

Ramped thermal analysis for isolating biologically meaningful soil organic matter fractions with distinct residence times

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Abstract. In this work, we assess whether or not ramped thermal oxidation coupled with determination of the radiocarbon content of the evolved CO₂ can be used to isolate distinct thermal fractions of SOM along with direct information on the turnover rate of each thermal fraction. Using a 30 year time-series of soil samples from a well characterized agronomic trial, we found that the incorporation of the bomb-spike in atmospheric ¹⁴CO₂ into thermal fractions of increasing resistance to thermal decomposition could be successfully modelled. With increasing temperature, which is proportional to activation energy, the mean residence time of the thermal fractions increased from 10 to 400 years. Importantly, the first four of five thermal fractions appeared to be a mixture of fast and increasingly slower cycling SOM. To further understand the composition of different thermal fractions, stepped pyrolysis-gas chromatography-mass spectrometry (py-GC/MS) experiments were performed at five temperatures ranging from 330 to 735 °C. The py-GC/MS data showed a reproducible shift in the chemistry of pyrolysis products across the temperature gradient trending from polysaccharides and lipids at low temperature to lignin and microbial-derived compounds at middle temperatures to aromatic and unknown compounds at the highest temperatures. Integrating the ¹⁴C and Py-GC-MS data suggests the organic compounds, with the exception of aromatic moieties likely derived from wildfire, with centennial residence times are not more complex but may be protected from pyrolysis, and likely also from biological mineralization, by interactions with mineral surfaces.

1 Introduction

Soil organic matter (SOM) consists of a spectrum of material from labile, rapidly cycling compounds to mineral-stabilized molecules that resist degradation for centuries. This spectrum of turnover rates is due to a combination of organic matter composition with varying reactivity, various degrees of interaction between organic and mineral phases and greatly varying microclimates more or less suited to microbial activity (Lehmann and Kleber, 2015; Ruamps et al., 2013; Schmidt et al., 2011). Soil scientists often deal with this complexity by fractionating SOM into what are thought to be more homogenous pools in terms of reactivity, composition or microbial accessibility depending on the particulars of the study (Christensen, 2001; von Lützow et al., 2007). Similarly, soil carbon cycle models typically divide SOM into conceptual pools with distinct mean turnover rates (Manzoni and Porporato, 2009). However, empiricists have been trying for decades with varying

30 degrees of success to link physically and chemically isolated fractions of SOM to the conceptual SOM pools in carbon cycle models (Skjemstad et al., 2004; Zimmermann et al., 2007).

Fractionation schemes, employing various physical, chemical, biological or thermal methods, are generally used to reduce the inherent complexity found in SOM. Both size fractionation and density separation are commonly used to separate particulate from mineral associated SOM (Elliott and Cambardella, 1991; Golchin et al., 1994; Sollins et al., 2009).
35 Hydrolysis with strong acid has been shown to isolate SOM that is consistently 100s to 1000s of years older than the bulk SOM (Paul et al., 2001). Biological fractionation involves the modeling of mass loss or CO₂ evolution during a laboratory incubation experiment (Grandy and Robertson, 2007; Schädel et al., 2013). Baldock et al. (2013), recognizing that fire-derived pyrogenic carbon (PyC) has distinct properties from plant or microbial products, combined physical size fractionation with the use of solid-state ¹³C NMR spectroscopy to mathematically isolate the PyC fraction from each size
40 fraction. There are advantages and disadvantages to each of these techniques (Poehlau et al., 2018), but even the most detailed fractionation schemes are unable to isolate homogenous SOM pools (Jastrow et al., 1996; Sanderman et al., 2013; Torn et al., 2013) which may be as much a result of methodological issues as the fact that there are multiple pathways for SOM formation (Cotrufo et al., 2013; Sokol et al., 2019).

Thermal analysis techniques, long used in petrochemical exploration and clay mineralogy, offer a promising alternative or
45 complement to physical- and chemical-based fractionation methods and are increasingly being applied to studies of SOM stability and loss (Peltre et al., 2013; Plante et al., 2009; Williams et al., 2018). The basic premise of this suite of tools is that by slowly heating a sample the energy needed to evolve the carbon at different temperatures, whether that comes from breaking an organic-organic bond or disrupting an organic-mineral bond or other association, can be quantified and that this energy yield is somewhat related to the energy requirements for enzymatic degradation of SOM (Williams and Plante, 2018).
50 While research suggests that more complex organic molecules have higher thermal stability (Lopez-Capel et al., 2005; Yang et al., 2006), contradictory results have also been reported (Rovira et al., 2008). Several studies have also found good correlations between thermal stability indices and biological stability (Peltre et al., 2013; Soucémariadin et al., 2018) and model-derived stable C pools (Cécillon et al., 2018). However, other studies have found that new carbon preferentially flowed into more thermally stable fractions (Helfrich et al., 2010; Schiedung et al., 2017) suggesting that the relationship
55 between thermal stability and SOM cycling concepts may not be straightforward.

A recent advance in thermal analysis is the coupling of a temperature-controlled oven to a vacuum line, termed ramped pyrolysis-oxidation (RPO), enabling collection of evolved gas at distinct temperature regions for subsequent ¹³C and ¹⁴C analysis (Rosenheim et al., 2008). Unlike interpretation of other thermal indices, ¹⁴C is a direct and powerful tracer of soil carbon cycling by providing information on the age and turnover rate of an isolated fraction of SOM (Trumbore, 2009;
60 Trumbore et al., 1996). In the first application of the RPO system to SOM, Plante et al. (2013) found that more thermally stable fractions also contained the oldest, most ¹⁴C-depleted carbon.

In this study, we present the first use of ramped thermal analysis with a time series of soil samples to investigate the carbon cycling rate of SOM with increasing thermal stability. The use of a multi-decadal time series of soil samples allows for modeling the uptake of the bomb-spike in atmospheric $^{14}\text{CO}_2$ due to nuclear weapons testing into SOM fractions in order to determine the turnover rate of those fractions (Baisden et al., 2013). We further couple the ^{14}C -based turnover time estimates with parallel chemical characterization of similar thermal fractions using a stepped pyrolysis-gas chromatography/mass spectrometry (py-GC/MS) approach. With the compound specific chemistry data we are aiming to be able to start to explain possible mechanisms controlling the turnover of the thermal fractions thereby providing new insights into the linkages between thermal stability, SOM composition and microbial cycling of SOM.

70 **2 Materials and Methods**

2.1 Trial and soil description

The soil used in this study come from a long-term agricultural research trial evaluating alternative crop rotations located at the Waite Research Institute in South Australia (34.967 S, 138.634 E) with a Mediterranean climate where 80% of the 626 mm of average rainfall occurs during the April-October growing season. The soil is classified as a Rhodoxeralf in the USDA taxonomy (Soil Survey Staff 1999) or a Chromic Luvisol in the WRB taxonomy (IUSS Working Group WRB 2015) with a fine sandy loam texture in the upper horizons, and a mean pH (H_2O) of 5.9 and clay content of 18% for the 0-10 cm layer (Grace et al., 1995). Full trial history including management records, agronomic performance and soil data have been reported elsewhere (Grace et al., 1995; Sanderman et al., 2017). These data including monthly climate records can be accessed from the CSIRO Data Access Portal (doi: 10.4225/08/55E5165EC0D29).

80 For this study, we have chosen to focus on the permanent pasture treatment because previous work (Sanderman et al., 2017) found the greatest uptake of the bomb-spike in atmospheric $^{14}\text{CO}_2$ in this treatment. This particular trial strip (#29) was under a wheat/pea rotation from 1925 until 1950 but then converted to an improved pasture by sowing a mix of annual rye grass, subterranean clover and phalaris in 1950 and was then managed consistently with simulated grazing (i.e. hand mowing) and periodic re-sowing until the end of the trial in 1996. In April of 1963, 1973, 1983 and 1993 soil samples from the top 10 cm were collected as a composite of 20 cores taken along the center line of this 90 m long strip trial. Soils were dried at 40 °C for > 48 hr before being stored in glass jars prior to subsampling in 2015. Previous analyses (Sanderman et al., 2016, 2017) suggest that SOM and the proportion in a particulate fraction only varied slightly throughout the 1963-1993 period (Table 1). The large change in $\Delta^{14}\text{C}$ was due to uptake and loss of the bomb-spike in atmospheric $^{14}\text{CO}_2$.

