



1 **N₂O and N₂ losses from simulated injection of biogas digestate**
2 **depend mainly on soil texture, moisture and temperature**

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10 **Abstract.** Biogas digestate (BD) is increasingly used as organic fertiliser, but has a high potential for NH_3 losses.
11 Its proposed injection into soils as a counter-measure has been suggested to promote the generation of N_2O , leading
12 to a potential trade-off. Furthermore, the effect on N_2 losses after injection of BD into soil has not yet been
13 evaluated. We performed a simulated BD injection experiment in a helium-oxygen atmosphere to examine the
14 influence of soil substrate (loamy sand, clayey silt), water-filled pore space (WFPS; 35, 55, 75%), temperature
15 (2°C , 15°C) and application rate (0, 160, 320 kg N ha^{-1}) as a proxy for row spacing of injection on the emissions
16 of N_2O , N_2 , and CO_2 . To determine the potential capacity for these gaseous losses, we incubated under anaerobic
17 conditions by purging with helium for the last 24 h of incubation. N_2O and N_2 emissions as well as the $\text{N}_2/(\text{N}_2\text{O}+\text{N}_2)$
18 ratio depended on soil type and increased with WFPS and temperature, indicating a crucial role of soil gas
19 diffusivity for the formation of these gases in agricultural soils. However, the emissions did not increase with the
20 application rate of BD, i.e. a broader spacing of injection slits, probably due to an inhibitory effect of the high
21 NH_4^+ content of BD. Our results suggest that the risk of N_2O and N_2 losses even after injection of relatively large
22 amounts of BD seems to be small for dry to wet sandy soils and acceptable when regarding simultaneously reduced
23 NH_3 emissions for dry silty soils.

24 1 Introduction

25 Nitrous oxide (N_2O) is a potent greenhouse gas (Myhre et al., 2013), with agriculture being the largest single
26 source of anthropogenic N_2O emissions, contributing about 4.1 Tg $\text{N}_2\text{O-N yr}^{-1}$ or 66% of total gross anthropogenic
27 emissions mainly as a result of mineral nitrogen (N) fertiliser and manure application (Davidson and Kanter, 2014).
28 The generation of nitrogen gas (N_2) is of agronomic interest in terms of nutrient management, since these gaseous
29 losses may imply a significant loss of N from the soil/plant system (Friedl et al., 2016; Cameron et al., 2013).
30 However, from an environmental stance, N_2 is innocuous and, thus, the preferred type of gaseous N-loss from soil
31 (Davidson et al., 2015). Further, emission and deposition of ammonia (NH_3) is of environmental concern, e.g.,
32 through acidification and conversion to N_2O (Ferm, 1998; Mosier et al., 1998). In general, the improvement of N
33 use efficiency and thus the decrease of N losses in crop production are paramount in the presence of challenges
34 like food security, environmental degradation and climate change (Zhang et al., 2015).
35 In Germany, the increased demand for renewable energy sources like methane from biogas plants entails an
36 expanded amount of digestion residues (biogas digestate, BD) used as organic amendment in agriculture (Möller
37 and Müller, 2012). Digestion in biogas reactors increases pH and the proportion of ammonium (NH_4^+) and narrows



38 the C to N ratio due to the depletion of labile C fractions of the feedstock (Möller and Müller, 2012). Compared
39 to undigested amendments like slurry, these altered chemical properties may promote biochemical reactions in the
40 soil that are responsible for the formation of gaseous N species like N_2O , N_2 and especially NH_3 (Nkoa, 2013). In
41 general, the overall effect of BD on gaseous N losses from soil is still under debate (Möller, 2015).
42 Significant losses of N as NH_3 can occur within the first hours after manure application (Quakernack et al., 2012).
43 To reduce NH_3 losses, the application of BD by injection is recommended. But this measure can simultaneously
44 increase the potential for N_2O losses compared to surface-application due to a reduction of local O_2 availability
45 (Wulf et al., 2002; Velthof and Mosquera, 2011; Webb et al., 2010) and the occurrence of high nutrient
46 concentrations in the injection band (Dell et al., 2011). Hence, these conditions during the initial phase after
47 injection of BD foster microsites favourable for microbial denitrification, which may promote also the formation
48 of N_2 due to anaerobic conditions (Köster et al., 2015).
49 There is a wealth of biotic and abiotic processes in soils that produce N_2O and N_2 , most of which are enhanced by
50 anaerobic or at least hypo-aerobic conditions (Butterbach-Bahl et al., 2013). Also the amounts and the relative
51 share of N_2 and N_2O in the overall gaseous N emissions depend – among other factors like the favoured reduction
52 of NO_3^- rather than N_2O as alternative electron acceptor – on the degree of O_2 restriction (Firestone and Davidson,
53 1989). Soil physical and biotic factors (i.e. diffusion and consumption of O_2) as well as their interactions control
54 the aerobic status of a soil. Diffusion of O_2 depends on the porosity of the soil substrate in conjunction with water-
55 filled pore space (WFPS), while O_2 is consumed by heterotrophic respiration which depends on mineral N content,
56 carbon (C) availability as well as on temperature (Ball, 2013; Uchida et al., 2008; Maag and Vinther, 1999).
57 Simultaneously, the supply of substrates for microorganisms is determined by liquid diffusion rates in soil water
58 and, thus, by WFPS (Blagodatsky and Smith, 2012; Maag and Vinther, 1999). However, nutrient concentrations
59 and WFPS should theoretically be controlled especially by the row spacing between the injection bands, since for
60 a given amount of BD per area, a wider spacing requires a higher concentration of BD application in the band and
61 *vice versa* with consequences for microbial activity and O_2 availability within the band. There are studies that
62 examined the effect of manure injection depth on N_2O emissions (Webb et al., 2010), but we are not aware of
63 studies on the effect of row spacing in general as well as application rate as a proxy in particular.
64 Hence, there is a general lack of knowledge about effects of manure injection on gaseous N-losses and especially
65 about the effects of BD and row spacing and how they interact with O_2 limiting factors like soil texture and WFPS,
66 as well as temperature and heterotrophic respiration. These knowledge gaps are caused not least by methodological
67 issues with the determination of N_2 fluxes. Thus, we applied the helium-oxygen ($He-O_2$) incubation technique



68 (Butterbach-Bahl et al., 2002) in a laboratory experiment to evaluate the effect of above suggested factors on the
69 generation of N₂O and N₂ after simulated injection of BD into soils. Simultaneously, CO₂ flux was determined as
70 an indicator for microbial O₂ consumption, O₂ diffusion and also for the degradability of organic C applied with
71 BD (Blagodatsky and Smith, 2012). We hypothesised that (1) N₂O and N₂ emissions will increase with WFPS and
72 temperature, (2) these gaseous N losses will also be affected by BD application rate, i.e. injection row-spacing,
73 and (3) the fine textured clayey silt will induce higher gaseous N losses than the coarse loamy sand.

74 2 Material and Methods

75 2.1 Selected soils, sampling of soil cores and biogas digestate

76 Two soils were selected and both were treated with three levels of WFPS and three quantities of BD (Table 1),
77 resulting in 18 factor combinations with three repetitions each. Temperature was increased from 2 °C during the
78 first two days to 15 °C for the last three days of the incubation. Intact soil cores (diameter 0.072 m, height 0.061
79 m) were taken with sample rings in the range from 0–0.10 m depth from two sites with different textures, i.e. sandy
80 loam and clayey silt. The sandy loam samples were gathered from a stagnic luvisol (IUSS Working Group WRB,
81 2006) located in Gülzow (North-East Germany) in the ground moraine of the Weichselian glacial period at 53° 48'
82 35" N and 12° 4' 20" E. The clayey silt samples were gathered from a haplic luvisol located in Dornburg between
83 the foothills and the lowlands of Central Germany at 51° 0' 8" N and 11° 39' 25" E (see Table 2 for more details
84 on soil characteristics). After field sampling, the soil cores were dried for 48 h at 40 °C.

