application (*P*<0.05).
- 238

239 Table (3): Soil salinity indicator in saline-sodic soil after 60 and 150 days of incubation

Treatments	Incubation Period	pН	ECe	Ex-Na <sup>+</sup>	CEC	ESP
	days		dsm <sup>-1</sup>	meq100g-1	Cmol+kg <sup>-1</sup>	%
Initial	Zero time	8.84±0.05	5.43±0.10	4.00±0.18	7.56±0.24	53.12±1.93
Control	60	$8.76 \pm 0.05$	$5.42\pm0.10$	$3.39\pm0.21$	7.93±0.24	$42.78 \pm 1.93$
Control	150	8.94±0.08	$5.48 \pm 0.02$	3.83±0.28	8.26±0.64	46.49±4.16
<b>S</b> 1	60	8.34±0.12	4.11±0.07	3.24±0.29	14.23±0.49	22.76±1.46
51	150	8.03±0.06	3.66±0.23	2.90±0.26	15.03±0.25	19.28±1.57
\$2	60	$8.10\pm0.11$	3.10±0.12	$3.06\pm0.11$	$16.20\pm0.83$	18.91±1.73
52	150	7.83±0.10	3.00±0.17	2.43±0.06	17.80±0.2	13.66±0.2
C1	60	8.51±0.04	5.17±0.13	$3.96 \pm 0.058$	12.33±0.31	30.67±0.33
CI	150	8.60±0.06	$5.42 \pm 0.10$	4.46±0.15	14.43±0.51	30.99±2.01
C2	60	$8.47 \pm 0.04$	5.41±0.21	$4.57 \pm 0.44$	15.67±0.40	$29.34 \pm 2.09$
C2	150	8.58±0.05	5.70±0.53	4.63±0.56	16.03±0.25	28.86±3.14
C181	60	8.17±0.36	4.65±0.32	$5.50\pm0.20$	$14.90\pm0.20$	26.90±0.35
CISI	150	8.06±0.43	4.67±0.02	3.50±0.26	15.93±0.11	21.97±1.76
A 1	60	7.98±0.03	$3.84\pm0.18$	$2.19\pm0.30$	8.83±0.24	$24.89 \pm 4.14$
AI	150	7.86±0.13	3.54±0.35	$1.75 \pm 0.18$	9.26±0.92	19.13±3.72
A 1 S 1	60	7.93±0.06	4.51±0.08	3.06±0.11	15.83±0.29	19.36±0.63
AISI	150	7.89±0.05	4.03±0.07	2.07±0.11	16.12±0.33	12.85±0.49
A1C1	60	$8.24 \pm 0.18$	$4.674 \pm 0.03$	$4.00\pm0.15$	14.25±0.35	$28.07 \pm 0.83$
AICI	150	8.23±0.01	4.485±0.03	3.54±0.36	$14.30 \pm 0.42$	24.72±1.83
I CD.	60	0.275	0.892	2.051	0.827	13.72
L5D0.05	150	0.278	0.413	0.475	0.801	4.180

 $\begin{array}{ll} 240 & \mbox{Data correspond to means of four replicates $\pm$ standard deviation. LSD_{0.05}$= least significant difference (LSD) test \\ 241 & \mbox{at } (p \le 0.05). \end{array}$ 

242

# 243 3.1.1. The soil pH changed after the incubation period

The influences of the eco-friendly organic wastes incubation with Azospirillum inoculation 244 245 on the decrease soil pH compared to initial condition and control treatment followed the order: S2 (spent grain 25.5 gkg<sup>-1</sup>) > A1 (5% of soil weight) > A1S1 (Mix A1 and S1) > S1 (spent grain 12.8 246  $gkg^{-1}$  > C1S1 (Mix C1 and S1) > C2 (47  $gkg^{-1}$ ) > C1 (Compost 23.5  $gkg^{-1}$ ). Shown (Fig 2) soil pH 247 after 60 days of incubation was higher in the soil initial condition compared to the spent grain (S2) 248 249 and Azospirillum (A1), treatments were 8.10, 7.93 and 7.98, for the treatments S2, A1S1, and A1 respectively, (P<0.05). While the corresponding values after 150 days of incubation were 7.83, 7.86 250 and 7.89, for the S2, A1 and A1S1 respectively, (P<0.05). The inherent soil pH values after 150 days 251





were between 7.8 and 7.89 are ideal for P-availability, while pH values below 5.5 and between 7.5 252 253 and 8.5 limits P-availability to plants due to fixation by aluminum, iron, or calcium (Rashad et al., 254 2016). The S2 treatment affected positively the soil pH greater than compost treatments and control. 255 Accordingly, this reduction in the pH values might be due to increase of the groups of carboxyl and 256 phenol as a result of decomposing of eco-friendly organic wastes and the reduction of bicarbonate in 257 the soil. (Hafez et al., 2019) reported that a decrease of pH might be due to the addition of spent grain only or with Azospirillum inoculation in calcareous and sandy soils reduces soil pH sharply. The 258 difference in the pH values between the S2 and the A1S1 was a highly significant camper to initial 259 and control. The remediation of saline-sodic soils by organic matter sources has long been known to 260261 facilitate the reclamation of saline-sodic soils (Sastre-Conde et al., 2015).



262

Figure (2): Effect of *Azospirillum* and eco-friendly organic wastes on pH in saline-sodic soil after 60 and 150 days of incubation. Data points and error bars represent means and standard errors (n = 4).

## 265 3.1.2. Soil ESP indicator

266 The soil ESP was decreased with the spent grain and Azospirillum increasing (P < 0.05). The soil ESP was statistically significantly higher in the inoculation Azospirillum with spent grain in 267 268 saline-sodic soil than that in the compost and control treatments (P<0.05). In the present study, the middle and high salinities exhibited greater salt content and ESP, and less Table (3) and Figure (3). 269 270 The ESP after 60 days of incubation in the saline-sodic soil was 53.10, 19.35, 20.91 and 23.64 % for the initial condition, A1S1, S2, and S1, respectively. The corresponding percentages for the 150 days 271 272 was similar to the trend of pH were 53.10, 12.85, 13.66 and 19.28 % for the initial condition, A1S1, S2, and S1, respectively. The addition of spent grain with Azospirillium (A1S1) to the saline-sodic 273 soil contributed a statistically significantly lower amount of salt to the soil. The spent grain decreased 274 275 the amount of exchangeable Na<sup>+</sup> and salinity than the compost and initial condition, this reducing for





ESP due to mixing Azospirillium with spent grain enhancement of the soil activity of carbon 276 degrading extracellular enzymes and microbial decomposition rates. There was a significantly 277 278 positive correlation between soil salt content and eco-friendly organic wastes amendments. Soil 279 microbes are essential in soil carbon mineralization and accumulation, which determines soil fertility. 280The Azospirillium increased microbial biomass carbon which decreased the exchangeable 281 Na<sup>+</sup> in soil solution, this result was positively correlated with organic wastes incubation in salinesodic soil. The addition of organic matter played the very important role for decreased soil salinity 282 and enhancement the soil microbial biomass, this result agreement with (Zhang et al., 2015, 2019; 283 Zhen et al., 2014) demonstrated that the low soil microbial carbon was attributed to the low soil 284 organic carbon that resulted from little carbon input into the soils for the scarcity of vegetation under 285 high salt condition. (Mahmoodabadi et al., 2013; Sastre-Conde et al., 2015) attributed the decrease of 286 soil EC, SAR, and ESP by organic wastes and chemical amendments under saline-sodic soil. (Meena 287 et al., 2019) further demonstrated that lower organic matter in saline soil is more likely to be due to 288 the decreased organic sources than the increased decomposition rate. 289



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291 292

293

Figure (3): Effect of *Azospirillum* and eco-friendly organic wastes on ESP in saline-sodic soil after 60 and 150 days of incubation. Data points and error bars represent means and standard errors (n = 4).