2.2 Ramped pyrolysis oxidation

90 Ramped pyrolysis oxidation (RPO) was performed using the Dirt Burner, a custom built evolved gas system at the National Ocean Sciences Accelerator Mass Spectrometry (NOSAMS) facility, where a sample can be linearly heated under either

pyrolyzing or oxidizing conditions. The evolved gases are then oxidized to CO₂, measured on an in-line infrared gas analyzer and trapped for subsequent analysis of ¹³C and ¹⁴C composition. Initial instrument development is described by Rosenheim et al. (2008) with upgrades described by Plante et al. (2013). See Hemingway et al. (2017b) for a complete description of current instrument configuration and operating conditions. In this investigation we operated the Dirt Burner only in oxidizing mode. It has been demonstrated that the distribution of activation energies and ¹⁴C age of thermal fractions for soil are similar under oxidizing and pyrolysis modes (Grant et al., 2019). The 1973 soil sample was initially run using a fast ramp (20 ° C min⁻¹) where only CO₂ concentration was recorded. Using an inversion method (Hemingway et al., 2017a), five distinct thermal fractions were identified from this thermogram with temperature ranges of 100-325, 325-400, 400-445, 445-100 515 and > 515 ° C (Figure A1). Subsequently, 45 mg of each soil sample were combusted on the Dirt Burner using a slow temperature ramp (5 ° C min⁻¹) and the evolved CO₂ in each of these five fractions was cryogenically collected and purified for subsequent analyses. The RPO fractions were then split for ¹³C analysis on a dual-inlet isotope ratio mass spectrometer (McNichol et al., 1994b) and ¹⁴C composition via accelerator mass spectrometry after graphitization (McNichol et al., 1994a). Stable isotope data are expressed in δ¹³C notation (‰) relative to Vienna Pee Dee Belemnite (VPDB) standard. Radiocarbon data, after using ¹³C data to correct for mass dependent fractionation, are reported from NOSAMS as fraction modern (Fm, where the 1950 atmosphere is assigned a value of 1.0) and subsequently converted to the geochemical Δ¹⁴C notation (Stuiver and Polach, 1977) for bomb-spike modeling.

2.3 Pyrolysis gas chromatography-mass spectrometry

We determined the relative percentages and ratios of chemical classes using pyrolysis-mass spectrometry/gas chromatography (py-GC/MS) using methods described previously (Grandy et al., 2009; Kallenbach et al., 2015; Wickings et al., 2011). However, in contrast to our previous studies in which we used a single pyrolysis temperature, here we used a ‘ramp’ or stepped approach by pyrolyzing the same sample at five distinct, sequentially increasing temperatures: 330, 396, 444, 503 and 735 ° C (Buurman et al., 2007; Hempfling and Schulten, 1990; Williams et al., 2014). Thus, the same sample was pyrolyzed five times corresponding with each of these temperatures. Temperatures were chosen to correspond with the temperature ranges from the RPO analysis. Samples were pyrolyzed at each temperature and pyrolysis products transferred to a GC where compounds were separated on a 60 m capillary column with a starting temperature of 40°C followed by a temperature ramp of 5°C min⁻¹ to 270°C followed by a final ramp (30°C min⁻¹) to 300°C. Compounds were immediately transferred from the GC to an ion trap MS where they were ionized at 70 eV in the EI mode with the source temperature held at 200°C, detected via electron multiplier, and identified using a compound library built using the National Institute of Standards and Technology (NIST) database and published literature as reported previously (Grandy et al. 2007; Grandy et al., 2009). Individual compounds were classified by their source as polysaccharide, aromatic, phenolic, protein, N-bearing (non-proteins) or unknown. Proteins include pyridines, pyrroles and indole that have been previously identified as pyrolysis products of proteins (Schulten and Schnitzer, 1997). The compounds that are in the unknown category are identified but can be derived in nature or due to pyrolysis from different sources (e.g. both protein and aromatic).

125 2.4 Data analysis

The turnover time of each thermal fraction was determined by modeling the incorporation of the bomb-spike in atmospheric $^{14}\text{CO}_2$ into each fraction (Baisden et al., 2013). We have applied a steady state soil carbon turnover model to each thermal fraction where carbon inputs (C_{in}) are portioned into each model pool proportional to the fractional distribution of carbon in each pool (f_{pool}) with the $^{14}\text{C}/^{12}\text{C}$ ratio of the previous year's atmospheric CO_2 (data from Currie et al., 2011). Carbon losses
130 follow first-order kinetics with a characteristic decay rate for each pool (k_{pool}) and shifts in $^{14}\text{C}/^{12}\text{C}$ ratio are also affected by radioactive decay ($\lambda = 1.21 \times 10^{-4} \text{ yr}^{-1}$). Turnover time (τ_{pool}) is simply the inverse of the decay rate for each pool. First, we assumed that each thermal fraction is a single homogenous pool and solve the turnover model to find a single k value by minimizing the sum of square errors (SSE) between observed and predicted $\Delta^{14}\text{C}$ data for the four years of data (1963, 1973, 1983 and 1993) for that particular thermal fraction. Initial results suggested that most of the thermal fractions could not be
135 represented as a single homogenous pool, so we then applied a two-pool model by assuming a fixed k for the fast cycling pool ($k_{fast} = 0.25 \text{ yr}^{-1}$) and allowed the size (f_{slow}) and decay rate (k_{slow}) of the slow cycling pool to vary while minimizing the SSE between observed and predicted $\Delta^{14}\text{C}$ data for each thermal fraction. Given the limited number of degrees of freedom, we decided not to allow for the simultaneous optimization of all three parameters. The value for k_{fast} was set at 0.25 yr^{-1} after performing a simple sensitivity analysis where k_{fast} was varied from 0.1 to 1.0 yr^{-1} and f_{slow} and k_{slow} were optimized. This
140 exercise suggested that the overall lowest mean RMSE across thermal fractions was achieved with $k_{fast} = 0.25 \text{ yr}^{-1}$. Model performance was assessed by calculating the root mean square error (RMSE) between observed and predicted $\Delta^{14}\text{C}$ values.

The stepped py-GC/MS data are reported as percent relative abundance for each identified compound. We present these data qualitatively in two ways. First, data from each thermal interval were averaged across years and shifts in the eight major compound classes are shown. Second, all compounds that averaged $> 1\%$ abundance across all 20 samples (4 years x 5
145 thermal intervals) were included in a non-metric multidimensional scaling analysis (e.g. Grandy et al., 2009) after constructing a resemblance matrix using Euclidean distance from the percent abundance data.

3 Results

3.1 RPO results

All samples produced nearly identical thermograms (Figure 1) suggesting that the activation energies assigned to the 1973
150 sample (Figure A1) are applicable to samples from all four decades. Fractions 2 and 3 contained the greatest proportion of SOM, with the distribution across all five fractions being consistent between across all years (Table B1). Increasing $\delta^{13}\text{C}$ values were found from F1 to F3 with little additional increase from F3 to F5 (Figure 2a). A greater incorporation of the bomb-spike in ^{14}C was found in the lower temperature fractions with no modern carbon seen in the highest temperature fraction (Figure 2b). There wasn't enough sample for ^{13}C analysis of F1 or F5 in 1963. Additionally, there was not enough

155 CO₂ for ¹⁴C analysis of F5 in 1963. The Δ¹⁴C value for F1 in 1973 is included in Figure 2b but removed from subsequent bomb-spike modeling.

3.2 Bomb-spike turnover modelling results

The single-pool steady-state model to trace the incorporation of the bomb-spike in atmospheric ¹⁴CO₂ into the thermal fractions indicated that turnover times increased from 37 to 386 years with increasing temperature (Table 2). However, with the exception of F5 a single-pool model resulted in unacceptably high error with RMSE values ranging from 25 to 45% for the other fractions. Visually it was clear that the single pool solution could not capture the dynamics of the increase and subsequent decrease in Δ¹⁴C in F1-F4 (Figure C1). A two-pool model with a fast cycling pool (τ = 4 yr) and a variably sized slower cycling pool was generally able to capture these dynamics with RMSE values below 13.8% (Table 2). With increasing activation energy of the thermal fraction, the proportion of fast cycling carbon decreased (Figure 3a) and the turnover time of the slow cycling fraction increased (Figure 3b).

As an independent first-order check on the reasonableness of MRT results for the thermal fractions, the inventory-weighted MRT of the bulk soil was 16.5 years matching the results of applying a 3-pool turnover model to the bulk ¹⁴C data which varied from 11-20 years depending on the model structure (Sanderman et al., 2017).

3.3 Py-GC/MS results

170 A total of 172 individual compounds were identified in one or more samples. Many of these compounds were a minor fraction of only a single sample. When these compounds were classified by source, there were sharp differences in the dominant chemical compound classes released at each pyrolysis temperature (Figure 4a). Lipids (42.5 ± 2.74 %) and polysaccharides (42.4 ± 2.12 %) were the dominant classes of compounds released at the lowest pyrolysis temperature (330 °C). Relative lipid abundance was <10% of the pyrolysis products for all higher temperatures and <1% relative abundance at the highest pyrolysis temperature (735 °C). Polysaccharides (51.2 ± 1.68 %) remained an abundant component of the pyrolysis products at a pyrolysis temperature of 396 °C and 444 (27.4 %) °C but were <10% relative abundance at the two highest temperatures. At the fourth temperature level (503 °C), N-bearing compounds, proteins, phenols, and unknown compounds dominated the chemical signature and were all ~15-20% relative abundance. Phenols (31.4%) were the most abundant compound class released at the highest temperature (735 °C) while proteins, aromatics and compounds of unknown origin were all >15% relative abundance.