85 Both sites have been cultivated with similar crop rotations used as feedstock for biogas production and have been
86 amended with biogas digestate for the past nine years. The crop rotation on the sandy loam consisted of maize
87 (*Zea mays* L.), rye (*Secale cereale* L.), sorghum (*Sorghum bicolor* (L.) MOENCH), winter triticale (×
88 *Triticosecale* Wittmack), ryegrass (*Lolium perenne* L.) and winter wheat (*Triticum aestivum* L.). The only
89 difference in the crop rotation on the clayey silt was the cultivation of sudangrass (*Sorghum × drummondii*)
90 instead of sorghum.

91 The biogas digestate used for the incubation was obtained from a biogas plant at 'Gut Dalwitz', an organic farm
92 in northeast Germany. The feedstock for the anaerobic fermentation in the plant consisted of 60 % maize, 20 %
93 solid cattle manure, 10 % dry chicken manure and 10 % rye. The digestate was analysed by 'LUFA', Rostock,
94 Germany and had a pH of 8.3, 2.91% organic C, 0.16% dissolved organic C (DOC), 0.54% N and 0.27% NH₄-N
95 in undried material with a dry matter content of 9.4%.



96 **2.2 Adjustment of WFPS and addition of N**

97 For adjustment of WFPS, the dry and undisturbed soil cores were moistened dropwise with the respective quantity
98 of water calculated based on the bulk density and an assumed particle density of 2.65 g cm^{-3} , reduced by the
99 expected moisture input from subsequent addition of BD. Since injection bands *in situ* have a thickness comparable
100 to the sample rings we used, the soil cores were mixed with the biogas digestate and finally repacked to simulate
101 the concentration of BD in soil after injection. The amounts of added BD were calculated with an assumed injection
102 of 160 kg N ha^{-1} into soil with row spaces of 0.15 and 0.30 m, which are common ranges used by injection
103 machinery. The application rate per band of 160 kg N ha^{-1} at 0.30 m spacing would correspond to the concentration
104 of BD resulting from the injection of 320 kg N ha^{-1} at 0.15 m spacing. Thus, for convenience, we will denote the
105 different levels of BD by amount based on a row spacing of 0.15 m. After this procedure, the soil cores were sealed
106 with plastic lids and stored immediately at $2 \text{ }^{\circ}\text{C}$ until the beginning of the incubation within a week.

107 **2.3 Determination of gas fluxes**

108 The measurements of N_2 , N_2O and CO_2 fluxes were applied following the He- O_2 method (Scholefield et al., 1997;
109 Butterbach-Bahl et al., 2002). Six soil cores (i.e. the repetitions of two factor combinations at a time, Table 3) were
110 placed simultaneously in special gas-tight incubation vessels inside a climate chamber. Analyses were conducted
111 in the laboratory of the Institute for Landscape Biogeochemistry, Leibniz Centre for Agricultural Landscape
112 Research (ZALF), Müncheberg, Germany. Before flux measurements, the vessels were evacuated moderately
113 (0.047 bar) and flushed with an artificial He/ O_2 gas mixture (20.49 % O_2 , 345.5 ppm CO_2 , 359 ppb N_2O , 1863 ppb
114 CH_4 , 2.46 ppm N_2 , rest He) four times consecutively to remove ambient N_2 . Subsequently, the air temperature of
115 the climate chamber was set to $2 \text{ }^{\circ}\text{C}$ and a continuous He/ O_2 gas flow rate of 15 ml min^{-1} was applied to the vessel
116 headspaces for 72 h to remove residues of N_2 from soil cores. After this pre-incubation, during the following two
117 days, the headspace concentration of N_2O and CO_2 was measured once daily in the morning. To compensate for a
118 possibly lower precision of the detector for N_2 in relation to the detector for N_2O and CO_2 , N_2 concentrations were
119 measured consecutively three times daily in the morning. Immediately after the last measurement on the second
120 day, the temperature was set to $15 \text{ }^{\circ}\text{C}$ and the measurements were continued for another two days. Finally, the
121 He/ O_2 gas mixture was substituted by pure He and, following 24 h of acclimatisation, gas measurements were
122 carried out once again (Figure 1) to determine the current potential for N_2O and N_2 generation in a completely



123 anaerobic soil matrix. The settings of the chromatographs for gas analyses are described in Eickenscheidt et al.
124 (2014). Gas fluxes were calculated according to Eq. (1):

$$125 \quad f = \frac{M \times p \times v \times dc}{R \times T \times A}, \quad (1)$$

126 where f is the flux (N_2 and CO_2 : $\text{mg m}^{-2} \text{h}^{-1}$, N_2O : $\mu\text{g m}^{-2} \text{h}^{-1}$), M the molar mass in g mol^{-1} (N_2 : 28, CO_2 : 44, N_2O :
127 44), p the air pressure (Pa), v the air flow (L h^{-1}), R the gas constant ($8.31 \text{ J mol}^{-1} \text{ K}^{-1}$), T the temperature inside the
128 chamber (K), A the area of the incubation vessel (m^2), and dc the difference of gas concentrations (N_2 and CO_2 :
129 ppm, N_2O : ppb).

130 The lowest detectable fluxes were checked every day for each vessel and were on average 0.427 ($1\sigma = 0.271$) mg
131 $\text{N}_2\text{-N m}^{-2} \text{h}^{-1}$, 3.6 (3.1) $\mu\text{g N}_2\text{O-N m}^{-2} \text{h}^{-1}$ and 0.918 (0.693) $\text{mg CO}_2\text{-C m}^{-2} \text{h}^{-1}$. Estimated fluxes from the soil cores
132 smaller than the respective detection limits of each day were set to zero.

133 **2.4 Soil analyses after incubation**

134 After incubation, the soil cores were stored at 2°C until they were extracted with 0.1 M KCl solution (soil to
135 extract ratio 1:4) and analysed for NH_4^+ and nitrate (NO_3^-) by spectrophotometry according to DIN ISO 14256
136 with a continuous flow analyser ‘CFA-SAN’, Skalar Analytical B.V., the Netherlands and for DOC by combustion
137 according to DIN ISO 10694 with an analyser ‘RC 612’, Leco Instruments GmbH, Germany.

138 **2.5 Statistical analysis**

139 All statistical analyses were done using R version 3.2.3 (R Core Team, 2015) with the data of the consecutive
140 measuring days two and four (2 and 15°C , respectively, under He-O_2 atmosphere) to allow for establishment of a
141 new flux equilibrium on day three after temperature change and, thus, minimising the effect of time. Data from
142 the vessels with the factor combination of 35% WFPS and 160 kg N ha^{-1} with clayey silt had to be omitted due to
143 technical reasons during sample preparation. For the final period of pure He headspace, some gas concentration
144 data are missing due to logistical reasons. For the loamy sand this affects all WFPS levels with 160 kg N ha^{-1} (N_2
145 and N_2O), the treatment 75% WFPS with 320 kg N ha^{-1} (N_2O and CO_2) and for the clayey silt the treatment 35%
146 WFPS without amendment (N_2O and CO_2).