# 294 3.1.3. Soil exchangeable Na<sup>+</sup> and electrical conductivity

295 The soil Ex-Na<sup>+</sup> was statistically significantly higher in the inoculation Azospirillum with 296 spent grain in saline-sodic soil than that in the other treatments (P<0.05). In the present study, the saline-sodic soil before amendments incubation was contained high content of the Ex-Na<sup>+</sup> and ESP, 297 298 and less organic matter compared to the after amendments incubation (Table 3). The effect of spent grain levels and Azospirillum treatments on Ex-Na<sup>+</sup> concentration were consistently higher (P< 0.05) 299 300 than compost levels, control and the initial (Table 3). The general influences of the Azospirillum inoculation with eco-friendly organic wastes incubation on the decrease in the soil Ex-Na<sup>+</sup> 301 302 concentration compared to initial condition and control treatment, followed the order: A1 >A1S1 >S2 >S1 >C1S1 >control >initial >C2 >C1. Shown (Table 3), the soil Ex-Na<sup>+</sup> for compost treatments 303





after 60 days of incubation were 5.0, 3.80 and 4.0 meq100g<sup>-1</sup>, for the C2, C1 and A1C1 respectively, 304 (P<0.05). While the corresponding values after 150 days of incubation was higher in the compost 305 306 only and compost with Azospirillium treatments compared to the initial condition and control, were 4.6, 4.4 and 3.54 meq100g<sup>-1</sup> for the treatments C2, C1 and A1C1 respectively. In the control and 307 initial Ex-Na<sup>+</sup> ranged between 3.4 and 4 meq100g<sup>-1</sup> after 150 days of incubation, while in A1, after 308 150 days Ex-Na<sup>+</sup> was 1.75 meq100g<sup>-1</sup>. This confirms that soil salinization was effectively maintained 309 through the control treatment irrigation with saline water, low rainfall since Ex-Na<sup>+</sup> decreased in A1 310 from 4 in initial to 1.75 meq100g<sup>-1</sup> for spent grain and Azospirillium treatments. The relative 311 reduction in Ex-Na<sup>+</sup> concentration after 150 days corresponded to 56.25 and 48.25% for A1 and 312 A1S1 respectively, compared to the initial condition. Obviously, as the application levels of compost 313 314 increased, the ECe and Ex-Na<sup>+</sup> concentrations increased (Table 3). There were significant differences among the treatments in the Ex-Na<sup>+</sup> concentration. 315

316 On the other hand, the higher concentrations of ECe and Ex-Na<sup>+</sup> can lead to deterioration of soil physical properties. The compost treatments was negative a contributed statistically significantly 317 higher amount of ECe and Ex-Na<sup>+</sup> to the soil compared the control. I think this result due to the 318 compost treatments is contained very high salts content before adding to soil, resulting in increased 319 Na<sup>+</sup> concentration in the compost residues. Therefore, the application of pig compost could increase 320 soil salinity in areas of low precipitation and drainage where the salts are not leached through the soil 321 profile or on borderline saline soils. Therefore, the characterization of organic wastes is mandatory 322 323 before adding to soil.

On the other hand the Azospirillum with spent grain treatments decreased amount of ECe and 324 325 Ex-Na<sup>+</sup> than the compost and initial condition, this reducing for the Ex-Na<sup>+</sup> due to spent grain with Azospirillum enhancement of the soil microbial activity of carbon-degrading extracellular enzymes, 326 increased humic substance, chelating the sodium in soil to sodium humate form and enhancement of 327 CEC under A1S1 and A1 treatments in saline-sodic soil incubated. Similar results were reported by 328 329 (Mahmoodabadi et al., 2013; Rojas-Tapias et al., 2012; Shrivastava and Kumar, 2015; Zhang et al., 2015) who reported that the applied organic wastes and PGPB, depending on their chemical 330 331 composition might have a positive impact on soil properties, especially Na<sup>+</sup> and EC. (Ashraf et al., 2004), who found that inoculation with exopolysaccharide- producing bacteria could restrict Na<sup>+</sup> 332 influx into roots. Further, (Zhang et al., 2008) reported that inoculation with Bacillus bacteria could 333 mediate the level of salt tolerance in soil. 334

### 335 3.1.4. Cation Exchangeable Sodium Percentage (CEC)

336 A progressive increase in CEC was observed in all treatments due to the accumulation of 337  $Ca^{2+}$  and  $Mg^{2+}$  release with the eco-friendly organic wastes amendments. In particular, CEC





increased significantly during the spent grain and Azospirillum treatments (P < 0.05), when it reached 338 15.5 and 15.3 Cmol+kg<sup>-1</sup> in A1S1 and S2 (Table 3) after 60 days of incubation. The A1S1treatment 339 increased CEC and decreased Ex-Na<sup>+</sup> led to a significant reduction (P < 0.05) for the ESP indicator 340 341 (Table 3). On the other hand, the corresponding percent for the 150 days was similar to the trend of SOC were 7.50, 17.80, 16.12 and 16.01 Cmol+kg<sup>-1</sup> for the initial condition, S2, A1S1, and C2, 342 respectively. As a result, in soil, no significant differences were observed between Azospirillum, eco-343 344 friendly organic wastes, and the control. The effect of Azospirillum inoculation with spent grain on the CEC was confirmed when comparing to CEC of the different treatments of compost, (Fig 4). 345 CEC increased by 137.3, 115 and 113.7% in S21, A1S, and C2 respectively, after 150 days of soil 346 incubation. Generally, the reported concentrations in organically treated soil with application rate of 347 Azospirillum, spent grain and compost are safe and sufficient for macronutrient consumption. This is 348 349 results in agreement with other researchers (Shrivastava and Kumar, 2015; Trivedi et al., 2017).