We identified 23 individual compounds that averaged more than one 1% relative abundance averaged across all samples (Figure 4b). These 23 compounds collectively represented from about ~70 to 90% of the total sample relative abundance at the five different temperatures (Table D1). These compounds included three aromatic compounds, one lignin derivative, one lipid, two N-bearing compounds, two phenols, five polysaccharides, five proteins, and four compounds that fell into the

185 unknown origin. The most abundant compound was hexadecanoic acid methyl ester (palmitic acid) with a mean relative abundance across all samples of about 10% but composing 42% of F1 (Table D1). The next two most abundant compounds were both phenols and included phenol and 4-methyl phenol, both increasing with increasing temperature, followed by the common polysaccharide pyrolysis product furfural, primarily found in the lower temperature fractions.

4 Discussion

190 While this study has only examined one soil under consistent management over four decades, the combined results from the modeling of the incorporation of bomb-spike ^{14}C into thermal fractions and the stepped py-GC/MS analysis here suggest that evolved gas analyses can be powerful analytical tools for understanding the complexities of SOM cycling. With increasing activation energy, turnover times increased from decades to centuries (Figure 3) and there was a consistent (i.e. repeatable across time) strong shift in OM chemistry (Figure 4).

195 This investigation was partially framed in the context of testing evolved gas analysis as a tool for isolating biologically meaningful fractions of SOM. The results indicate that each of the five thermal fractions contained unique information. Each of the thermal fractions represented pools with a diverse mixture of organic materials with ^{14}C -based turnover times ranging from 10 to 400 years. A wide distribution of activation energy (Figure A1) was needed to describe each thermal fraction. Additionally, a one-pool turnover model could not represent the dynamics of ^{14}C in all but the most thermally stable fraction
200 (Table 3). This finding of heterogeneity within an isolated fraction also plagues other physical and chemical techniques for isolating carbon fractions (Sanderman et al. 2013; Torn et al. 2013). Here, by combining stable isotope data, ^{14}C -based turnover modeling and compound specific chemical characterization of the thermal fractions, we have found clues as to reasons for the heterogeneity of SOM within these fractions which provide insights into pathways of SOM formation and longer-term stabilization.

205 At the two lowest temperature intervals, where the ^{14}C data suggested rapid turnover rates, the polysaccharides comprised > 40% of the identified compounds. The dominant polysaccharide products at the two lowest temperatures included furfural, 3-furaldehyde, and levoglucosenone. Furfural is a pyrolysis product of hexoses and pentoses as well as uronic acids. The hexoses originate from both plant and microbial residues, while the pentoses are primarily microbial in origin. Levoglucosenone is a pyrolysis product of neutral sugars such as glucose, galactose and mannose (Saiz-Jimenez et al., 1979;
210 Saiz-Jimenez and De Leeuw, 1986). These polysaccharides may be part of partially decomposed residues that dominate the light or particulate fractions of SOM or be part of other soil carbon pools not strongly protected from thermal degradation by association with minerals. Across all thermal fractions, polysaccharide abundance was strongly correlated with the proportion of fast cycling carbon in that fraction.

Identifiable lignin derivatives (e.g. syringol and guaicol) were most abundant in the second and third thermal fractions but never constituted more than 10% of either thermal fraction. While the chemical complexity of lignin likely affords some thermal stability, as suggested by the greater activation energy than the thermal fraction dominated by polysaccharides, the ¹⁴C data suggest that lignin is not a particularly persistent compound in this soil. This finding is consistent with past studies findings that there is little recognizable lignin found in the more stable clay fractions of many soils (Baldock et al., 1997; Grandy and Neff, 2008; Schulten and Leinweber, 2000), although amounts may vary across different mineral types and environmental contexts (Kramer et al., 2012).

Besides polysaccharides, hexadecanoic acid methyl ester (palmitic acid) was the other dominant constituent of the lowest temperature fraction (Table D1). This compound is a major constituent of microbial cell walls, especially fungi, and has been strongly correlated to both microbial biomass and activity (Zelles et al., 1992). The relative dominance of this lipid at low pyrolysis temperatures suggests that compared to other compounds it is not protected from degradation by either association with minerals or chemical complexity. There is a growing body of evidence suggesting that a substantial portion of stable SOM in many soils is microbial in origin (Kallenbach et al., 2015; Knicker, 2011; Miltner et al., 2012) which could lead to the misconception that microbial products are inherently more stable than plant-derived compounds. The findings here suggest that palmitic acid, the most abundant lipid that we identified, which is most likely of microbial origin, is not a very thermally stable component in soil with a much faster than average MRT. This may arise because these lipids are not forming direct covalent bonds with mineral surfaces nor are they forming other complexes (e.g. with metal oxides) that could make them resistant to thermal and biological degradation.

At the third and fourth highest temperatures phenols, proteins and N-bearing compounds became more relatively abundant. These phenols can originate from degraded lignin monomers but also other aromatics of plant or microbial origin. The proteins and other N-bearing compounds are important components of mineral-associated N. The presence of heterocyclic N (e.g. our furans, pyridines and pyrroles) is a common finding of studies using py-GC/MS (Schulten and Schnitzer, 1997; Leinweber et al. 2014). The formation of humic substances (de Assis et al., 2012) as well as the pyrolysis process itself (Hatcher et al., 2001) and fire (González-Pérez et al., 2004) are potential sources of heterocyclic N but there are plant and microbial inputs that contain also sources these may be secondary contributors compared to microbial synthesis (Leinweber et al. 2014; Paul, 2016). The pyrroles and pyridines, for example, may largely originate from microbes and Kallenbach et al. (2016) found in an artificial soil system with only glucose inputs that a variety of heterocyclic N compounds can be derived from the pyrolysis of microbial cells.

We have previously found that polysaccharides, proteins and N-bearing compounds are the dominant mineral-associated chemical fractions (Grandy et al., 2007; Grandy and Neff, 2008) as well as an increasing proportion of these compounds with soil depth (Rothstein et al., 2018). Anticipating that these same compounds would also dominate the slowest cycling highest temperature fraction, we found that aromatic compounds and compounds from an unknown source, which included

compounds such as toluene, the eleventh most abundant compound and that may be derived from pyrolyzing aromatics or proteins, were relatively abundant. There have been reports that pyrolysis of clay fractions can produce matrix effects that inflate the abundance of aromatic compounds (Schulten and Leinweber, 1993). We have seen no detectable evidence for this in our previous studies, which generally show very low abundance of aromatics in fine fractions. The relatively low clay content and dominance of illite in our soils also point to lower potential for matrix effects. More likely, the high relative abundance of stable aromatics, phenols and compounds of unknown origin can be attributed to the history of fire at this Mediterranean sites. Previous studies at the site indicate that ~30% of the total SOM is made up of pyrogenic aromatics (Sanderman et al., 2017).

The findings from this preliminary investigation add to the growing body of literature using evolved gas analysis as a tool for understanding soil organic matter dynamics. The two unique aspects of this work were using a time series of soil samples to calculate the MRT of different thermal fractions and relating these MRTs to the chemical composition of each fraction. We found that mean residence time increased with increasing activation energy. Modeling the incorporation of the bomb-spike in $^{14}\text{CO}_2$ indicated that the thermal fractions were, except for the most stable fraction, heterogeneous mixtures of fast and slow cycling SOM. Compound specific analysis demonstrated distinct assemblages of organic compounds were found with increasing thermal stability. These findings together suggest similar thermal activation energies may not equate to similar biological accessibility of the same material and care needs to be taken not to over interpret results from thermal analysis alone. Important follow-up work includes analyzing just the mineral associated OM fraction to better understand this potential disconnect between thermal and biological stability. By coupling evolved gas analysis with radiocarbon and compound specific chemical analyses, new insights into the formation, stabilization and fate of SOM may be possible.

265 **Data availability**

Data from the Waite Permanent Rotation Trial are available for download from the CSIRO Data Access Portal (doi: 10.4225/08/55E5165EC0D29). Radiocarbon and py-GC/MS data are given in appendices B and D.

Author contribution

270 JS conceived the study. JS carried out radiocarbon measurements and interpretation. ASG carried out py-GC/MS measurements and interpretation. JS and ASG contributed equally to manuscript preparation.

Competing interests

The authors declare that they have no conflict of interest.