147 To account for repeated measurement of vessels, linear mixed effect models were applied with package ‘lmerTest’
148 (Kuznetsova et al., 2015) for fluxes of each gas type. The three pseudo-replicated fluxes from the N_2 measurements
149 of each vessel were averaged for each day to obtain the same number of observations as for N_2O and CO_2 fluxes.



150 The fixed structure of models included soil type, WFPS, amount of digestate, temperature, NO_3^- contents after
151 incubation as well as the fluxes of N_2O (in the model for N_2) and CO_2 (in the models for N_2 , N_2O and $\text{N}_2/[\text{N}_2+\text{N}_2\text{O}]$
152 product ratio). Soil NH_4^+ and DOC were omitted since they showed high autocorrelation with the amount of BD
153 applied. The individual soil core was set as random effect with regard to lack of independence of consecutive
154 measurements. The model responses for N_2 , N_2O and CO_2 where log transformed ($\ln[\text{value} + 1]$) since these gas
155 fluxes from soils usually show lognormal distributions (Kaiser et al., 1998). The function ‘step’ was used for
156 automatic backward selection of models based on AIC (Akaike’s ‘An Information Criterion’). Shapiro-Wilk
157 normality test ($\alpha = 0.05$) was applied to check residuals for normal distribution. Three outliers, identified by ‘Scale-
158 Location’ plots, were omitted in each model for N_2 as well as for N_2O and five outliers were discarded in the model
159 for $\text{N}_2/(\text{N}_2+\text{N}_2\text{O})$ product ratio to obtain normality. For mixed effect models, the p -values of the summaries were
160 calculated with t -test based on Satterthwaite’s approximations.

161 Cumulated gas fluxes were estimated with a bootstrap method using function ‘auc.mc’ of R package ‘flux’ version
162 0.3-0 (Jurasiński et al., 2012) for the R statistical software version 3.2.3 (R Core Team, 2015). In short, the fluxes
163 for the period of aerobic headspace were cumulated in 100 iterations, while for each run 2 fluxes were omitted
164 randomly. Then, the resulting data were used to calculate means and standard deviations.

165 3 Results

166 3.1 Soil NH_4^+ , NO_3^- and DOC contents

167 The calculated application of NH_4^+ -N from BD per kg soil approximated for the sandy loam 247.0 mg (160 kg N
168 ha^{-1}) and 494.0 mg (320 kg N ha^{-1}), and for the clayey silt 266.0 mg (160 kg N ha^{-1}) and 532.0 mg (320 kg N ha^{-1}).
169 The NO_3^- content of BD was negligible. After incubation, the recovered NH_4^+ -N contents increased with the
170 level of amendment with BD in both soils and were not affected by WFPS, with the exception of treatments of
171 clayey silt with 35% WFPS (Fig. 2). In the loamy sand, the mean amounts of NH_4^+ -N per kg soil ranged from 8.5
172 to 10.0 mg (no amendment), from 170.4 to 185.6 mg (160 kg N ha^{-1}) and from 273.7 to 314.0 mg (320 kg N ha^{-1}).
173 In the clayey silt, NH_4^+ -N contents per kg soil reached only 1.8 to 8.8 mg (no amendment), 89.7 to 98.9 mg (160
174 kg N ha^{-1}) and 146.8 to 194.0 mg (320 kg N ha^{-1}) and, thus, roughly half the amounts of the clayey silt. However,
175 in contrast to the loamy sand, the clayey silt showed also substantial NO_3^- contents between 25.7 (35% WFPS
176 without amendment) and 49.8 mg NO_3^- -N (kg soil^{-1}) (55% WFPS with 160 kg N ha^{-1}). Negligible amounts of NO_3^-



177 were detected in the loamy sand after incubation: except for a mean value of 2.4 mg NO₃⁻-N (kg soil)⁻¹ from the
178 unamended treatment with 75% WFPS, the values of all other treatments ranged between 0.2 and 0.5 mg.
179 The amounts of recovered DOC increased with the application rate of BD, but with different magnitudes for both
180 soils. While mean values from 38.6 (55 % WFPS without amendment) to 500.1 mg DOC per kg soil (75 % WFPS,
181 320 kg N ha⁻¹) were determined for the loamy sand after incubation, lower mean values from 18.9 (55 % WFPS
182 without amendment) to 358.1 mg (35 % WFPS, 320 kg N ha⁻¹) were found in the clayey silt, where the respective
183 second highest values were considerably lower for both soils (loamy sand: 362.2 mg for 75 % WFPS with 160 kg
184 N ha⁻¹, clayey silt: 105.9 mg for 75 % WFPS with 320 kg N ha⁻¹) (Table 4).

185 3.2 N₂O fluxes

186 The mean N₂O fluxes from the loamy sand at 2 °C under the He-O₂-atmosphere were virtually zero and, thus,
187 negligible (Fig. 3, Day 2 in Table A1). This was similar at 15°C with the exception of 35% WFPS without digestate
188 (0.1 mg N₂O-N m⁻² h⁻¹, Fig. 3, Day 4 in Table A1). The clayey silt showed much larger fluxes than the loamy sand:
189 even at 2 °C, up to 1.5 mg N₂O-N m⁻² h⁻¹ were detected (55% WFPS with 160 kg N ha⁻¹). After shifting the
190 temperature to 15 °C, the same factor combination had a mean flux of 6.2 mg N₂O-N m⁻² h⁻¹ and the other treatment
191 means emitted between 1.0 and 3.0 mg N₂O-N m⁻² h⁻¹ with the exception of incubations with 35% WFPS, where
192 fluxes were smaller. The sand showed weak N₂O emissions, independent of temperature and WFPS as well as the
193 amount of BD application. In contrast, the emissions of the clayey silt increased with temperature and were highest
194 with intermediate WFPS and amount of BD, i.e. 55% and 160 kg N ha⁻¹, respectively. Surprisingly, at 15 °C,
195 increasing the amount of BD up to 320 kg N ha⁻¹ did not increase the observed N₂O efflux; rather it decreased the
196 efflux significantly ($p < 0.05$, Tuckey's HSD) at 55% and also, but not significantly, at 75% WFPS (Fig. 3, Table
197 A1). However, this effect was not noticed at 35% WFPS due to generally low emissions at this moisture level.
198 According to the linear mixed model for N₂O fluxes in aerobic conditions, WFPS, amount of digestate,
199 temperature, NO₃⁻ content of soil after incubation and CO₂ fluxes had significant ($p < 0.001$) effects on N₂O flux
200 (Table 5).

201 Under anaerobic headspace conditions, the overall highest mean N₂O flux was observed from the clayey silt at
202 35% WFPS and 320 kg N ha⁻¹ (11.7 mg N₂O-N m⁻² h⁻¹). The same soil showed a tendency of decreasing N₂O
203 fluxes with increasing WFPS. Fluxes were largest with an amendment of 160 kg N ha⁻¹. In the loamy sand, the
204 pure He-atmosphere induced increasing mean N₂O fluxes (up to 1.3 mg N₂O-N m⁻² h⁻¹) with increasing WFPS
205 (Fig. 3, Table A1). So, the anaerobic headspace induced a change only in the loamy sand by increasing emissions.