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353 354

- Figure (4): Effect of *Azospirillum* and eco-friendly organic wastes on CEC in saline-sodic soil after 60 and 150 days of incubation. Data points and error bars represent means and standard errors (n = 4).
- 355

# 356 3.2. Soil biological parameters after incubation period

#### 357 **3.2.1.** Dehydrogenase and urease activities

358 The soil biological parameters dehydrogenase and urease activities showed in Figure (5) and figure (6) revealed that there were significant differences in dehydrogenase and urease 359 activities in saline-sodic soil between all treatments, and also significant differences were recorded 360 361 when using different levels of eco-friendly organic wastes and Azospirillum concentrations followed the order A1S1 $\ge$  S2  $\ge$  A1>C1S1 > C2 > A1C1  $\ge$  S1 > C1 > control > initial. Also, the differences in 362 dehydrogenase and urease activities were highly significant (P<0.05) when combined with different 363 364 levels of spent grain with Azospirillum or only spent grain, compared with different levels of compost and control. This is results in agreement with other researchers (Liang et al., 2014; Meena et al., 365





366 2019), in soils treated with farmyard manure; dehydrogenase activity is increasing. The high organic matter content enhanced the production of microbial activity as a result of soil organic carbon during 367 the mineralization process of organic matter. Generally, the mineralized organic carbon increased as 368 the application rate of spent grain with Azospirillum increased. (Liang et al., 2014) reported that 369 application of organic amendments increased microbial activity and urease activities in all particle-370 size fractions as compared to those in control. Our results are in the same direction with the findings 371 of (Rojas-Tapias et al., 2012; Scherer et al., 2011; Shi et al., 2019) which proposed that the 372 treatments with high organic amendments might exhibit the greatest microbial activity and 373 dehydrogenase activity. The soil enzyme activities play a very important role in nutrients recycling, 374 making them accessible to plants and microorganisms (Ai et al., 2015). 375



377

Figure (5): Effect of *Azospirillum* and eco-friendly organic wastes on Dehydrogenase enzyme ( $\mu$ g TPF ml<sup>-1</sup>) in saline-sodic soil after 60 and 150 days of incubation. Data points and error bars represent means and standard errors (n = 4).



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Figure (6): Effect of *Azospirillum* and eco-friendly organic wastes on Urease enzyme in saline-sodic soil after 60 and 150 days of incubation. Data points and error bars represent means and standard errors (n = 4).





#### 384 3.2.2. Soil organic carbon

385 The soil organic carbon (SOC) values are shown in Table (4) and Figure (7). The SOC percentages after 60 days of incubation were 0.1, 0.081, 0.64, 1.16, 0.55, 1.0, 0.98, 0.056, 0.58 and 386 0.51 for initial condition, control, S1, S2, C1, C2, C1S1, A1, A1S1 and A1C1, respectively. There 387 388 were significant differences among the treatments. The trend of SOC after 60 days was similar to the trend of SOC after 150 days of incubation, where the SOC percentages were 0.1, 0.0712, 0.54, 1.05, 389 0.47, 0.88, 0.83, 0.26, 0.50 and 0.39 for initial condition, control, S1, S2, C1, C2, C1S1, A1, A1S1 390 and A1C1, respectively. The spent grain (S2) possessed the highest values of SOC while the A1 and 391 control showed the lowest value. The spent grain amendments increased the carbon contents in 392 saline-sodic soil up to 12 gkg<sup>-1</sup>. The use of organic wastes with Azospirillum increased saline-sodic 393 394 soil organic carbon contents. On the other hand, the Azospirillum bacteria increased the SOC after 395 150 days greeter than after 60 days, this result Azospirillum increased of decomposition rate was 396 highly significant after 150 days compared with 60 days of incubation. The large increase of SOC and TN amounts with organic amendments application might be attributed to the increased 397 398 particulate organic matter derived from manure since the organic material added to the soil was initially located in the coarser fraction (Liang et al., 2014; Zhang et al., 2015). The high organic 399 400 matter content enhanced the production of soil organic carbon as a result of micro-organisms activity 401 during the mineralization process of organic matter. Generally, the mineralized organic carbon 402 increased as the application rate of spent grain and compost increased. The presented results of SOC are in agreement with the results presented by (Luo et al., 2018; Nadeem et al., 2014; Nassar et al., 403 2014; Scherer et al., 2011; Zhang et al., 2015) 404



407Figure (7): Effect of Azospirillum and eco-friendly organic wastes on SOC in saline-sodic soil after 60408and 150 days of incubation. Data correspond to means of four replicates  $\pm$  standard error. Different409letters indicate statistically significant differences ( $p \le 0.05$ ) among treatments.



410



Treatments	Incubation Period	Total N	Ava P	Ava K	SOC
	Day	%	m	gkg-1	%
Initial	Zero time	0.0021	1.2	78.0	0.1
Control	60	0.0030	1.95	86.31	0.096
Control	150	0.0047	2.06	92.66	0.071
<b>S</b> 1	60	0.0280	3.73	158.47	0.597
51	150	0.0950	5.72	179.33	0.547
52	60	0.0633	6.81	268.06	1.175
32	150	0.1482	14.28	284.33	1.052
01	60	0.0160	2.66	121.71	0.576
CI	150	0.0840	3.72	145.20	0.471
<b>C</b> 2	60	0.0360	5.45	206.24	0.968
C2	150	0.1090	8.45	237.66	0.885
C161	60	0.0567	7.22	200.00	1.169
CISI	150	0.1250	10.00	237.66	0.835
A 1	60	0.0460	8.94	88.366	0.055
AI	150	0.1437	13.13	117.66	0.267
A 1 C 1	60	0.0583	8.81	166.14	0.576
AISI	150	0.1717	9.63	195.00	0.504
4101	60	0.0324	5.22	139.20	0.501
AICI	150	0.099	7.15	180.50	0.396
I CD	60	0.005	0.452	11.0	0.208
L5D0.05	150	0.025	0.761	17.74	0.065

#### Table (4): Soil N, P, K and SOC in saline-sodic soil after 60 and 150 days of incubation

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## 413 3.3. Soil fertility indicator after incubation periods

## 414 3.3.1. N, P, and K<sup>+</sup> concentration in saline-sodic soil

415 The effects of different amendments on soil N are shown in Table (4). Azospirillum 416 inoculation not only alleviated the inhibitory effect of salinity on soluble and total macro-nutrients 417 but also induced a marked and progressive increase in N, P and K concentrations. Figure (8) show total N concentrations. Total nitrogen is the sum of nitrate (NO<sub>3</sub><sup>-</sup>), nitrite (NO<sub>2</sub>), organic nitrogen and 418 ammonia (all expressed as N). The total N percentages for the saline-sodic soil after 60 days of 419 420 incubation were 0.0021, 0.003, 0.028, 0.063, 0.016, 0.036, 0.056, 0.046, 0.058 and 0.032% for the initial condition, control, S1, S2, C1, C2, S1C1, A1, A1S1 and A1C1 treatments, respectively. The 421 422 corresponding percentages for the saline-sodic soil after 150 days of incubation were 0.0021, 0.0047, 0.095, 0.148, 0.084, 0.109, 0.125, 0.143, 0.171 and 0.099%. The spent grain with Azospirillum 423 inoculation was a good supplier for the total nitrogen at both incubation periods in comparison to the 424 initial total N. 425

The compost treatment supplied a lower total N in comparison to the *Azospirillum* and spent grain. This is due to the high initial nitrogen percentage in spent grain and enhancements decomposition rate and mineralization process by *Azospirillum* bacteria. The total N was greater after 150 days of incubation than after 60 days of incubation in saline-sodic soil. Is in accordance with the report of (Liang *et al.*, 2014; Sastre-Conde *et al.*, 2015; Yu *et al.*, 2019; Zhang *et al.*, 2015),





that a majority part of organic fertilizer is released to the soil N, P and K<sup>+</sup> after application of agricultural waste compost and bio-fertilizer in saline-sodic soils. Accordingly, the spent grain with *Azospirillum* used in the present study can supply a growing plant with plentiful of nitrogen after 60 and 150 days of incubation and this period was enough for decomposition organic matter and mineralization of nutrients. Soil microbial communities regulate principal ecosystem processes such as soil nutrient cycling and organic matter formation and decomposition (Sant'Anna *et al.*, 2018; Trivedi *et al.*, 2017).