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References

- 280 de Assis, C. P., González-Pérez, J. A., de la Rosa, J. M., Jucksch, I., Mendonça, E. de S. and González-Vila, F. J.: Analytical pyrolysis of humic substances from a Latosol (Typic Hapludox) under different land uses in Minas Gerais, Brazil, *J. Anal. Appl. Pyrolysis*, 93, 120–128, doi:10.1016/J.JAAP.2011.10.005, 2012.
- Baisden, W. T., Parfitt, R. L., Ross, C., Schipper, L. a. and Canessa, S.: Evaluating 50 years of time-series soil radiocarbon data: Towards routine calculation of robust C residence times, *Biogeochemistry*, 112(1–3), 129–137, doi:10.1007/s10533-285 011-9675-y, 2013.
- Baldock, J. A., Oades, J. M., Nelson, P. N., Skene, T. M., Golchin, A. and Clarke, P.: Assessing the extent of decomposition of natural organic materials using solid-state ¹³C NMR spectroscopy, *Aust. J. Soil Res.*, 35(5), 1061–1084, 1997.
- Baldock, J. A., Sanderman, J., Macdonald, L. M., Puccini, A., Hawke, B. A., Szarvas, S. and MCGowan, J.: Quantifying the allocation of soil organic carbon to biologically significant fractions, *Soil Res.*, 51(8), 561–576, 2013.
- 290 Buurman, P., Peterse, F. and Almendros Martin, G.: Soil organic matter chemistry in allophanic soils: a pyrolysis-GC/MS study of a Costa Rican Andosol catena, *Eur. J. Soil Sci.*, 58(6), 1330–1347, 2007.
- Cécillon, L., Baudin, F., Chenu, C., Houot, S., Jolivet, R., Kätterer, T., Lutfalla, S., Macdonald, A., Oort, F. van, Plante, A. F. and others: A model based on Rock-Eval thermal analysis to quantify the size of the centennially persistent organic carbon pool in temperate soils, *Biogeosciences*, 15(9), 2835–2849, 2018.
- 295 Christensen, B. T.: Physical fractionation of soil and structural and functional complexity in organic matter turnover, *Eur. J. Soil Sci.*, 52(3), 345–353, 2001.
- Cotrufo, M. F., Wallenstein, M. D., Boot, C. M., Deneff, K. and Paul, E.: The Microbial Efficiency-Matrix Stabilization (MEMS) framework integrates plant litter decomposition with soil organic matter stabilization: do labile plant inputs form stable soil organic matter?, *Glob. Chang. Biol.*, 19(4), 988–995, doi:10.1111/gcb.12113, 2013.
- 300 Currie, K. I., Brailsford, G., Nichol, S., Gomez, A., Sparks, R., Lassey, K. R. and Riedel, K.: Tropospheric ¹⁴CO₂ at Wellington, New Zealand: the world's longest record, *Biogeochemistry*, 104(1–3), 5–22, 2011.
- Elliott, E. T. and Cambardella, C. A.: Physical separation of soil organic matter, *Agric. Ecosyst. Environ.*, 34(1–4), 407–419, 1991.

- 305 Golchin, A., Oades, J. M., Skjemstad, J. O. and Clarke, P.: Soil structure and carbon cycling, *Soil Res.*, 32(5), 1043–1068, 1994.
- González-Pérez, J. A., González-Vila, F. J., Almendros, G. and Knicker, H.: The effect of fire on soil organic matter—a review, *Environ. Int.*, 30(6), 855–870, doi:10.1016/J.ENVINT.2004.02.003, 2004.
- Grace, P., Oades, J., Keith, H. and Hancock, T.: Trends in wheat yields and soil organic carbon in the Permanent Rotation Trial at the Waite Agricultural Research Institute, South Australia, *Aust. J. Exp. Agric.*, 35(7), 857, doi:10.1071/EA9950857, 310 1995.
- Grandy, A. S. and Neff, J. C.: Molecular C dynamics downstream: the biochemical decomposition sequence and its impact on soil organic matter structure and function, *Sci. Total Environ.*, 404, 221–446, 2008.
- Grandy, A. S. and Robertson, G. P.: Land-Use Intensity Effects on Soil Organic Carbon Accumulation Rates and Mechanisms, *Ecosystems*, 10(1), 59–74, doi:10.1007/s10021-006-9010-y, 2007.
- 315 Grandy, A. S., Neff, J. C. and Weintraub, M. N.: Carbon structure and enzyme activities in alpine and forest ecosystems, *Soil Biol. Biochem.*, 39(11), 2701–2711, doi:10.1016/J.SOILBIO.2007.05.009, 2007.
- Grandy, A. S., Strickland, M. S., Lauber, C. L., Bradford, M. A. and Fierer, N.: The influence of microbial communities, management, and soil texture on soil organic matter chemistry, *Geoderma*, 150, 278–286, 2009.
- Grant, K. E., Galy, V. V., Chadwick, O. A. and Derry, L. A.: Thermal oxidation of carbon in organic matter rich volcanic 320 soils: insights into SOC age differentiation and mineral stabilization, *Biogeochemistry*, 144(3), 291–304, doi:10.1007/s10533-019-00586-1, 2019.
- Hatcher, P. G., Dria, K. J., Kim, S. and Frazier, S. W.: Modern analytical studies of humic substances, *Soil Sci.*, 166(11), 770–794, 2001.
- Helfrich, M., Flessa, H., Dreves, A. and Ludwig, B.: Is thermal oxidation at different temperatures suitable to isolate soil 325 organic carbon fractions with different turnover?, *J. Plant Nutr. Soil Sci.*, 173(1), 61–66, 2010.
- Hemingway, J. D., Rothman, D. H., Rosengard, S. Z. and Galy, V. V.: An inverse method to relate organic carbon reactivity to isotope composition from serial oxidation, *Biogeosciences*, 14(22), 5099–5114, 2017a.
- Hemingway, J. D., Galy, V. V., Gagnon, A. R., Grant, K. E., Rosengard, S. Z., Soulet, G., Zigah, P. K. and McNichol, A. P.: Assessing the blank carbon contribution, isotope mass balance, and kinetic isotope fractionation of the ramped

- 330 pyrolysis/oxidation instrument at NOSAMS, *Radiocarbon*, 59(1), 179–193, 2017b.
- Hempfling, R. and Schulten, H.-R.: Chemical characterization of the organic matter in forest soils by Curie point pyrolysis-GC/MS and pyrolysis-field ionization mass spectrometry, *Org. Geochem.*, 15(2), 131–145, 1990.
- Jastrow, J. D., Miller, R. M. and Boutton, T. W.: Carbon dynamics of aggregate-associated organic matter estimated by carbon-13 natural abundance, *Soil Sci. Soc. Am. J.*, 60(3), 801–807, 1996.
- 335 Kallenbach, C. M., Grandy, A. S., Frey, S. D. and Diefendorf, A. F.: Microbial physiology and necromass regulate agricultural soil carbon accumulation, *Soil Biol. Biochem.*, 91, 279–290, doi:10.1016/j.soilbio.2015.09.005, 2015.
- Kallenbach, C. M., Frey, S. D. and Grandy, A. S.: Direct evidence for microbial-derived soil organic matter formation and its ecophysiological controls, *Nat. Commun.*, 7, 13630, doi:10.1038/ncomms13630https://www.nature.com/articles/ncomms13630#supplementary-information, 2016.
- 340 Knicker, H.: Soil organic N - An under-rated player for C sequestration in soils?, *Soil Biol. Biochem.*, 43(6), 1118–1129, doi:10.1016/j.soilbio.2011.02.020, 2011.
- Kramer, M. G., Sanderman, J., Chadwick, O. A., Chorover, J. and Vitousek, P. M.: Long-term carbon storage through retention of dissolved aromatic acids by reactive particles in soil, *Glob. Chang. Biol.*, doi:10.1111/j.1365-2486.2012.02681.x, 2012.
- 345 Lehmann, J. and Kleber, M.: The contentious nature of soil organic matter, *Nature*, 528(7580), 60, 2015.
- Lopez-Capel, E., Sohi, S. P., Gaunt, J. L. and Manning, D. A. C.: Use of thermogravimetry--differential scanning calorimetry to characterize modelable soil organic matter fractions, *Soil Sci. Soc. Am. J.*, 69(1), 136–140, 2005.
- von Lützow, M., Kögel-Knabner, I., Ekschmitt, K., Flessa, H., Guggenberger, G., Matzner, E. and Marschner, B.: SOM fractionation methods: Relevance to functional pools and to stabilization mechanisms, *Soil Biol. Biochem.*, 39(9), 2183–2207, doi:10.1016/j.soilbio.2007.03.007, 2007.
- 350 Manzoni, S. and Porporato, A.: Soil carbon and nitrogen mineralization: Theory and models across scales, *Soil Biol. Biochem.*, 41(7), 1355–1379, doi:10.1016/j.soilbio.2009.02.031, 2009.
- McNichol, A. P., Jones, G. A., Hutton, D. L., Gagnon, A. R. and Key, Rm.: The rapid preparation of seawater ¹⁴C₂ for radiocarbon analysis at the National Ocean Sciences AMS facility, *Radiocarbon*, 36(2), 237–246, 1994a.