206 3.3 N₂ fluxes

207 From the loamy sand, no N₂ fluxes were detected at 2 °C under He-O₂-atmosphere (Fig. 4, Day 2 in Table A2).
208 Under the same conditions, the clayey silt showed mean fluxes from zero (all incubations with 35% WFPS) to 1.4
209 mg N₂ m⁻² h⁻¹ (all incubations with 75% WFPS). After increasing the temperature to 15 °C, again, the sandy loam
210 released mostly negligible amounts of N₂, except for 0.5 mg m⁻² h⁻¹ with 55% WFPS and 320 kg N ha⁻¹ (Fig. 4,
211 Day 4 in Table A2). In contrast, up to 3.8 mg N₂ m⁻² h⁻¹ (75% WFPS with 160 kg N ha⁻¹) were detected in the
212 clayey silt, where, however, also no fluxes were detected in all incubations with 35% WFPS. Put simply,
213 temperature had a small effect on N₂ emissions from the sandy loam, but WFPS and amount of BD showed no
214 consistent influence. In contrast, the clayey silt presented clearly increasing emissions with increasing temperature,
215 WFPS and also with the application of BD, where a raise from 160 up to 320 kg N ha⁻¹ at 15 °C, however, resulted
216 in slightly, but not significantly ($p > 0.05$, Tuckey's HSD), decreased fluxes (Fig. 4, Table A2). The summary of
217 the linear mixed model for N₂ fluxes under aerobic conditions revealed that soil type, WFPS and N₂O flux had
218 significant ($p < 0.001$) effects on N₂ flux (Table 5).
219 After switching the atmosphere to pure He, the N₂ fluxes from the sandy loam increased more than 60-fold. In
220 contrast to aerobic conditions, all measured factor combinations showed mean fluxes from 3.3 (35% WFPS
221 without N) to 35.1 mg N₂ m⁻² h⁻¹ (55% with 320 kg N ha⁻¹), where the fluxes from amended treatments were always
222 higher than fluxes from the unamended ones (Fig. 2, Day 5 in Table A2). For the clayey silt, compared with aerobic
223 atmosphere, mean fluxes increased slightly to 1.9 mg N₂ m⁻² h⁻¹ in unamended treatments and more remarkably to
224 9.3 mg N₂ m⁻² h⁻¹ in amended ones, still not reaching the amounts observed for the sandy loam. This implies that
225 the N₂ emissions were increased from both soils under anaerobic headspace conditions, but the loamy sand
226 exhibited a much more intense reaction.

227 3.4 N₂/(N₂ + N₂O-N) product ratio

228 No clear trend of the product ratio of N₂/(N₂ + N₂O-N) was found for incubations of the loamy sand. However,
229 there was a clear distinction of the ratios for this soil under aerobic and anaerobic atmospheres: while the ratios
230 were close to zero in the former, they were close to 1 in the latter (Fig. 5). In contrast, in the clayey silt the ratios
231 increased with WFPS and were affected by digestate amendment under both the aerobic and the anaerobic
232 atmospheres, where the highest ratios (up to 0.8) were found in treatments without digestate and at least 55%
233 WFPS. The digestate-amended treatments showed mostly ratios around or above 0.5, with exception of the 35%



234 WFPS treatments, which had ratios close to zero. According to the linear mixed model, the product ratio under
235 aerobic conditions was affected significantly ($p < 0.001$) by soil type, amount of digestate and temperature (Table
236 5).

237 3.5 CO₂ fluxes

238 CO₂ fluxes showed clear differences between the soils: under all combinations of temperature and oxygen, the
239 fluxes were always larger from the loamy sand compared with the clayey silt (Table A3). In general, the mean
240 fluxes from the loamy sand increased with the amount of digestate from 8.3 to 57.6 (aerobic atmosphere at 2°C),
241 from 34.0 to 168.7 (aerobic at 15 °C) and from 11.2 to 87.9 mg CO₂-C m⁻² h⁻¹ (anaerobic at 15°C), but showed no
242 obvious pattern with WFPS. Although the mean fluxes from the clayey silt were also always smallest in the
243 unamended treatments, there was no clear trend of fluxes with the amount of amendment. There was a slight trend
244 of decreasing fluxes with increasing WFPS for the clayey silt. However, the predictive power of WFPS on CO₂-C
245 fluxes was minor since it was eliminated during stepwise regression fitting. By contrast, soil type, amount of
246 digestate and temperature had significant ($p < 0.001$) effects (Table 5).

247 4 Discussion

248 4.1 Increased BD application rate did not increase N₂O and N₂ losses probably due to inhibitory effect 249 of high NH₄⁺ concentrations

250 The overall N₂O fluxes corresponded well with those from other studies with similar incubation conditions and
251 application rates of BD in terms of N ha⁻¹ (Severin et al., 2015; Senbayram et al., 2012; Köster et al., 2015).
252 However, the latter studies assumed a distribution of BD into soil by a cultivator, which implies a smaller
253 concentration of BD compared to its occurrence in injection slits. Although we observed differences in N₂O
254 emissions between soils, soil type was not confirmed as a significant effect. Nevertheless, WFPS and temperature,
255 which are well known controllers of N₂O generation (Maag and Vinther, 1999), showed significant influences.
256 Both are physical (by gas diffusion) and biological (by Q_{10} and consequently O₂ consumption by respiration)
257 proxies for O₂ availability, respectively (Maag and Vinther, 1999; Ball, 2013). Accordingly, the CO₂ flux (as
258 respiration product of O₂) generally increased with temperature and was also identified as significant by regression
259 selection.



260 The mean N_2 fluxes of up to 0.5 (loamy sand) and 3.8 mg N m⁻² h⁻¹ (clayey silt) at 15° C (Fig. 5, Table A1) were
261 considerably smaller than the mean fluxes of up to 13.0 mg m⁻² h⁻¹ observed by Köster et al. (2015) during the first
262 five days of their incubation. Although the amount of BD in terms of applied N (250 kg ha⁻¹) was comparable,
263 Köster et al. (2015) used a higher WFPS of 90%, which may have increased the generation of N_2 . In contrast to
264 N_2O emission rates, the observed N_2 fluxes depended not only on WFPS, but also on soil type (Table 5), most
265 likely due to the direct influence of soil structure on diffusivity and, thus, the supply with O_2 (Balaine et al. 2016;
266 Butterbach-Bahl et al. 2013). N_2O flux showed also a significant effect during regression selection for N_2 . N_2O is
267 the direct precursor of N_2 in many N_2 -producing processes and, hence, the flux of the latter depends on the
268 availability of the former. However, temperature showed no significant effect.

269 The $N_2/(N_2+N_2O)$ ratios were significantly determined only by soil type and WFPS: while no clear trend was
270 observable for the loamy sand, there was a pronounced effect in the clayey silt (Fig 4). We attribute the lack of a
271 trend in the loamy sand to generally adverse conditions for the formation of N_2O and N_2 . Contrary, the influence
272 of WFPS apparently mirrored favourable conditions in the clayey silt (Table 5). Simultaneously, with increasing
273 WFPS, the reduction of N_2O accelerates as an alternative electron acceptor under reduced O_2 supply (Benckiser et
274 al., 2015). Nevertheless, a sufficient soil moisture is required to supply NH_4^+ , NO_3^- and DOC for microbial activity
275 (Blagodatsky and Smith, 2012) and may be the reason why no or rather small fluxes of the investigated gaseous
276 N species were generally found in our treatments with 35% WFPS.