The soil available P and K<sup>+</sup> values are shown in Table (4) and Fig (9) were significantly 438 increased by eco-friendly organic wastes with Azospirillum treatments compared to the initial 439 condition. The trend of available P and K after 60 days was similar to the trend of available P and K 440 441 after 150 days of incubation, the available P and K, concentrations followed the order  $S_2 > A_1 >$ A1S1 > C1S1 > C2 > S1 > C1 > A1C1 > control > initial respectively. The spent grain (S2) possessed442 443 the highest values of P and K while the A1C1 and control showed the lowest value. The available P and K were significantly (P < 0.05) increased by spent grain (S2) treatment compared to control 444 treatment after 150 days 12 and 3.64 folds increase in P and K respectively. The spent grain 445 amendments increased the available P and K contents in saline-sodic soil up to 12 mgkg<sup>-1</sup> in S2 and 446 A1 respectively. 447

448 The spent grain application rates with Azospirillum were superior compared to the same rate of compost for soil P and K. The high rate of spent grain (S2) significantly produced larger increases 449 in soil P than in the other organic wastes amendment, while A1 was statistically similar to S2 and 450 both significantly increased soil P compared to other organic wastes amendments shown Figure (9). 451 452 Generally soil N, P and K increased as the application rate of Azospirillum or spent grain increased, and all of the organic treatments were effective at adding N, P and K to saline-sodic soil. 453 Furthermore, spent grain and Azospirillum should be applied before planting to give sufficient time 454 for natural oxidation of organic wastes materials, which in turn enhances the soil available nutrients. 455 456 Similar to the results of the present study, incubation of some organic sources resulted high total N. Our results also concur with those of (Barra et al., 2019; Ouyang et al., 2013; Trivedi et al., 2017) 457 458 who found that the application of composted poultry manure, organic fertilizer, and bio-fertilizers 459 inoculation enhances the benefit of P-fertilization led to the increase of available P in saline-sodic soils. This increase in available P is due to the reduction in pH as a result of organic matter 460 decomposition. Furthermore, the application of bio-fertilizer or FYM significantly increased total N, 461 available P and K for saline soils. (Liang et al., 2014; Yu et al., 2019). 462









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Figure (8): Effect of *Azospirillum* and eco-friendly organic wastes on total (N) in saline-sodic soil after 60 and 150 days of incubation. Data correspond to means of four replicates  $\pm$  standard error. Different letters indicate statistically significant differences (p  $\leq$  0.05) among treatments.





Figure (9): Effect of *Azospirillum* and eco-friendly organic wastes on available (P) in saline-sodic soil after 60 and 150 days of incubation. Data points and error bars represent means and standard errors (n = 4)

<sup>469</sup> 470

	471	Table (5): Soil	micronutrien	ts in saline	e-sodic soil	after 60 and	d 150 days	of incubation
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Treatments	Incubation Period	Fe <sup>2+</sup>	Zn <sup>2+</sup>	Mn <sup>2+</sup>	Cu <sup>2+</sup>	B+
	Day			mgkg-1		
Initial	Zero time	$0.08 \pm 0.01$	Nd*	0.11±0.02	0.003±0.01	Nd*
Control	60	$0.74\pm0.01$	$0.332\pm0.06$	$0.619 \pm 0.07$	0.093±0.01	$0.058 \pm 0.01$
Control	150	$1.27 \pm 0.08$	0.477±0.04	$1.089 \pm 0.10$	0.264±0.02	0.081±0.009
<b>S</b> 1	60	3.33±0.19	0.925±0.03	2.049±0.09	$0.742 \pm 0.08$	$0.445 \pm 0.021$
51	150	4.27±0.32	$1.335 \pm 0.13$	2.625±0.05	$1.020 \pm 0.13$	$0.768 \pm 0.151$
\$2	60	8.01±0.41	2.633±0.10	5.989±0.66	$1.344\pm0.11$	0.716±0.012
52	150	9.15±0.44	3.273±0.13	7.461±0.72	1.776±0.02	$1.110 \pm 0.18$
C1	60	$2.62\pm0.05$	0.753±0.03	1.657±0.13	$0.403 \pm 0.02$	$0.378 \pm 0.04$
CI	150	$3.12 \pm 0.06$	$1.059 \pm 0.14$	$2.334 \pm 0.22$	0.501±0.08	$0.510 \pm 0.02$
C2	60	$4.40\pm0.14$	$1.439\pm0.11$	3.522±0.05	0.851±0.02	$0.574 \pm 0.02$
C2	150	5.16±0.023	$2.178 \pm 0.07$	4.719±0.19	1.131±0.05	$0.858 \pm 0.05$
C181	60	$5.96 \pm 0.55$	1.473±0.07	$4.602 \pm 1.64$	0.868±0.30	0.591±0.18
C151	150	6.64±0.65	$2.010 \pm 0.18$	6.159±0.27	$1.329 \pm 0.10$	$1.143 \pm 0.14$
A 1	60	4.03±0.03	1.113±0.13	2.595±0.07	0.711±0.11	$0.350 \pm 0.02$
711	150	$4.68 \pm 0.21$	$1.488 \pm 0.12$	$3.183 \pm 0.18$	0.993±0.16	0.501±0.07
A 1 S 1	60	12.58±0.40	2.904±0.04	5.956±0.11	1.531±0.17	0.916±0.04
71151	150	13.80±0.36	4.125±0.39	7.600±0.37	1.945±0.24	1.275±0.36
A1C1	60	2.38±0.11	$0.865 \pm 0.09$	$1.789\pm0.11$	$0.467 \pm 0.05$	$0.439 \pm 0.05$
AICI	150	3.81±0.28	$1.308 \pm 0.35$	2.193±0.16	0.660±0.04	$0.534 \pm 0.08$
I SDa of	60	0.484	0.140	1.022	0.228	0.114
1.500.05	150	0.585	0.361	0.541	0.207	0.271

472 Data correspond to means of four replicates  $\pm$  standard deviation. LSD<sub>0.05</sub>= least significant difference (LSD) test