- 355 McNichol, A. P., Osborne, E. A., Gagnon, A. R., Fry, B. and Jones, G. A.: TIC, TOC, DIC, DOC, PIC, POC unique aspects in the preparation of oceanographic samples for ^{14}C -AMS, *Nucl. Instruments Methods Phys. Res. Sect. B Beam Interact. with Mater. Atoms*, 92(1–4), 162–165, 1994b.
- Miltner, A., Bombach, P., Schmidt-Brücken, B. and Kästner, M.: SOM genesis: Microbial biomass as a significant source, *Biogeochemistry*, 111(1–3), 41–55, doi:10.1007/s10533-011-9658-z, 2012.
- 360 Paul, E. A.: The nature and dynamics of soil organic matter: Plant inputs, microbial transformations, and organic matter stabilization, *Soil Biol. Biochem.*, 98, 109–126, doi:10.1016/j.soilbio.2016.04.001, 2016.
- Paul, E. A., Collins, H. P. and Leavitt, S. W.: Dynamics of resistant soil carbon of Midwestern agricultural soils measured by naturally occurring ^{14}C abundance, *Geoderma*, 104(3–4), 239–256, 2001.
- Peltre, C., Fernández, J. M., Craine, J. M. and Plante, A. F.: Relationships between Biological and Thermal Indices of Soil Organic Matter Stability Differ with Soil Organic Carbon Level, *Soil Sci. Soc. Am. J.*, 77(6), 2020, doi:10.2136/sssaj2013.02.0081, 2013.
- 365 Plante, A. F., Fernández, J. M. and Leifeld, J.: Application of thermal analysis techniques in soil science, *Geoderma*, 153(1–2), 1–10, doi:10.1016/j.geoderma.2009.08.016, 2009.
- Plante, A. F., Beupré, S. R., Roberts, M. L. and Baisden, T.: Distribution of radiocarbon ages in soil organic matter by thermal fractionation, *Radiocarbon*, 55(2), 1077–1083, 2013.
- 370 Poeplau, C., Don, A., Six, J., Kaiser, M., Benbi, D., Chenu, C., Cotrufo, M. F., Derrien, D., Gioacchini, P., Grand, S. and others: Isolating organic carbon fractions with varying turnover rates in temperate agricultural soils--A comprehensive method comparison, *Soil Biol. Biochem.*, 125, 10–26, 2018.
- Rosenheim, B. E., Day, M. B., Domack, E., Schrum, H., Benthien, A. and Hayes, J. M.: Antarctic sediment chronology by programmed-temperature pyrolysis: Methodology and data treatment, *Geochemistry, Geophys. Geosystems*, 9(4), 2008.
- 375 Rothstein, D. E., Toosi, E. R., Schaetzl, R. J. and Grandy, A. S.: Translocation of Carbon from Surface Organic Horizons to the Subsoil in Coarse-Textured Spodosols: Implications for Deep Soil C Dynamics, *Soil Sci. Soc. Am. J.*, 82(4), 969–982, doi:10.2136/sssaj2018.01.0033, 2018.
- Rovira, P., Kurz-Besson, C., Coûteaux, M.-M. and Ramón Vallejo, V.: Changes in litter properties during decomposition: A study by differential thermogravimetry and scanning calorimetry, *Soil Biol. Biochem.*, 40(1), 172–185, doi:10.1016/j.soilbio.2007.07.021, 2008.
- 380

- Ruamps, L. S., Nunan, N., Pouteau, V., Leloup, J., Raynaud, X., Roy, V. and Chenu, C.: Regulation of soil organic C mineralisation at the pore scale, *FEMS Microbiol. Ecol.*, 86(1), 26–35, doi:10.1111/1574-6941.12078, 2013.
- 385 Saiz-Jimenez, C. and De Leeuw, J. W.: Chemical characterization of soil organic matter fractions by analytical pyrolysis-gas chromatography-mass spectrometry, *J. Anal. Appl. Pyrolysis*, 9(2), 99–119, doi:10.1016/0165-2370(86)85002-1, 1986.
- Saiz-Jimenez, C., Haider, K. and Meuzelaar, H. L. C.: Comparisons of soil organic matter and its fractions by pyrolysis mass-spectrometry, *Geoderma*, 22(1), 25–37, doi:10.1016/0016-7061(79)90037-5, 1979.
- Sanderman, J., Fillery, I. R. P., Jongepier, R., Massalsky, a., Roper, M. M., MacDonald, L. M., Maddern, T., Murphy, D. V. and Baldock, J. a.: Carbon sequestration under subtropical perennial pastures II: Carbon dynamics, *Soil Res.*, 51(7–8), 771–
390 780, doi:10.1071/SR12351, 2013.
- Sanderman, J., Baisden, W. T. and Fallon, S.: Redefining the inert organic carbon pool, *Soil Biol. Biochem.*, 92, doi:10.1016/j.soilbio.2015.10.005, 2016.
- Sanderman, J., Creamer, C., Baisden, W. T., Farrell, M. and Fallon, S.: Greater soil carbon stocks and faster turnover rates with increasing agricultural productivity, *SOIL*, 3(1), doi:10.5194/soil-3-1-2017, 2017.
- 395 Schädel, C., Luo, Y., Evans, R. D., Fei, S. and Schaeffer, S. M.: Separating soil CO₂ efflux into C-pool-specific decay rates via inverse analysis of soil incubation data, *Oecologia*, 171(3), 721–732, 2013.
- Schiedung, M., Don, A., Wordell-Dietrich, P., Alcántara, V., Kuner, P. and Guggenberger, G.: Thermal oxidation does not fractionate soil organic carbon with differing biological stabilities, *J. Plant Nutr. Soil Sci.*, 180(1), 18–26, 2017.
- Schmidt, M. W. I., Torn, M. S., Abiven, S., Dittmar, T., Guggenberger, G., Janssens, I. A., Kleber, M., Kögel-Knabner, I.,
400 Lehmann, J., Manning, D. A. C. and others: Persistence of soil organic matter as an ecosystem property, *Nature*, 478(7367), 49, 2011.
- Schulten, H.-R. and Leinweber, P.: Pyrolysis-field ionization mass spectrometry of agricultural soils and humic substances: Effect of cropping systems and influence of the mineral matrix, *Plant Soil*, 151, 77–90, 1993.
- Schulten, H.-R. and Leinweber, P.: New insights into organic-mineral particles: composition, properties and models of
405 molecular structure, *Biol. Fertil. Soils*, 30(5–6), 399–432, doi:10.1007/s003740050020, 2000.
- Schulten, H. R. and Schnitzer, M.: The chemistry of soil organic nitrogen: a review, *Biol. Fertil. Soils*, 26(1), 1–15, doi:10.1007/s003740050335, 1997.

- Skjemstad, J. O., Spouncer, L. R., Cowie, B. and Swift, R. S.: Calibration of the Rothamsted organic carbon turnover model (RothC ver. 26.3), using measurable soil organic carbon pools, *Aust. J. Soil Res.*, 42(1), 79–88, doi:10.1071/SR03013, 2004.
- 410 Sokol, N. W., Sanderman, J. and Bradford, M. A.: Pathways of mineral-associated soil organic matter formation: Integrating the role of plant carbon source, chemistry, and point of entry, *Glob. Chang. Biol.*, 25(1), 12–24, 2019.
- Sollins, P., Kramer, M. G., Swanston, C., Lajtha, K., Filley, T., Aufdenkampe, A. K., Wagai, R. and Bowden, R. D.: Sequential density fractionation across soils of contrasting mineralogy: evidence for both microbial- and mineral-controlled soil organic matter stabilization, *Biogeochemistry*, 96(1–3), 209–231, doi:10.1007/s10533-009-9359-z, 2009.
- 415 Soucémarianadin, L., Cécillon, L., Chenu, C., Baudin, F., Nicolas, M., Girardin, C. and Barré, P.: Is Rock-Eval 6 thermal analysis a good indicator of soil organic carbon lability?--A method-comparison study in forest soils, *Soil Biol. Biochem.*, 117, 108–116, 2018.
- Stuiver, M. and Polach, H. A.: Discussion reporting of 14 C data, *Radiocarbon*, 19(3), 355–363, 1977.
- Torn, M. S., Kleber, M., Zavaleta, E. S., Zhu, B., Field, C. B. and Trumbore, S. E.: A dual isotope approach to isolate soil carbon pools of different turnover times, *Biogeosciences*, 10(12), 8067–8081, doi:10.5194/bg-10-8067-2013, 2013.
- 420 Trumbore, S.: Radiocarbon and soil carbon dynamics, *Annu. Rev. Earth Planet. Sci.*, 37, 47–66, 2009.
- Trumbore, S. E., Chadwick, O. A. and Amundson, R.: Rapid exchange between soil carbon and atmospheric carbon dioxide driven by temperature change, *Science (80-.)*, 272(5260), 393–396, 1996.
- Wickings, K., Grandy, A. S., Reed, S. and Cleveland, C.: Management intensity alters decomposition via biological pathways, *Biogeochemistry*, 104(1–3), 365–379, doi:10.1007/s10533-010-9510-x, 2011.
- 425 Williams, E. K. and Plante, A. F.: A bioenergetic framework for assessing soil organic matter persistence, *Front. Earth Sci.*, 6, 143, 2018.
- Williams, E. K., Rosenheim, B. E., McNichol, A. P. and Masiello, C. A.: Charring and non-additive chemical reactions during ramped pyrolysis: Applications to the characterization of sedimentary and soil organic material, *Org. Geochem.*, 77, 106–114, 2014.
- 430 Williams, E. K., Fogel, M. L., Berhe, A. A. and Plante, A. F.: Distinct bioenergetic signatures in particulate versus mineral-associated soil organic matter, *Geoderma*, 330, 107–116, 2018.

Yang, H., Yan, R., Chen, H., Zheng, C., Lee, D. H. and Liang, D. T.: In-depth investigation of biomass pyrolysis based on three major components: hemicellulose, cellulose and lignin, *Energy & Fuels*, 20(1), 388–393, 2006.