277 In our study, one treatment (clayey silt, 55% WFPS, 160 kg N ha⁻¹) showed exceptionally large mean N_2O fluxes
278 of up to 7.1 mg N m⁻² h⁻¹ (Fig. 3, Table A1). This could be evidence that injection of such moderate amounts may
279 favour much larger losses of N_2O compared to an even distribution of BD in soils due to larger substrate
280 concentration in injection slits. However, with higher amendments (i.e. 320 kg N ha⁻¹), we observed surprisingly
281 partially significant ($p < 0.05$, Tuckey's HSD) reductions of N_2O and a decreasing tendency of N_2 emissions (Table
282 A1, Table A2). In line with this, the amount of BD showed a significant effect during the regression selection on
283 N_2O , but not on N_2 fluxes (Table 5). A coherent reason for the rather smaller emissions of high amended (320 kg
284 N ha⁻¹) treatments might be the inhibitory effect of NH_3 on nitrification, where Anthonisen et al. (1976) found an
285 inhibition by concentrations from 0.1 to 150 mg NH_3 L⁻¹. Since the application rate in the treatments with 320 kg
286 N ha⁻¹ amounted to approximately 500 mg NH_4^+ (kg soil)⁻¹ (Fig. 3) and, thus, relative high concentrations of NH_3
287 in the rather alkaline milieu of BD (Möller and Müller, 2012), we consider this inhibitory effect as an argument
288 for the missing increase of N_2O and N_2 . Additionally, the amount of NH_4^+ fixed as NH_3 by soil organic matter
289 increases with pH and, moreover, this fixed NH_3 is not readily extractable by the KCl method we have applied



290 (Kissel et al., 2008). This is consistent with the observation of generally higher N_2O and N_2 fluxes from the clayey
291 silt since clay increases the sorption capacity of soils for NH_4^+ and may, thus, reduce the inhibitory effect on
292 nitrification (Kissel et al., 2008).

293 An increasing application of BD tended also to decrease the $\text{N}_2/(\text{N}_2+\text{N}_2\text{O})$ ratio, but this effect was also not
294 significant ($p > 0.05$, Tuckey's HSD). In general, NO_2^- and NO_3^- from oxidation of applied NH_4^+ are preferably
295 reduced compared to N_2O during nitrifier-denitrification and denitrification, respectively (Benckiser et al., 2015).
296 Consequently, the share of N_2O builds up with the supply of BD.

297 The inhibitory effect is in line with the strong influence of NO_3^- content of the soils after incubation (Table 5),
298 suggesting a dominant role of denitrifying processes in the generation of N_2O (Butterbach-Bahl et al., 2013).
299 Indeed, coupled nitrification-denitrification and bacterial denitrification have been found to dominate the
300 production of N_2O directly after application of BD (Köster et al., 2011; Senbayram et al., 2009). Notably, in
301 contrast to the clayey silt, no or negligible concentrations of NO_3^- were found in all treatments with loamy sand.
302 Although we have not determined NO_2^- , we speculate that it was a substantial source for reduction by nitrifier
303 denitrification, especially during the anaerobic headspace conditions at the end of the incubation. Actually, high
304 NH_4^+ loads in conjunction with alkaline conditions are typical for BD (Möller and Müller, 2012) favour NO_2^-
305 accumulation and may be the reason for the relatively small NO_3^- recovery in both soils (van Cleemput and
306 Samater, 1995).

307 **4.2 Differentiated effects on N_2O and N_2 fluxes controlled by diffusivity**

308 Apparently, the tested factors affected the N_2O and N_2 fluxes from both soils in a different way. A specific soil
309 characteristic that exhibits such a fundamental control on biogeochemical processes such as denitrifying processes
310 is the diffusivity for O_2 (Ball, 2013; Letey et al., 1980; Parkin and Tiedje, 1984), which is a main soil characteristic
311 responsible for the appearance of anaerobic microsites. In general, diffusivity integrates the soil porosity, i.e., pore
312 continuity and size as well as WFPS, which control both soil N_2O and N_2 emissions (Balaine et al., 2016; Letey et
313 al., 1980; Ball, 2013). Soils with a coarser texture like the loamy sand have a higher proportion of macro-pores
314 and thus a higher saturated conductivity and gas diffusion compared with fine textured soils like the clayey silt we
315 used (Groffman and Tiedje, 1991). This let us expect conditions that are more favourable for N_2O and N_2
316 generation in the latter due to inferior diffusion characteristics and, thus, a smaller O_2 supply. Actually, although
317 we incubated the soils at comparable levels of WFPS and BD amendments, the apparent lower diffusivity led to
318 larger N_2O and N_2 production in the treatments with the clayey silt in relation to the loamy sand.



319 The role of the distinct diffusivities of both soils is corroborated by our observations of the gas fluxes in anaerobic
320 headspace. With switching the He-O₂ atmosphere in the headspace to pure He, the denitrification potential can be
321 tested because anaerobicity eliminates respiration processes that use O₂ as electron acceptor (Parkin and Tiedje,
322 1984). We acknowledge e.g. DNRA and anammox as possible additional sources of N₂O and N₂, respectively,
323 under these conditions, but we were not able to quantify their contribution. The anaerobic headspace induced a
324 considerable increase of N₂O fluxes in the loamy sand, but not in the clayey silt. Concurrently, the N₂ fluxes
325 increased in both soils, but pronounced, i.e. more than 60-fold, in the sandy loam. These observed changes resulting
326 from oxygen deprivation imply that, during the previous aerobic conditions, the diffusivity of the sandy loam was
327 too high to allow for a reasonable establishment of anaerobic microsites, while the clayey silt ensured at least a
328 moderate gas diffusion to maintain hypo-aerobic conditions. However, the large production rates indicate that also
329 the loamy sand harboured the necessary microbial community able to generate N₂ as soon as the atmospheric
330 conditions become favourable. In general, only N₂O fluxes from treatments with negligible fluxes during the
331 previous aerobic period increased under anaerobic conditions. This included all treatments with loamy sand and
332 the highly amended clayey silt with 35% WFPS (Fig. 3, Table A1). At the same time, there was a reduction of
333 N₂O fluxes in all other clayey silt treatments. However, when we take a closer look at the simultaneous changes
334 of N₂ fluxes after atmosphere change, virtually all of these treatments showed increased rates. Hence, there was
335 an enhanced reduction of N₂O to N₂, which is reflected in the increased N₂/(N₂ + N₂O) ratio (Fig. 5) and points to
336 intensified reduction of N₂O due to the lack of oxygen (Parkin and Tiedje, 1984). The much larger N₂ fluxes from
337 the loamy sand compared to the clayey silt might have been caused additionally by small NO₃⁻ availability (Fig.
338 2) and a high availability of C (Table 4), which promoted the reduction of N₂O to N₂ (Benckiser et al., 2015).
339 Alternatively, the much smaller increase of N₂ fluxes from the clayey silt could have resulted from depleted
340 mineral N stocks (NO₃⁻ and NH₄⁺) due to the previous gaseous N losses during the course of incubation. However,
341 the cumulated fluxes of both N₂ and N₂O amounted to a maximum absolute loss of 9.4 (1σ = 0.3) mg N per kg soil
342 in the clayey silt with 160 kg N ha⁻¹ and 55% WFPS, which was roughly 3.5% of the calculated NH₄⁺-N applied
343 with BD (Fig. 2). Thus, we found no evidence for any shortage of substrate in the clayey silt during the subsequent
344 anaerobic headspace conditions. On the other hand, the N₂/(N₂+N₂O) ratios increased only slightly (Fig. 5) and, in
345 contrast to the loamy sand, there were still significant N₂O fluxes in the clayey silt (Fig. 3), which point to still
346 reasonable stocks of NO₃⁻ in the latter (Benckiser et al., 2015). In fact, the NO₃⁻ stock was greater in the clayey silt
347 than in loamy sand after incubation (Fig. 2). Thus, we suggest that the gas fluxes were unaffected by the change
348 to anaerobic headspace in the clayey silt due to already low O₂ concentrations as a result of poor diffusivity. In



349 conclusion, distinct gas diffusivities of both soils can be supposed as the main reason for the differing N_2O and N_2
350 fluxes.