473 at  $(p \le 0.05)$ . Nd<sup>\*</sup>= not detected





#### 3.3.2. Soil micro-nutrients ( $Fe^{2+}$ , $Zn^{2+}$ , $Mn^{2+}$ , $Cu^{2+}$ and $B^+$ ) 475

476 The soil amendments were found significantly impact on micro-nutrients. Table (5) shows that  $Fe^{2+}$ ,  $Zn^{2+}$ ,  $Mn^{2+}$ ,  $Cu^{2+}$ , and  $B^+$  concentrations after 60 days of incubation had similar trend  $Fe^{2+}$ , 477 Zn<sup>2+</sup>, Mn<sup>2+</sup>, Cu<sup>2+</sup> and B<sup>+</sup> concentrations after 150 days of incubation. Fe<sup>2+</sup>, Zn<sup>2+</sup>, Mn<sup>2+</sup>, Cu<sup>2+</sup> and B<sup>+</sup> 478 concentrations followed the order A1S1 > S2 > C1S1 > C2 > A1 > S1 > A1C1 > C1 > control > initial.479 480 The micronutrients, the largest increases were observed with A1S1 treatment while the lowest increases were observed with C1 and control treatment after 60 and 150 days of saline-sodic soil 481 incubated. Available micronutrients were significantly (P < 0.05) increased by Azospirillum with 482 spent grain (A1S1) treatment compared to control treatment 18.9, 12.42, 12.66, 20.86 and 21.90 folds 483 increase in Fe<sup>2+</sup>, Zn<sup>2+</sup>, Mn<sup>2+</sup>, Cu<sup>2+</sup>, and B<sup>+</sup>, respectively. The Azospirillum and spent grain application 484 rates (A1S1 and S2) were superior compared to the same rate of compost for Fe<sup>2+</sup>, Zn<sup>2+</sup>, and Mn<sup>2+</sup> 485 (Fig 10). The superiority of the Azospirillum with spent grain was due to its effect on lowering soil 486 487 pH as well as solubility and chelation effect of organic wastes in saline-sodic soil (Pii et al., 2015a). Accordingly, the application of A1S1 to enrich soil with micronutrients is highly recommended in 488 489 saline-sodic soils. The mineralization of eco-friendly organic wastes compounds leads to decrease pH in saline-sodic soil, which increases micronutrient availability in the soil. The presented results are in 490 agreement with the results presented by (Rashad et al., 2016; Wang and Cai, 2006; Zhang et al., 491 2019) who reported that the available micro-nutrients are increased by using agro-industrial organic 492 493 wastes and microbial community.





Figure (10): Effect of Azospirillum and eco-friendly organic wastes on micro-nutrients (Fe<sup>2+</sup>, Zn<sup>2+</sup>, and 496 Mn<sup>2+</sup>) in saline-sodic soil after 60 and 150 days of incubation. Data points and error bars represent means 497 and standard errors (n = 4). 498

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#### 502 CONCLUSION

503 The results of the present study confirm our hypotheses that (i) Application of spent grain 504 doses with Azospirillum inoculation reduced the ESP, increased the beneficial microbes and the 505 activity of enzymes from decomposition of the eco-friendly organic wastes and (ii) Soil fertility 506 increased by improvement of macro and micro-nutrients in saline-sodic soil after 60 and 150 days of incubation. The results also showed that the spent grain and Azospirillum treatments enhanced the 507 production of soil organic carbon, the activity of enzymes, soil macro and micro-nutrients which 508 decreased soil salinity in saline-sodic soil as a result of micro-organisms activity during the 509 510 mineralization process of spent grain with Azospirillum treatments.

In the saline-sodic soil, the concentration of monovalent total N, available P, and available 511  $K^+$  also Fe<sup>2+</sup>, Zn<sup>2+</sup>, Mn<sup>2+</sup>, Cu<sup>2+</sup>, and B<sup>+</sup> were affected significantly by the application of Azospirillum 512 with spent grain under controlled laboratory conditions. In the final application of Azospirillum in 513 514 saline-sodic soil has a negligible effect on the final SOC content but it has a strong effect on soil 515 dehydrogenase, urease, pH, EC, ESP, and Ex-Na<sup>+</sup>. At the end of experiments, monovalent cations 516 concentrations, as well as soil EC and ESP, were smaller after 150 days than 60 days of incubation. Finally, the soil salinity indicator decreased significantly, even for eco-friendly organic wastes with 517 518 Azospirillum, after 150 days of incubation. Results suggested that the applied spent grain with 519 Azospirillum, depending on their biological and chemical properties might have a positive effect on soil properties, especially in the absence of compost. 520

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## 530 **REFERENCES**

- Ahmad, P., Azooz, M.M., and Prasad, M.N.V.: Salt Stress in Plants, Salt Stress in Plants:
  Signalling, Omics and Adaptations. Springer New York, New York, NY. https://doi.org/10.1007/978-1-4614-6108-1, 2013
- Ai, C., Liang, G., Sun, J., He, P., Tang, S., Yang, S., Zhou, W., and Wang, X.: The alleviation of
  acid soil stress in rice by inorganic or organic ameliorants is associated with changes in soil
  enzyme activity and microbial community composition. Biol. Fertil. Soils 51, 465–477.
  https://doi.org/10.1007/s00374-015-0994-3, 2015
- 538 Anderson, J.P.E., Page, A.L., Miller, R.H. and Keeney, D.R.: Methods of Soil Analysis and Soil





539 Respiration. https://doi.org/10.2134/agronmonogr9.2.2ed.c41, 1982

539	Respiration. https://doi.org/10.2134/agronmonogr9.2.2ed.c41, 1982			
540	Ashraf, M., Hasnain, S., Berge, O., and Mahmood, T.: Inoculating wheat seedlings with			
541	exopolysaccharide-producing bacteria restricts sodium uptake and stimulates plant growth			
542	under salt stress. Biol. Fertil. Soils 40, 157-162. https://doi.org/10.1007/s00374-004-0766-y,			
543	2004			
544	Bano, A., and Fatima, M.: Salt tolerance in Zea mays (L). following inoculation with Rhizobium			
545	and Pseudomonas. Biol. Fertil. Soils 45, 405–413. https://doi.org/10.1007/s00374-008-0344-9,			
546	2009			
547	Barra, P.J., Pontigo, S., Delgado, M., Parra-Almuna, L., Duran, P., Valentine, A.J., Jorquera, M.A.,			
548	and Mora, M. de la L.: Phosphobacteria inoculation enhances the benefit of P-fertilization on			
549	Lolium perenne in soils contrasting in P-availability. Soil Biol. Biochem. 136, 107516.			
550	https://doi.org/10.1016/j.soilbio.2019.06.012, 2019			
551	Casida, L.E., Klein, D.A., and Santoro, T.: Soil Dehydrogenase Activity. Soil Sci. 98, 371-376.			
552	https://doi.org/10.1097/00010694-196412000-00004, 1964			
553	Climate-Data.org.: climate-data.org. https://doi.org/https://en.climate-			
554	data.org/africa/egypt/alexandria-governorate/alexandria-515/., 2019			
555	Corwin, D.L., and Yemoto, K.: Salinity: Electrical Conductivity and Total Dissolved Solids.			
556	Methods Soil Anal. 2, 0. https://doi.org/10.2136/msa2015.0039, 2017			
557	Dodd, K., Guppy, C.N., Lockwood, P. V., and Rochester, I.J.: The effect of sodicity on cotton:			
558	Does soil chemistry or soil physical condition have the greater role? Crop Pasture Sci. 64, 806.			
559	https://doi.org/10.1071/CP13078, 2013			
560	Hafez, M., Popov, A.I., and Rashad, M.: Influence of Agro-industrial Wastes and Azospirillum on			
561	Nutrients Status and Grain Yield under Corn Plant Growth in Arid Regions. Biosci. Res. 16,			
562	2119-2130. https://doi.org/www.isisn.org Bioscience, 2019			
563	Hayat, R., Ali, S., Amara, U., Khalid, R., and Ahmed, I.: Soil beneficial bacteria and their role in			
564	plant growth promotion: a review. Ann. Microbiol. 60, 579-598.			
565	https://doi.org/10.1007/s13213-010-0117-1, 2010			
566	Hu, Y., and Schmidhalter, U.: Drought and salinity: A comparison of their effects on mineral			
567	nutrition of plants. J. Plant Nutr. Soil Sci. 168, 541-549.			
568	https://doi.org/10.1002/jpln.200420516, 2005			
569	Kandeler, E., and Gerber, H.: Short-term assay of soil urease activity using colorimetric			
570	determination of ammonium. Biol. Fertil. Soils 6, 68–72. https://doi.org/10.1007/BF00257924,			
571	1988			
572	Karagöz, K., Ateş, F., Karagöz, H., Kotan, R., and Çakmakçi, R.: Characterization of plant growth-			
573	promoting traits of bacteria isolated from the rhizosphere of grapevine grown in alkaline and			
574	acidic soils. Eur. J. Soil Biol. 50, 144-150. https://doi.org/10.1016/j.ejsobi.2012.01.007, 2012			
575	Liang, Q., Chen, H., Gong, Y., Yang, H., Fan, M., and Kuzyakov, Y.: Effects of 15 years of manure			
576	and mineral fertilizers on enzyme activities in particle-size fractions in a North China Plain			
577	soil. Eur. J. Soil Biol. 60, 112-119. https://doi.org/10.1016/j.ejsobi.2013.11.009, 2014			
578	Lordan, R., O'Keeffe, E., Tsoupras, A., and Zabetakis, I.: Total, Neutral, and Polar Lipids of			
579	Brewing Ingredients, By-Products and Beer: Evaluation of Antithrombotic Activities. Foods 8,			
580	171. https://doi.org/10.3390/foods8050171, 2019			