435 Zelles, L., Bai, Q. Y., Beck, T. and Beese, F.: Signature fatty-acids in phospholipids and lipopolysaccharides as indicators of microbial biomass and community structure in agricultural soils, *Soil Biol. Biochem.*, 24(4), 317–323, doi:10.1016/0038-0717(92)90191-y, 1992.

Zimmermann, M., Leifeld, J., Schmidt, M. W. I., Smith, P. and Fuhrer, J.: Measured soil organic matter fractions can be related to pools in the RothC model, *Eur. J. Soil Sci.*, 58(3), 658–667, doi:10.1111/j.1365-2389.2006.00855.x, 2007.

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Table 1. Previously reported organic matter properties measured on the soils used in this study.

Year	SOC (%)	TN (%)	C/N	$\Delta^{14}\text{C}$ (‰)	$\delta^{13}\text{C}$ (‰)	Size Fractions ¹		Adelaide Fractions ²		
						f _{>50μm}	f _{<50μm}	f _{POC}	f _{HOC}	f _{ROC}
1963	2.80	0.25	11.3	16.7	-26.5	0.27	0.73	0.17	0.54	0.29
1973	2.49	0.22	11.2	132.4	-26.0	0.24	0.76	0.16	0.55	0.29
1983	2.48	0.23	10.8	118.4	-26.4	0.23	0.77	0.15	0.55	0.30
1993	2.46	0.24	10.4	95.6	-26.6	0.23	0.77	0.16	0.55	0.28

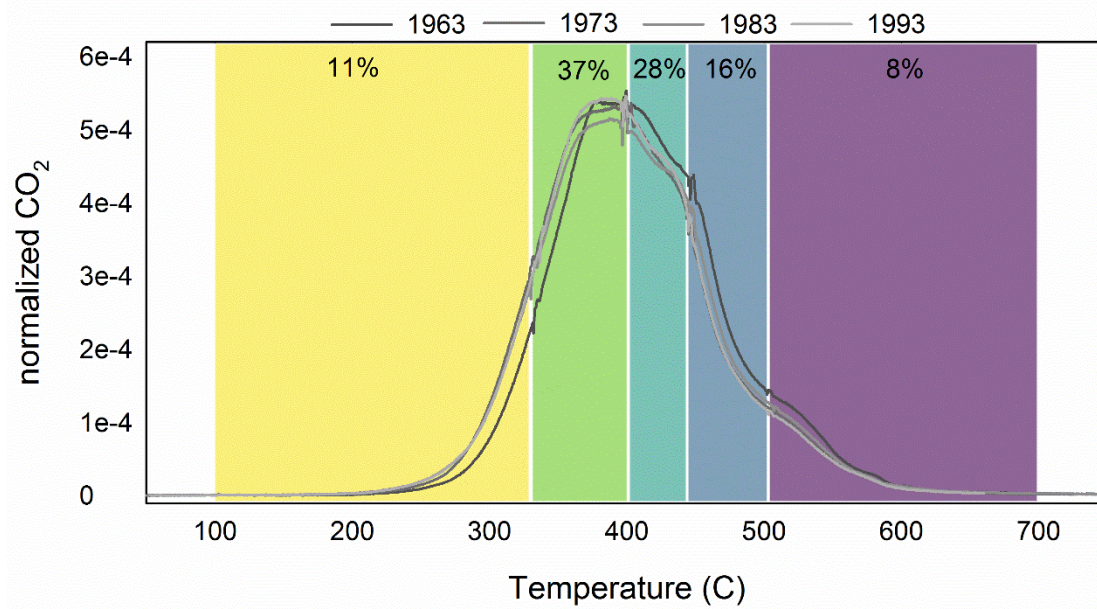
¹measured distribution of SOC into > and < 50 μm size fractions following dispersion and wet sieving

²predicted distribution into particulate, humus and resistant organic carbon (POC, HOC and ROC) fractions (Baldock et al. 2013a) using a mid-infrared spectroscopy based predictive model (Baldock et al. 2013b)

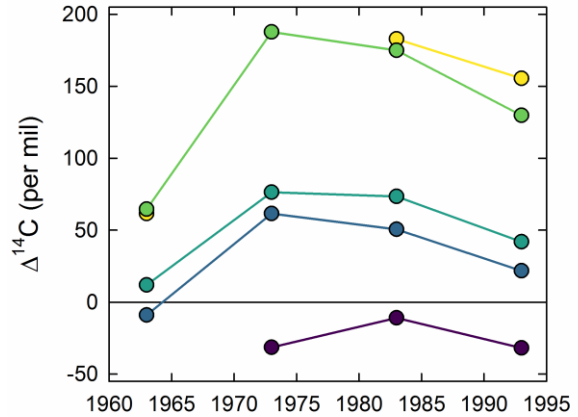
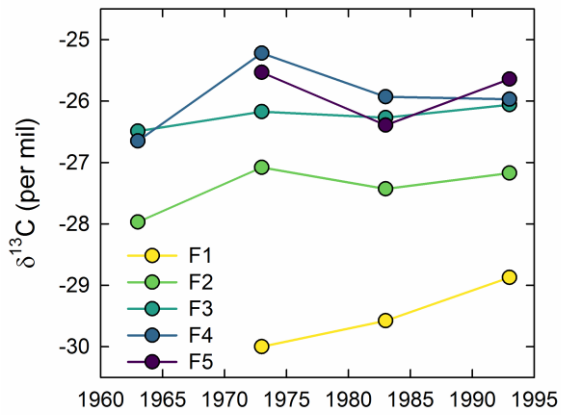
Table 2. Results from bomb-spike soil carbon turnover modeling.

Fraction	E (kJ mol ⁻¹)	<u>One-pool</u>		<u>Two-pool</u>				RMSE
		τ (yrs)	RMSE	f_{fast}	f_{slow}	τ_{fast}	τ_{slow}	
F1	142.0	37.3	28.6	0.38	0.62	4.0	56.16	7.1
F2	155.2	34.1	45.2	0.30	0.70	4.0	65.80	13.8
F3	167.5	99.9	27.1	0.16	0.84	4.0	197.2	7.9
F4	178.1	132.4	24.9	0.16	0.84	4.0	297.3	5.3
F5	198.0	386.3	8.9	0.00	1.00	4.0	390.4	8.7

450



455 **Figure 1. Proportion of total CO₂ evolved with temperature as samples were oxidized with a ramp rate of 5 ° C min⁻¹. Thermal fractions are shaded with colours corresponding to Figures 2-4 and mean percent of total is given for each fraction.**



(a)

(b)

460 **Figure 2. Variation in (a) $\delta^{13}\text{C}$ and (b) $\Delta^{14}\text{C}$ in the five thermal fractions across the four decades of soil sampling. There wasn't enough sample for ^{13}C analysis of F1 or F5 in 1963. Additionally, there was not enough CO_2 for ^{14}C analysis of F5 in 1963. $\Delta^{14}\text{C}$ value from F1 from 1973 not shown in (b) as this data point was removed as an outlier from the turnover modeling.**