351 In interaction with soil diffusivity, also respiration affects the aerobicity of a soil matrix by concurrent consumption
352 and formation of O_2 and CO_2 , respectively. Similarly to N_2O and N_2 generation by denitrifying processes,
353 respiration depends on the microbial availability of carbon as well. Although anaerobic digestion reduces readily
354 degradable organic matter in BD, a ‘labile’ fraction usually remains, but the biodegradability of the respective
355 residual organic carbon is variable, depending on the origin of BD (Askri et al., 2015). However, DOC could be
356 used as an approximate indicator for microbial availability of carbon, though not all DOC might be readily
357 degradable (Cook and Allan, 1992). Generally, the DOC contents after our incubation increased with application
358 rate of BD (Table 4), but the DOC contents were always smaller in the clayey silt both in the not amended and
359 especially in the amended treatments. This might reflect a stronger sorption of C and thus a lower availability for
360 respiration in the clayey silt compared to the loamy sand (Kaiser and Guggenberger, 2000). If we compare the
361 DOC recoveries with the cumulated flux rates of CO_2 over the incubation cycle, we find a good regression fit (R^2
362 = 0.91, $p < 0.001$) for both soils (Fig. 6) indicating a sufficient availability of C from BD for respiration and, thus,
363 implicitly also for denitrification processes (Reddy et al., 1982). Moreover, as increased DOC enhanced respiration
364 (Table A3), it consequently affected O_2 consumption and, thus, also the emergence of anaerobic microsites (Azam
365 et al., 2002), preferably in the highly amended treatments. Although CO_2 fluxes were mostly higher in the
366 treatments with 320 kg compared to 160 kg $N\ ha^{-1}$, this behaviour was not generally reflected in the emissions of
367 N_2O and N_2 which might be a result of the inhibitory effect of high NH_4^+ loads on nitrification (see chapter 4.1).
368 However, the $N_2/(N_2O+N_2)$ ratios implied a tendency of fostered N_2O reduction due to a shortage of alternative
369 electron acceptors like O_2 in the highly amended treatments. Additionally, temperature influenced indirectly the
370 aerobic status of the soils due to increased microbial activity (Q_{10}) and, thus, respiration (Maag and Vinther, 1999).

371 **4.3 No indications for BD induced short-term priming effect**

372 We further checked for a short-term priming effect after amendment with BD as suggested recently by Coban et
373 al. (2015). After balancing cumulated net CO_2 -C-fluxes (difference between amended and unamended treatments)
374 against the calculated DOC-C application with BD for the period of aerobic headspace, we found no evidence for
375 a short-term priming effect. In the loamy sand with 160 kg $N\ ha^{-1}$, between 76% (35% WFPS) and 103% (75%
376 WFPS) of the DOC-C had been respired (data not shown). In the respective treatments with 320 kg $N\ ha^{-1}$, the
377 CO_2 -C losses ranged from 47% (35% WFPS) to 76% (75% WFPS). By contrast, only between 11% (320 kg $N\ ha^{-1}$



378 ¹) and 42% (160 kg N ha⁻¹) has been respired in the clayey silt (both at 55% WFPS). However, if one would
379 consider the period after BD application for a longer time than we, the light loamy sand may be vulnerable for C
380 losses after BD application than the claggy clayey silt.

381 5 Conclusions

382 As hypothesised, N₂O and N₂ emissions as well as the N₂/(N₂O+N₂) ratio increased with WFPS and temperature,
383 most probably due to restricted supply and enhanced consumption, respectively, of O₂. Contrary to our second
384 hypothesis, the gaseous losses of N₂O and N₂ did not increase with the application rate of BD. This indicates an
385 inhibitory effect of high NH₃ and NH₄⁺ concentrations, respectively, on nitrification, which is found typically in
386 biogas digestates. However, the N₂/(N₂O+N₂) ratio tended to decrease with application rate as supposed, probably
387 due to a copious supply with NO₂⁻ and NO₃⁻ from oxidised BD-NH₄⁺. Confirming our third hypothesis, the fine
388 textured clayey silt induced higher gaseous N losses and a higher N₂/(N₂O+N₂) ratio than the coarse loamy sand
389 by the apparent distinct diffusivities of both soils. Overall, there was a larger potential for formation of N₂O in the
390 fine-textured clayey silt compared to the coarse loamy sand after injection of BD. However, the loamy sand showed
391 a large potential for N₂ formation under anaerobic headspace conditions, indicating the occurrence of an
392 appropriate denitrifying community. In summary, the risk of N₂O and N₂ losses even after injection of relatively
393 large amounts of BD seems to be small for dry to wet sandy soils and acceptable when regarding simultaneously
394 reduced NH₃ emissions for dry silty soils. However, further investigations are needed in regard to study different
395 types of soil and BD, the duration of the observed effects and their reliability for field conditions.

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402 References

403 Anthonisen, A. C., Loehr, R. C., Prakasam, T. B. S., and Srinath, E. G.: Inhibition of Nitrification by Ammonia
404 and Nitrous Acid, J Water Pollut Con F, 48, 835–852, 1976.