581 Luo, S., Wang, S., Tian, L., Shi, S., Xu, S., Yang, F., Li, X., Wang, Z., and Tian, C.: Aggregate-582 related changes in soil microbial communities under different ameliorant applications in saline-





583	sodic soils. Geoderma 329, 108-117. https://doi.org/10.1016/j.geoderma.2018.05.023, 2018
584	Mader, P.: Soil Fertility and Biodiversity in Organic Farming. Science (80 ). 296, 1694-1697.
585	https://doi.org/10.1126/science.1071148, 2002
586	Mahmoodabadi, M., Yazdanpanah, N., Sinobas, L.R., Pazira, E., and Neshat, A.: Reclamation of
587	calcareous saline sodic soil with different amendments (I): Redistribution of soluble cations
588	within the soil profile. Agric. Water Manag. 120, 30-38.
589	https://doi.org/10.1016/j.agwat.2012.08.018, 2013
590	Meena, M.D., Yadav, R.K., Narjary, B., Yadav, G., Jat, H.S., Sheoran, P., Meena, M.K., Antil,
591	R.S., Meena, B.L., Singh, H.V., Singh Meena, V., Rai, P.K., Ghosh, A., and Moharana, P.C.:
592	Municipal solid waste (MSW): Strategies to improve salt affected soil sustainability: A review.
593	Waste Manag. 84, 38-53. https://doi.org/10.1016/j.wasman.2018.11.020, 2019
594	Munns, R.: Comparative physiology of salt and water stress. Plant, Cell Environ. 25, 239-250.
595	https://doi.org/10.1046/j.0016-8025.2001.00808.x, 2002
596	Mussatto, S.I.: Brewer's spent grain: a valuable feedstock for industrial applications. J. Sci. Food
597	Agric. 94, 1264–1275. https://doi.org/10.1002/jsfa.6486, 2014
598	Mussatto, S.I., and Roberto, I.C.: Chemical characterization and liberation of pentose sugars from
599	brewer's spent grain. J. Chem. Technol. Biotechnol. 81, 268-274.
600	https://doi.org/10.1002/jctb.1374, 2006
601	Nadeem, S.M., Ahmad, M., Zahir, Z.A., Javaid, A., and Ashraf, M.: The role of mycorrhizae and
602	plant growth promoting rhizobacteria (PGPR) in improving crop productivity under stressful
603	environments. Biotechnol. Adv. 32, 429–448.
604	https://doi.org/10.1016/j.biotechadv.2013.12.005, 2014
605	Nassar, I., Rashad, M., Aboukila, E., and Hafez, M.: Effect of Beer Industry Wastes Compost on
606	Some Physical and Chemical Properties of a Sandy Soil Effect of Beer Industry Wastes and
607	Compost on Some Physical and Chemical Properties of a Sandy Soil. J. Agric. Environ. Sci.
608	ISBN 1687-1464, 2014
609	Normandin, V., Kotuby-Amacher, J., and Miller, R.O.: Modification of the ammonium acetate
610	
	extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci.
611	extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998
611 612	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The</li> </ul>
611 612 613	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural</li> </ul>
611 612 613 614	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural ecosystem. Soil Tillage Res. 132, 30–38. https://doi.org/10.1016/j.still.2013.04.007, 2013</li> </ul>
611 612 613 614 615	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural ecosystem. Soil Tillage Res. 132, 30–38. https://doi.org/10.1016/j.still.2013.04.007, 2013</li> <li>Page, A.L., and Miller, R.H.: Methods of Soil Analysis. Part 2, Second Edition, in: A. L. Page,</li> </ul>
<ul> <li>611</li> <li>612</li> <li>613</li> <li>614</li> <li>615</li> <li>616</li> </ul>	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural ecosystem. Soil Tillage Res. 132, 30–38. https://doi.org/10.1016/j.still.2013.04.007, 2013</li> <li>Page, A.L., and Miller, R.H.: Methods of Soil Analysis. Part 2 , Second Edition, in: A. L. Page, Miller, R.H.D.R.K. (Ed.), Methods Of Soil Analysis.</li> </ul>
<ul> <li>611</li> <li>612</li> <li>613</li> <li>614</li> <li>615</li> <li>616</li> <li>617</li> </ul>	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural ecosystem. Soil Tillage Res. 132, 30–38. https://doi.org/10.1016/j.still.2013.04.007, 2013</li> <li>Page, A.L., and Miller, R.H.: Methods of Soil Analysis. Part 2, Second Edition, in: A. L. Page, Miller, R.H.D.R.K. (Ed.), Methods Of Soil Analysis. https://doi.org/10.2134/agronmonogr9.2.2ed.c41, 1982</li> </ul>
<ul> <li>611</li> <li>612</li> <li>613</li> <li>614</li> <li>615</li> <li>616</li> <li>617</li> <li>618</li> </ul>	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural ecosystem. Soil Tillage Res. 132, 30–38. https://doi.org/10.1016/j.still.2013.04.007, 2013</li> <li>Page, A.L., and Miller, R.H.: Methods of Soil Analysis. Part 2 , Second Edition, in: A. L. Page, Miller, R.H.D.R.K. (Ed.), Methods Of Soil Analysis. https://doi.org/10.2134/agronmonogr9.2.2ed.c41, 1982</li> <li>Pii, Y., Mimmo, T., Tomasi, N., Terzano, R., Cesco, S., and Crecchio, C.: Microbial interactions in</li> </ul>
<ul> <li>611</li> <li>612</li> <li>613</li> <li>614</li> <li>615</li> <li>616</li> <li>617</li> <li>618</li> <li>619</li> </ul>	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural ecosystem. Soil Tillage Res. 132, 30–38. https://doi.org/10.1016/j.still.2013.04.007, 2013</li> <li>Page, A.L., and Miller, R.H.: Methods of Soil Analysis. Part 2 , Second Edition, in: A. L. Page, Miller, R.H.D.R.K. (Ed.), Methods Of Soil Analysis. https://doi.org/10.2134/agronmonogr9.2.2ed.c41, 1982</li> <li>Pii, Y., Mimmo, T., Tomasi, N., Terzano, R., Cesco, S., and Crecchio, C.: Microbial interactions in the rhizosphere: beneficial influences of plant growth-promoting rhizobacteria on nutrient</li> </ul>