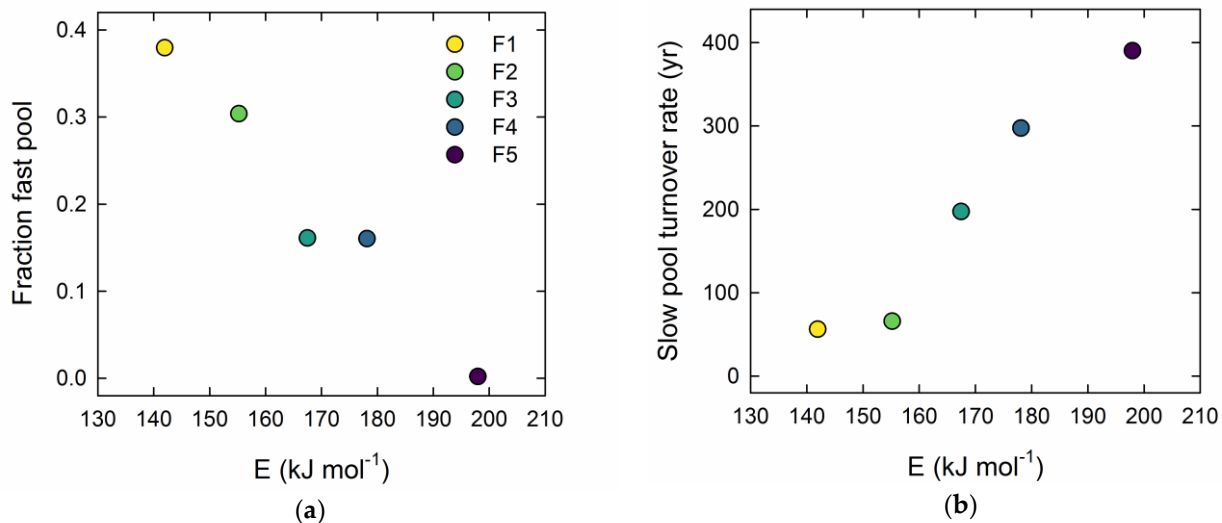
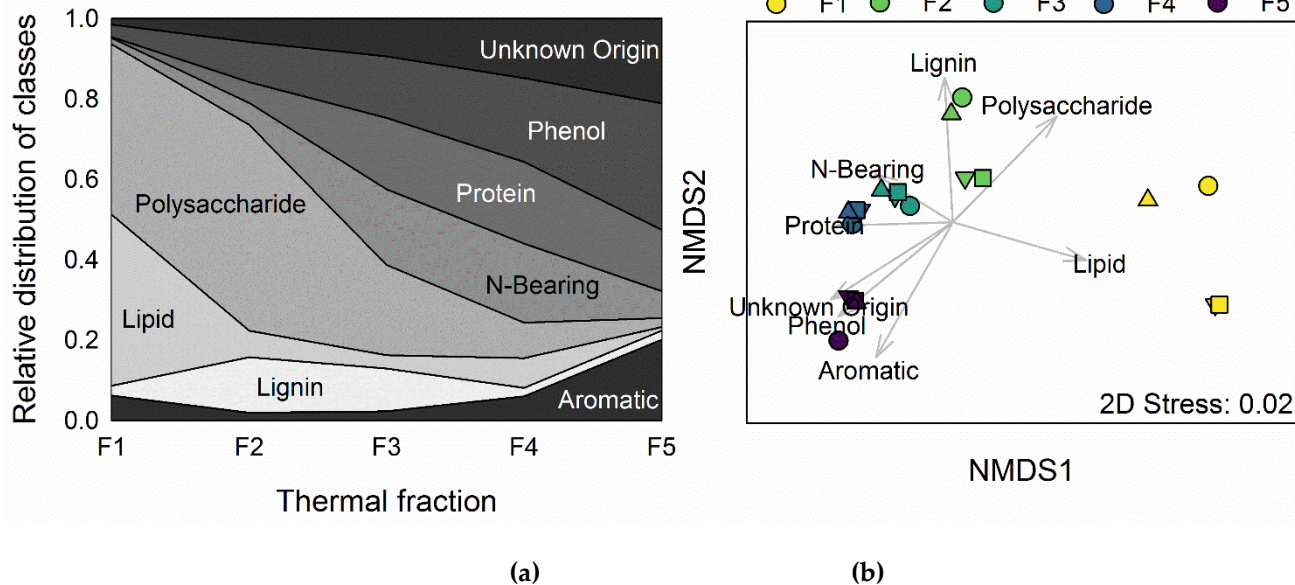


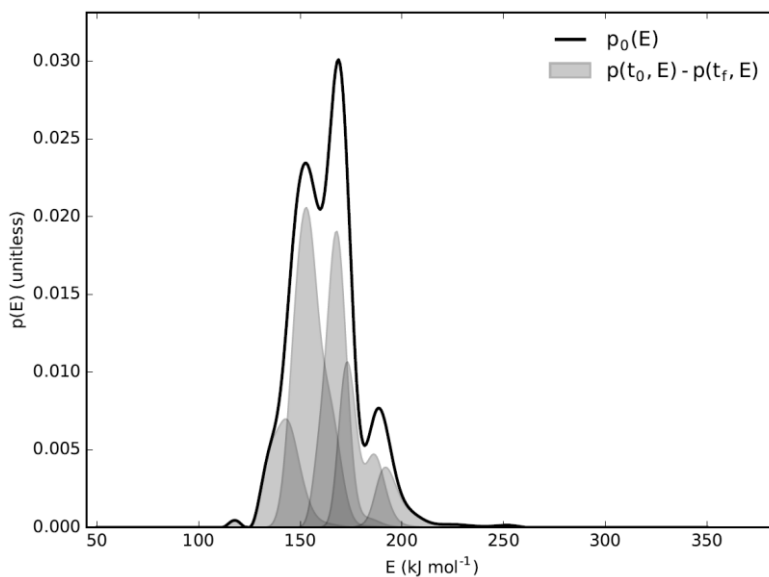
Figure 3. Two-pool ¹⁴C modeling results: (a) proportion of fast cycling carbon in thermal fraction and (b) turnover time (τ) of slow cycling pool plotted as a function of mean activation energy of each thermal fraction.



475 **Figure 4. Stepped pyrolysis gas chromatography-mass spectrometry results: (a) Mean distribution of major compound classes across the four years; (b) Non-metric multidimensional scaling plot of all compounds with > 1% mean abundance (n = 23). In (b) vectors represent correlations with major compound classes and different years are given by different symbols (● 1963, ▼ 1973, ■ 1983, ▲ 1993).**

Appendix A

480 Following acquisition of an initial thermogram (i.e. the trend of CO₂ evolution versus increasing temperature) for the 1973 sample, a mathematical inversion method was employed to deconvolve the evolved gas analysis into N number of pools with distinct activation energy profiles (Hemingway et al. 2017).



485

Figure A1. Proportion of activation energy (E) found with increasing activation energy. Distribution of E within each RPO fraction given by shaded regions.

490 **Table A1. Thermal fraction activation energies and distribution (s.d. = standard deviation) based on inversion analysis.**

Fraction	T0 (° C)	E (kJ/mol)	s.d.
1	100	141.96	8.21
2	325	155.21	7.76
3	400	167.47	5.98
4	445	178.13	8.00
5	515	197.95	13.24

Appendix B

495

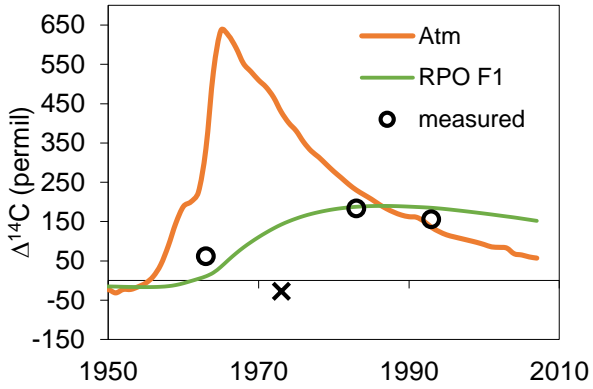
Table B1. Ramped oxidation isotope results.

Year	Fraction	frac of total C ¹	NOSAMS accession #	F modern	Fm Error	δ ¹³ C (‰)	Δ ¹⁴ C (‰)
1963	1	0.08	OS-131571	1.0632	0.0023	n.d.	61.5
	2	0.34	OS-131374	1.0662	0.0021	-27.97	64.5
	3	0.30	OS-131504	1.0135	0.0020	-26.49	11.9
	4	0.18	OS-131505	0.9926	0.0021	-26.65	-9.0
	5	0.10	no sample				
1973	1	0.11	OS-131506	0.9754	0.0033	-30.00	-27.3
	2	0.41	OS-131375	1.1910	0.0024	-27.08	187.7
	3	0.26	OS-131507	1.0793	0.0044	-26.17	76.3
	4	0.16	OS-131508	1.0645	0.0021	-25.22	61.5
	5	0.07	OS-131574	0.9713	0.0026	-25.53	-31.4
1983	1	0.11	OS-131509	1.1876	0.0026	-29.58	182.9
	2	0.37	OS-131376	1.1796	0.0023	-27.43	174.9
	3	0.28	OS-131377	1.0777	0.0026	-26.27	73.4
	4	0.17	OS-131510	1.0548	0.0022	-25.93	50.6
	5	0.07	OS-131573	0.9930	0.0026	-26.39	-11.0
1993	1	0.14	OS-131511	1.1615	0.0024	-28.87	155.5
	2	0.36	OS-131378	1.1357	0.0025	-27.17	129.8
	3	0.28	OS-131707	1.0473	0.0027	-26.06	41.9
	4	0.15	OS-131708	1.0271	0.0020	-25.97	21.8
	5	0.07	OS-131783	0.9732	0.0024	-25.64	-31.8

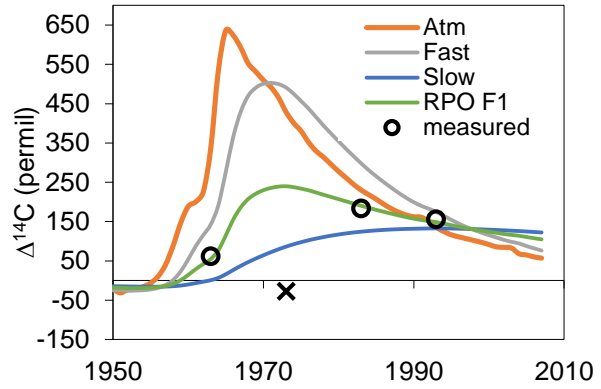
¹proportion of total pCO₂ found in each thermal split (i.e. fraction)

Appendix C

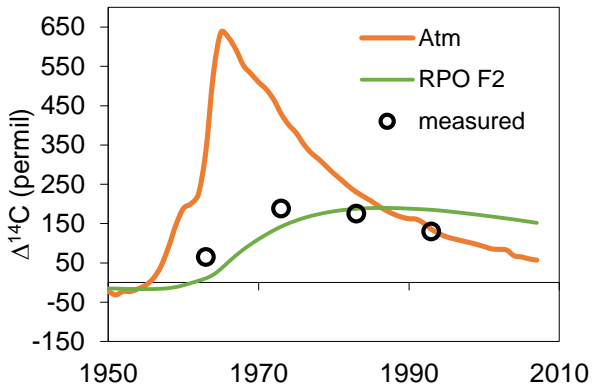
Bomb-spike turnover modeling results for one-pool and two-pool models.