- 405 Askri, A., Laville, P., Trémier, A., and Houot, S.: Influence of Origin and Post-treatment on Greenhouse Gas
 406 Emissions After Anaerobic Digestate Application to Soil, *Waste Biomass Valor*, 1–14, doi:10.1007/s12649-
 407 015-9452-6, 2015.
- 408 Azam, F., Müller, C., Weiske, A., Benckiser, G., and Ottow, J.: Nitrification and denitrification as sources of
 409 atmospheric nitrous oxide – role of oxidizable carbon and applied nitrogen, *Biol Fert Soils*, 35, 54–61,
 410 doi:10.1007/s00374-001-0441-5, 2002.
- 411 Balaine, N., Clough, T. J., Beare, M. H., Thomas, S. M., and Meenken, E. D.: Soil Gas Diffusivity Controls N₂O
 412 and N₂ Emissions and their Ratio, *Soil Sci Soc Am J*, 80, 529–540, doi:10.2136/sssaj2015.09.0350, 2016.
- 413 Ball, B. C.: Soil structure and greenhouse gas emissions: a synthesis of 20 years of experimentation, *Eur J Soil
 414 Sci*, 64, 357–373, doi:10.1111/ejss.12013, 2013.
- 415 Benckiser, G., Schartel, T., and Weiske, A.: Control of NO₃⁻ and N₂O emissions in agroecosystems: A review,
 416 *Agron Sustain Dev*, 1–16, doi:10.1007/s13593-015-0296-z, 2015.
- 417 Blagodatsky, S. and Smith, P.: Soil physics meets soil biology: Towards better mechanistic prediction of
 418 greenhouse gas emissions from soil, *Soil Biol Biochem*, 47, 78–92, doi:10.1016/j.soilbio.2011.12.015, 2012.
- 419 Butterbach-Bahl, K., Baggs, E. M., Dannenmann, M., Kiese, R., and Zechmeister-Boltenstern, S.: Nitrous oxide
 420 emissions from soils: how well do we understand the processes and their controls?, *Philos T Roy Soc B*,
 421 368, doi:10.1098/rstb.2013.0122, 2013.
- 422 Butterbach-Bahl, K., Willibald, G., and Papen, H.: Soil core method for direct simultaneous determination of N₂
 423 and N₂O emissions from forest soils, *Plant Soil*, 240, 105–116, doi:10.1023/A:1015870518723, 2002.
- 424 Cameron, K. C., Di, H. J., and Moir, J. L.: Nitrogen losses from the soil/plant system: a review, *Ann Appl Biol*,
 425 162, 145–173, doi:10.1111/aab.12014, 2013.
- 426 Coban, H., Miltner, A., Elling, F. J., Hinrichs, K. U., and Kästner, M.: The contribution of biogas residues to soil
 427 organic matter formation and CO₂ emissions in an arable soil, *Soil Biol Biochem*, 86, 108–115,
 428 doi:10.1016/j.soilbio.2015.03.023, 2015.
- 429 Cook, B. D. and Allan, D. L.: Dissolved organic carbon in old field soils: Total amounts as a measure of
 430 available resources for soil mineralization, *Soil Biol Biochem*, 24, 585–594, doi:10.1016/0038-
 431 0717(92)90084-B, 1992.
- 432 Davidson, E. A. and Kanter, D.: Inventories and scenarios of nitrous oxide emissions, *Environ Res Lett*, 9,
 433 105012, 2014.
- 434 Davidson, E. A., Suddick, E. C., Rice, C. W., and Prokopy, L. S.: More Food, Low Pollution (Mo Fo Lo Po): A
 435 Grand Challenge for the 21st Century, *J Environ Qual*, 44, doi:10.2134/jeq2015.02.0078, 2015.
- 436 Dell, C. J., Meisinger, J. J., and Beegle, D. B.: Subsurface Application of Manures Slurries for Conservation
 437 Tillage and Pasture Soils and Their Impact on the Nitrogen Balance, *J Environ Qual*, 40,
 438 doi:10.2134/jeq2010.0069, 2011.
- 439 Eickenscheidt, T., Heinichen, J., Augustin, J., Freibauer, A., and Drösler, M.: Nitrogen mineralization and
 440 gaseous nitrogen losses from waterlogged and drained organic soils in a black alder (*Alnus glutinosa* (L.)
 441 Gaertn.) forest, *Biogeosciences*, 11, 2961–2976, doi:10.5194/bg-11-2961-2014, 2014.
- 442 Ferm, M.: Atmospheric ammonia and ammonium transport in Europe and critical loads: a review, *Nutr Cycl
 443 Agroecosys*, 51, 5–17, doi:10.1023/A:1009780030477, 1998.



- 444 Firestone, M. K. and Davidson, E. A.: Microbiological basis of NO and N₂O production and consumption in soil,
445 in: Exchange of Trace Gases Between Terrestrial Ecosystems and the Atmosphere, Andreae, M. a. S. D.
446 (Ed.), Wiley, Chichester, 7–21, 1989.
- 447 Friedl, J., Scheer, C., Rowlings, D. W., McIntosh, H. V., Strazzabosco, A., Warner, D. I., and Grace, P. R.:
448 Denitrification losses from an intensively managed sub-tropical pasture – Impact of soil moisture on the
449 partitioning of N₂ and N₂O emissions, *Soil Biol Biochem*, 92, 58–66, doi:10.1016/j.soilbio.2015.09.016,
450 2016.
- 451 Groffman, P. M. and Tiedje, J. M.: Relationships between denitrification, CO₂ production and air-filled porosity
452 in soils of different texture and drainage, *Soil Biol Biochem*, 23, 299–302, doi:10.1016/0038-
453 0717(91)90067-T, 1991.
- 454 Jurasinski, G., Köbsch, F., Günther, A., and Beetz, S.: flux: Flux rate calculation from dynamic closed chamber
455 measurements, R package version 0.3-0, <http://CRAN.R-project.org/package=flux> (last access: 8 February
456 2017), 2014.
- 457 Kaiser, E. A., Kohrs, K., Kucke, M., Schnug, E., Heinemeyer, O., and Munch, J. C.: Nitrous oxide release from
458 arable soil: Importance of N-fertilization, crops and temporal variation, *Soil Biol Biochem*, 30, 1553–1563,
459 doi:10.1016/S0038-0717(98)00036-4, 1998.
- 460 Kaiser, K. and Guggenberger, G.: The role of DOM sorption to mineral surfaces in the preservation of organic
461 matter in soils, *Org Geochem*, 31, 711–725, doi:10.1016/S0146-6380(00)00046-2, 2000.
- 462 Kissel, D. E., Cabrera, M. L., and Paramasivam, S.: Ammonium, Ammonia, and Urea Reactions in Soils, in:
463 Nitrogen in Agricultural Systems, *Agronomy Monographs*, American Society of Agronomy, Crop Science
464 Society of America, Soil Science Society of America, Madison, WI, 101–155, 2008.
- 465 Köster, J. R., Cárdenas, L., Senbayram, M., Bol, R., Well, R., Butler, M., Mühlhling, K. H., and Dittert, K.: Rapid
466 shift from denitrification to nitrification in soil after biogas residue application as indicated by nitrous oxide
467 isotopomers, *Soil Biol Biochem*, 43, 1671–1677, doi:10.1016/j.soilbio.2011.04.004, 2011.
- 468 Köster, J. R., Cárdenas, L. M., Bol, R., Lewicka-Szczebak, D., Senbayram, M., Well, R., Giesemann, A., and
469 Dittert, K.: Anaerobic digestates lower N₂O emissions compared to cattle slurry by affecting rate and
470 product stoichiometry of denitrification – An N₂O isotopomer case study, *Soil Biol Biochem*, 84, 65–74,
471 doi:10.1016/j.soilbio.2015.01.021, 2015.
- 472 Kuznetsova, A., Brockhoff, P. B., and Christensen, R. H. B.: lmerTest: Tests in Linear Mixed Effects Models, R
473 package version 2.0-29, <http://CRAN.R-project.org/package=lmerTest> (last access: 8 February 2017), 2015.
- 474 Letey, J., Jury, W. A., Hadas, A., and Valoras, N.: Gas Diffusion as a Factor in Laboratory Incubation Studies on
475 Denitrification1, *J Environ Qual*, 9, doi:10.2134/jeq1980.00472425000900020012x, 1980.
- 476 Maag, M. and Vinther, F. P.: Effect of temperature and water on gaseous emissions from soils treated with
477 animal slurry, *Soil Sci Soc Am J*, 63, 858–865, 1999.
- 478 Möller, K.: Effects of anaerobic digestion on soil carbon and nitrogen turnover, N emissions, and soil biological
479 activity. A review, *Agron Sustain Dev*, 1–21, doi:10.1007/s13593-015-0284-3, 2015.
- 480 Möller, K. and Müller, T.: Effects of anaerobic digestion on digestate nutrient availability and crop growth: A
481 review, *Eng Life Sci*, 12, 242–257, doi:10.1002/elsc.201100085, 2012.