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611 612 613 614 615 616 617 618 619 620 621	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural ecosystem. Soil Tillage Res. 132, 30–38. https://doi.org/10.1016/j.still.2013.04.007, 2013</li> <li>Page, A.L., and Miller, R.H.: Methods of Soil Analysis. Part 2, Second Edition, in: A. L. Page, Miller, R.H.D.R.K. (Ed.), Methods Of Soil Analysis. https://doi.org/10.2134/agronmonogr9.2.2ed.c41, 1982</li> <li>Pii, Y., Minmo, T., Tomasi, N., Terzano, R., Cesco, S., and Crecchio, C.: Microbial interactions in the rhizosphere: beneficial influences of plant growth-promoting rhizobacteria on nutrient acquisition process. A review. Biol. Fertil. Soils 51, 403–415. https://doi.org/10.1007/s00374-015-0996-1, 2015a</li> </ul>
611 612 613 614 615 616 617 618 619 620 621 622	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural ecosystem. Soil Tillage Res. 132, 30–38. https://doi.org/10.1016/j.still.2013.04.007, 2013</li> <li>Page, A.L., and Miller, R.H.: Methods of Soil Analysis. Part 2 , Second Edition, in: A. L. Page, Miller, R.H.D.R.K. (Ed.), Methods Of Soil Analysis. https://doi.org/10.2134/agronmonogr9.2.2ed.c41, 1982</li> <li>Pii, Y., Mimmo, T., Tomasi, N., Terzano, R., Cesco, S., and Crecchio, C.: Microbial interactions in the rhizosphere: beneficial influences of plant growth-promoting rhizobacteria on nutrient acquisition process. A review. Biol. Fertil. Soils 51, 403–415. https://doi.org/10.1007/s00374-015-0996-1, 2015a</li> <li>Pii, Y., Penn, A., Terzano, R., Crecchio, C., Mimmo, T., and Cesco, S.: Plant-microorganism-soil</li> </ul>
611 612 613 614 615 616 617 618 619 620 621 622 623	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural ecosystem. Soil Tillage Res. 132, 30–38. https://doi.org/10.1016/j.still.2013.04.007, 2013</li> <li>Page, A.L., and Miller, R.H.: Methods of Soil Analysis. Part 2 , Second Edition, in: A. L. Page, Miller, R.H.D.R.K. (Ed.), Methods Of Soil Analysis. https://doi.org/10.2134/agronmonogr9.2.2ed.c41, 1982</li> <li>Pii, Y., Mimmo, T., Tomasi, N., Terzano, R., Cesco, S., and Crecchio, C.: Microbial interactions in the rhizosphere: beneficial influences of plant growth-promoting rhizobacteria on nutrient acquisition process. A review. Biol. Fertil. Soils 51, 403–415. https://doi.org/10.1007/s00374-015-0996-1, 2015a</li> <li>Pii, Y., Penn, A., Terzano, R., Crecchio, C., Mimmo, T., and Cesco, S.: Plant-microorganism-soil interactions influence the Fe availability in the rhizosphere of cucumber plants. Plant Physiol.</li> </ul>
611 612 613 614 615 616 617 618 619 620 621 622 623 624	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural ecosystem. Soil Tillage Res. 132, 30–38. https://doi.org/10.1016/j.still.2013.04.007, 2013</li> <li>Page, A.L., and Miller, R.H.: Methods of Soil Analysis. Part 2 , Second Edition, in: A. L. Page, Miller, R.H.D.R.K. (Ed.), Methods Of Soil Analysis. https://doi.org/10.2134/agronmonogr9.2.2ed.c41, 1982</li> <li>Pii, Y., Mimmo, T., Tomasi, N., Terzano, R., Cesco, S., and Crecchio, C.: Microbial interactions in the rhizosphere: beneficial influences of plant growth-promoting rhizobacteria on nutrient acquisition process. A review. Biol. Fertil. Soils 51, 403–415. https://doi.org/10.1007/s00374-015-0996-1, 2015a</li> <li>Pii, Y., Penn, A., Terzano, R., Crecchio, C., Mimmo, T., and Cesco, S.: Plant-microorganism-soil interactions influence the Fe availability in the rhizosphere of cucumber plants. Plant Physiol. Biochem. 87, 45–52. https://doi.org/10.1016/j.plaphy.2014.12.014, 2015b</li> </ul>
611 612 613 614 615 616 617 618 619 620 621 622 623 624 625	<ul> <li>extractant for the determination of exchangeable cations in calcareous soils. Commun. Soil Sci. Plant Anal. 29, 1785–1791. https://doi.org/10.1080/00103629809370069, 1998</li> <li>Ouyang, W., Shan, Y., Hao, F., Chen, S., Pu, X., and Wang, M.K.: Soil &amp; Tillage Research The effect on soil nutrients resulting from land use transformations in a freeze-thaw agricultural ecosystem. Soil Tillage Res. 132, 30–38. https://doi.org/10.1016/j.still.2013.04.007, 2013</li> <li>Page, A.L., and Miller, R.H.: Methods of Soil Analysis. Part 2 , Second Edition, in: A. L. Page, Miller, R.H.D.R.K. (Ed.), Methods Of Soil Analysis. https://doi.org/10.2134/agronmonogr9.2.2ed.c41, 1982</li> <li>Pii, Y., Mimmo, T., Tomasi, N., Terzano, R., Cesco, S., and Crecchio, C.: Microbial interactions in the rhizosphere: beneficial influences of plant growth-promoting rhizobacteria on nutrient acquisition process. A review. Biol. Fertil. Soils 51, 403–415. https://doi.org/10.1007/s00374-015-0996-1, 2015a</li> <li>Pii, Y., Penn, A., Terzano, R., Crecchio, C., Mimmo, T., and Cesco, S.: Plant-microorganism-soil interactions influence the Fe availability in the rhizosphere of cucumber plants. Plant Physiol. Biochem. 87, 45–52. https://doi.org/10.1016/j.plaphy.2014.12.014, 2015b</li> <li>Qadir, M., Tubeileh, A., Akhtar, J., Larbi, A., Minhas, P.S., and Khan, M.A.: Productivity</li> </ul>