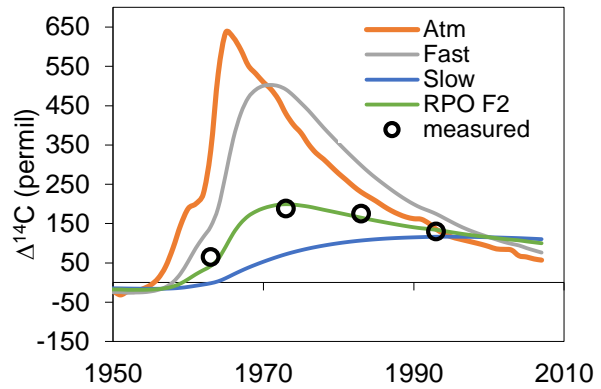
(a)



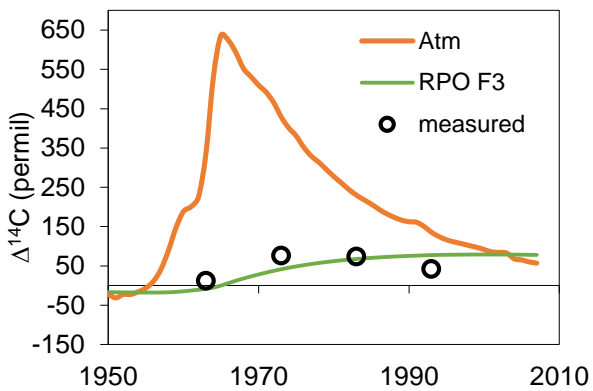
(b)



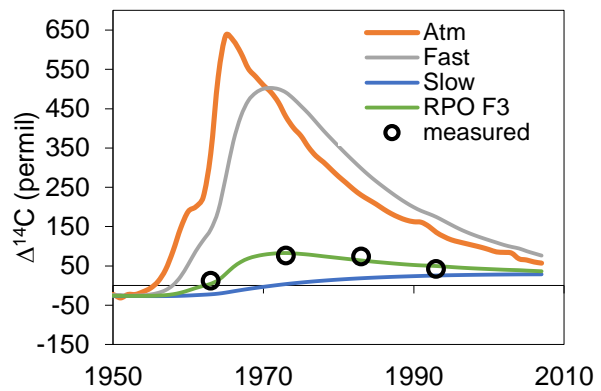
(c)



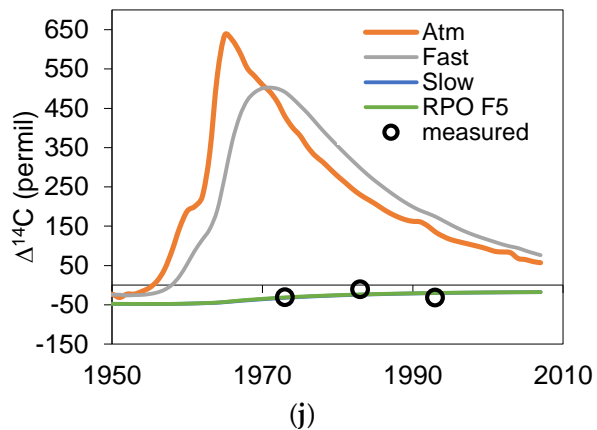
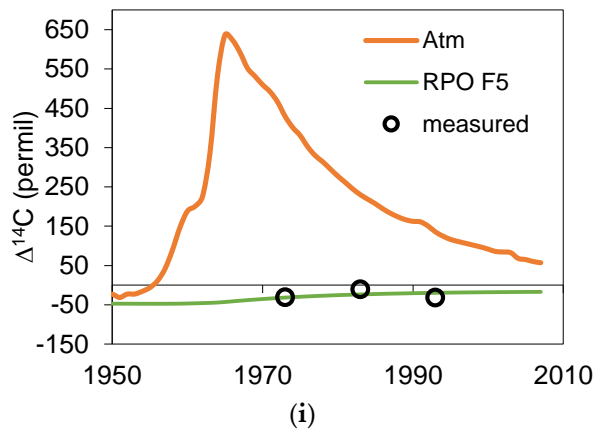
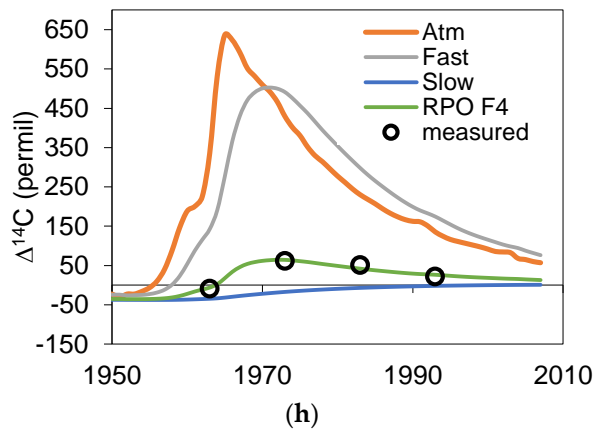
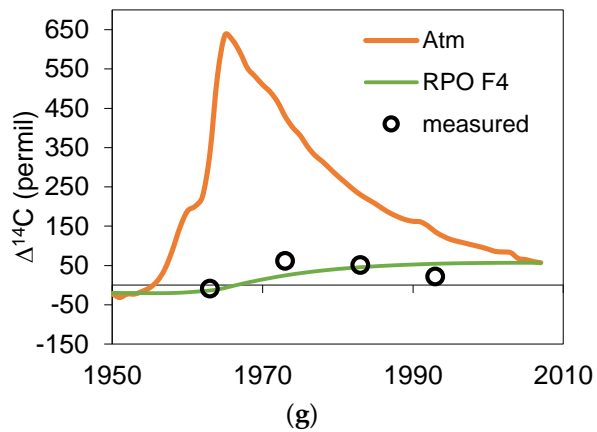
(d)



(e)



(f)



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Figure C1. One-pool (a, c, e, g, i) and two-pool (b, d, f, h, j) solutions to a steady-state soil carbon turnover model for the five thermal fractions. Southern hemisphere atmospheric record given in the background of each panel. For two-pool solutions the trend in $\Delta^{14}\text{C}$ are given for the fast and slow pools as well as the overall thermal fraction. The X in panels (a) and (b) refers to the $^{14}\text{CO}_2$ measurement deemed to be an outlier for purposes of bomb-spike modeling.

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Appendix D

Table D1. Most abundant compound classes (mean across years) identified by stepped Py-GC/MS.

Compound	Source	F1	F2	F3	F4	F5
Phenol, 3,4-dimethyl-	Aromatic	0.3	0.8	1.5	2.6	2.0
Fluorene	Aromatic	0.0	0.0	0.0	0.3	6.0
Naphthalene	Aromatic	0.0	0.0	0.0	0.8	5.0
Phenol, 2-methoxy- (Guaiacol)	Lignin	0.9	6.1	5.4	1.0	0.0
Hexadecanoic acid, methyl ester (Palmitic acid-C16)	Lipid	41.5	4.9	3.1	0.4	0.0
Pyrazolo[5,1-c][1,2,4]benzotriazin-8-ol	N-Bearing	0.0	1.3	11.7	13.0	1.2
1H-Pyrrole, 3-methyl-	N-Bearing	0.0	0.9	1.9	2.5	0.6
Phenol	Phenol	1.0	6.9	9.1	10.5	16.7
Phenol, 4-methyl-	Phenol	2.1	3.1	6.2	10.3	14.6
Furfural	Polysaccharide	16.0	13.4	4.1	0.2	0.0
3-Furaldehyde	Polysaccharide	10.2	8.3	0.9	0.0	0.0
Levoglucosenone	Polysaccharide	9.0	9.2	1.1	0.0	0.0
Benzofuran, 2,3-dihydro-	Polysaccharide	0.9	7.3	6.0	2.2	1.1
Furfural, 5-methyl-	Polysaccharide	5.7	7.8	3.1	0.5	0.0
Indole	Protein	0.2	2.5	5.0	3.7	4.1
Pyrrole	Protein	0.0	1.0	3.3	3.4	0.4
Benzyl nitrile	Protein	0.0	0.5	3.0	3.4	0.3
3-Methylindole	Protein	0.0	0.4	1.5	2.3	2.8
Styrene	Protein	0.0	0.0	0.9	3.6	2.2
Toluene	Unknown origin	0.0	0.5	2.2	5.8	5.9
1,3,5-Cycloheptatriene	Unknown origin	0.4	2.8	3.4	0.7	0.0
Phenanthrene	Unknown origin	0.1	0.0	0.0	0.1	6.9
Monobenzene	Unknown origin	0.0	0.9	2.6	2.2	0.0
% of all compound abundance.....		88.2	78.4	76.1	69.6	69.7