627	429–453. https://doi.org/10.1002/ldr.853, 2008
628	Rashad, M., Hafez, M., Emran, M., Aboukila, E., and Nassar. I.: Influence of Environment-Friendly
629	Organic Wastes on the Properties of Sandy Soil under Growing Zea mays L. in Arid Regions.
630	World Acad. Sci. Eng. Technol. 10, 588–594. https://doi.org/waset.org/1999.1/10005388, 2016
631	Rashad, M., Dultz, S., and Guggenberger, G.: Dissolved organic matter release and retention in an
632	alkaline soil from the Nile River Delta in relation to surface charge and electrolyte type.
633	Geoderma 158, 385–391. https://doi.org/10.1016/j.geoderma.2010.06.007, 2010
634	Rath, K.M., and Rousk, J.: Salt effects on the soil microbial decomposer community and their role
635	in organic carbon cycling: A review. Soil Biol. Biochem. 81, 108-123.
636	https://doi.org/10.1016/j.soilbio.2014.11.001.2, 2015
637	Renuka, N., Guldhe, A., Prasanna, R., Singh, P., and Bux, F.: Microalgae as multi-functional
638	options in modern agriculture: current trends, prospects and challenges. Biotechnol. Adv. 36,
639	1255-1273. https://doi.org/10.1016/j.biotechadv.2018.04.004, 2018
640	Richards, L.A.: Diagnosis and improvement of saline and alkali soils. Soil Sc., 1954
641	Rojas-Tapias, D., Moreno-Galván, A., Pardo-Díaz, S., Obando, M., Rivera, D., and Bonilla, R.:
642	Effect of inoculation with plant growth-promoting bacteria (PGPB) on amelioration of saline
643	stress in maize (Zea mays). Appl. Soil Ecol. 61, 264–272.
644	https://doi.org/10.1016/j.apsoil.2012.01.006, 2012
645	Sant'Anna, S.A.C., Martins, M.R., Goulart, J.M., Araújo, S.N., Araújo, E.S., Zaman, M., Jantalia,
646	C.P., Alves, B.J.R., and Boddey, R.M., Urquiaga, S.: Biological nitrogen fixation and soil N2O
647	emissions from legume residues in an Acrisol in SE Brazil. Geoderma Reg. 15, e00196.
648	https://doi.org/10.1016/j.geodrs.2018.e00196, 2018
649	Sastre-Conde, I., Carmen Lobo, M., Icela Beltrán-Hernández, R., and Poggi-Varaldo, H.M.:
650	Remediation of saline soils by a two-step process: Washing and amendment with sludge.
651	Geoderma 247-248, 140-150. https://doi.org/10.1016/j.geoderma.2014.12.002, 2015
652	Scherer, H.W., Metker, D.J., and Welp, G.: Effect of long-term organic amendments on chemical
653	and microbial properties of a luvisol. Plant, Soil Environ. 57, 513-518.
654	https://doi.org/10.17221/3283-PSE, 2011
655	Shi, S., Tian, L., Nasir, F., Bahadur, A., Batool, A., Luo, S., Yang, F., Wang, Z., and Tian, C.:
656	Response of microbial communities and enzyme activities to amendments in saline-alkaline
657	soils. Appl. Soil Ecol. 135, 16-24. https://doi.org/10.1016/j.apsoil.2018.11.003, 2019
658	Shrivastava, P., and Kumar, R.: Soil salinity: A serious environmental issue and plant growth
659	promoting bacteria as one of the tools for its alleviation. Saudi J. Biol. Sci. 22, 123-131.
660	https://doi.org/10.1016/j.sjbs.2014.12.001, 2015
661	Soltanpour, P.N., and Schwab, A.P.: A new soil test for simultaneous extraction of macro- and
662	micro-nutrients in alkaline soils. Commun. Soil Sci. Plant Anal. 8, 195-207.
663	https://doi.org/10.1080/00103627709366714, 1977
664	Trivedi, P., Delgado-Baquerizo, M., Jeffries, T.C., Trivedi, C., Anderson, I.C., Lai, K., McNee, M.,
665	Flower, K., Pal Singh, B., Minkey, D., and Singh, B.K.: Soil aggregation and associated
666	microbial communities modify the impact of agricultural management on carbon content.
667	Environ. Microbiol. 19, 3070-3086. https://doi.org/10.1111/1462-2920.13779, 2017
668	Wang, X., and Cai, Q.S.: Steel Slag as an Iron Fertilizer for Corn Growth and Soil Improvement in
669	a Pot Experiment1 1 Project supported by the National Natural Science Foundation of China
670	(No. 30270800). Pedosphere 16, 519–524. https://doi.org/10.1016/S1002-0160(06)60083-0,





671	2006

- 672 Wilson, G.W.T., Rice, C.W., Rillig, M.C., Springer, A., and Hartnett, D.C.: Soil aggregation and 673 carbon sequestration are tightly correlated with the abundance of arbuscular mycorrhizal fungi: long-term 674 Results from field experiments. Ecol. Lett. 12, 452-461. 675 https://doi.org/10.1111/j.1461-0248.2009.01303.x, 2009
- Yu, Y.-Y., Li, S.-M., Qiu, J.-P., Li, J.-G., Luo, Y.-M., and Guo, J.-H.: Combination of agricultural
  waste compost and biofertilizer improves yield and enhances the sustainability of a pepper
  field. J. Plant Nutr. Soil Sci. 182, 560–569. https://doi.org/10.1002/jpln.201800223, 2019
- 679 Zarea, M.J., Hajinia, S., Karimi, N., Mohammadi Goltapeh, E., Rejali, F., and Varma, A.: Effect of 680 Piriformospora indica and Azospirillum strains from saline or non-saline soil on mitigation of 681 the effects of NaCl. Soil Biol. Biochem. 45. 139-146. 682 https://doi.org/10.1016/j.soilbio.2011.11.006, 2012
- Zhang, H., Kim, M.-S., Sun, Y., Dowd, S.E., Shi, H., and Paré, P.W.: Soil Bacteria Confer Plant
  Salt Tolerance by Tissue-Specific Regulation of the Sodium Transporter HKT1. Mol. PlantMicrobe Interact. 21, 737–744. https://doi.org/10.1094/MPMI-21-6-0737, 2008
- Zhang, T., Wang, T., Liu, K., Wang, L., Wang, K., and Zhou, Y.: Effects of different amendments
  for the reclamation of coastal saline soil on soil nutrient dynamics and electrical conductivity
  responses. Agric. Water Manag. 159, 115–122. https://doi.org/10.1016/j.agwat.2015.06.002,
  2015
- Zhang, W. wen, Wang, C., Xue, R., and Wang, L. jie.: Effects of salinity on the soil microbial
  community and ssoil fertility. J. Integr. Agric. 18, 1360–1368. https://doi.org/10.1016/S20953119(18)62077-5, 2019
- Zhao, Y., Wang, S., Li, Y., Liu, J., Zhuo, Y., Chen, H., Wang, J., Xu, L., and Sun, Z.: Extensive
  reclamation of saline-sodic soils with flue gas desulfurization gypsum on the Songnen Plain,
  Northeast China. Geoderma 321, 52–60. https://doi.org/10.1016/j.geoderma.2018.01.033, 2018
- Zhen, Z., Liu, H., Wang, N., Guo, L., Meng, J., Ding, N., Wu, G., and Jiang, G.: Effects of manure
   compost application on soil microbial community diversity and soil microenvironments in a
- temperate cropland in China. PLOS One 9. https://doi.org/10.1371/journal.pone.0108555, 2014
- Zhuang, X., Chen, J., Shim, H., and Bai, Z.: New advances in plant growth-promoting rhizobacteria
  for bioremediation. Environ. Int. 33, 406–413. https://doi.org/10.1016/j.envint.2006.12.005,
  2007