- 482 Mosier, A., Kroeze, C., Nevison, C., Oenema, O., Seitzinger, S., and van Cleemput, O.: Closing the global N₂O
483 budget: nitrous oxide emissions through the agricultural nitrogen cycle - OECD/IPCC/IEA phase II
484 development of IPCC guidelines for national greenhouse gas inventory methodology, *Nutr Cycl*
485 *Agroecosys*, 52, 225–248, doi:10.1023/A:1009740530221, 1998.
- 486 Myhre, G., Shindell, D., Bréon, F. M., Collins, W., Fuglestedt, J., Huang, J., Koch, D., Lamarque, J. F., Lee,
487 D., and Mendoza, B.: Anthropogenic and natural radiative forcing, in: *Climate Change 2013: The Physical*
488 *Science Basis.: Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental*
489 *Panel on Climate Change*, Stocker, T. F., Qin, D., Plattner, G. K., Tignor, M., Allen, S. K., Boschung, J.,
490 Nauels, A., Xia, Y., Bex, V., and Midgley, P. M. (Eds.), Cambridge University Press, Cambridge, United
491 Kingdom, New York, NY, USA, 659–740, 2013.
- 492 Nkoa, R.: Agricultural benefits and environmental risks of soil fertilization with anaerobic digestates: a review,
493 *Agron Sustain Dev*, 1–20, doi:10.1007/s13593-013-0196-z, 2013.
- 494 Parkin, T. B. and Tiedje, J. M.: Application of a soil core method to investigate the effect of oxygen
495 concentration on denitrification, *Soil Biol Biochem*, 16, 331–334, doi:10.1016/0038-0717(84)90027-0,
496 1984.
- 497 Quakernack, R., Pacholski, A., Techow, A., Herrmann, A., Taube, F., and Kage, H.: Ammonia volatilization and
498 yield response of energy crops after fertilization with biogas residues in a coastal marsh of Northern
499 Germany, *Agr Ecosyst Environ*, 160, 66–74, doi:10.1016/j.agee.2011.05.030, 2012.
- 500 R Core Team: R: A Language and Environment for Statistical Computing, Vienna, Austria: [https://www.R-](https://www.R-project.org)
501 [project.org](https://www.R-project.org), (last access: 8 February 2017), 2017.
- 502 Reddy, K. R., Rao, P. S. C., and Jessup, R. E.: The Effect of Carbon Mineralization on Denitrification Kinetics
503 in Mineral and Organic Soils I, *Soil Sci Soc Am J*, 46, 62–68,
504 doi:10.2136/sssaj1982.03615995004600010011x, 1982.
- 505 Scholefield, D., Hawkins, J. M. B., and Jackson, S. M.: Development of a helium atmosphere soil incubation
506 technique for direct measurement of nitrous oxide and dinitrogen fluxes during denitrification, *Soil Biol*
507 *Biochem*, 29, 1345–1352, doi:10.1016/S0038-0717(97)00021-7, 1997.
- 508 Senbayram, M., Chen, R., Budai, A., Bakken, L., and Dittert, K.: N₂O emission and the N₂O/(N₂O + N₂) product
509 ratio of denitrification as controlled by available carbon substrates and nitrate concentrations, *Agr Ecosyst*
510 *Environ*, 147, 4–12, doi:10.1016/j.agee.2011.06.022, 2012.
- 511 Senbayram, M., Chen, R., Mühling, K. H., and Dittert, K.: Contribution of nitrification and denitrification to
512 nitrous oxide emissions from soils after application of biogas waste and other fertilizers, *Rapid Comm Mass*
513 *Sp*, 23, 2489–2498, doi:10.1002/rcm.4067, 2009.
- 514 Severin, M., Fuss, R., Well, R., Garlipp, F., and van den Weghe, H.: Soil, slurry and application effects on
515 greenhouse gas emissions, *Plant, Soil Environm*, 61, 344–351, 2015.
- 516 Uchida, Y., Clough, T. J., Kelliher, F. M., and Sherlock, R. R.: Effects of aggregate size, soil compaction, and
517 bovine urine on N₂O emissions from a pasture soil, *Soil Biol Biochem*, 40, 924–931,
518 doi:10.1016/j.soilbio.2007.11.007, 2008.
- 519 van Cleemput, O. and Samater, A. H.: Nitrite in soils: accumulation and role in the formation of gaseous N
520 compounds, *Fert Res*, 45, 81–89, doi:10.1007/BF00749884, 1995.



- 521 Velthof, G. L. and Mosquera, J.: The impact of slurry application technique on nitrous oxide emission from
522 agricultural soils, *Agr Ecosyst Environ*, 140, 298–308, doi:10.1016/j.agee.2010.12.017, 2011.
- 523 Webb, J., Pain, B., Bittman, S., and Morgan, J.: The impacts of manure application methods on emissions of
524 ammonia, nitrous oxide and on crop response—A review, *Agr Ecosyst Environ*, 137, 39–46,
525 doi:10.1016/j.agee.2010.01.001, 2010.
- 526 Wulf, S., Maeting, M., and Clemens, J.: Application technique and slurry co-fermentation effects on ammonia,
527 nitrous oxide, and methane emissions after spreading: II. Greenhouse gas emissions, *J Environ Qual*, 31,
528 1795–1801, 2002.
- 529 Zhang, X., Davidson, E. A., Mauzerall, D. L., Searchinger, T. D., Dumas, P., and Shen, Y.: Managing nitrogen
530 for sustainable development, *Nature*, 528, 51–59, 2015.
- 531



532 **Table 1: The examined factors soil texture, water-filled pore space (WFPS), and amount (i.e., concentration) of nitrogen**
533 **(N) applied with biogas digestate (BD) with their respective levels applied in the present study, resulting in 18**
534 **treatments with three replicates each. The temperature was manipulated consecutively during the incubation.**

Factor [n]	Levels		
Soil texture [2]	loamy sand	clayey silt	
WFPS (%) [3]	35	55	75
BD-N (kg ha ⁻¹) [3]	0	160	320
Temperature (°C) [2]	2	15	

535



536 **Table 2: Characteristics of both soils. Texture and mean values with standard deviations (in brackets) for carbon (C, $n = 9$), nitrogen (N, $n = 9$), pH ($n = 3$), bulk density (BD, $n = 3$)**
 537 **and mineral N (NO_3^- and NH_4^+ , $n = 3$) of both soils in 0–10 cm depth after field sampling.**

Texture	C (mg g^{-1}) ^a	N (mg g^{-1}) ^a	pH ^b	Bulk density (g cm^{-3}) ^c	NO_3^- (mg kg^{-1}) ^d	NH_4^+ (mg kg^{-1}) ^d
Loamy sand	6.99 (0.29)	0.67 (0.05)	7.2 (0.1)	1.4 (0.0)	1.0 (0.2)	0.6 (0.3)
Clayey silt	10.77 (0.28)	1.19 (0.06)	7.2 (0.0)	1.5 (0.0)	1.8 (0.2)	0.3 (0.2)

538 ^a measured with analyser “Truspec CNS”, Leco Instruments GmbH, Germany, performed according to ISO 10694 (“elemental analysis”) for C and according to ISO 13878

539 (“elemental analysis”) for N

540 ^b measured in H_2O with TiiraMaster85, Radiometer Analytical SAS, France, performed according to VDLUFA Methodenbuch, Vol. 1, chap. 5.1.1

541 ^c measured on 250 cm^3 soil cores

542 ^d measured with analyser “CFA-SAN”, Skalar Analytical B.V., the Netherlands, performed according to ISO 14256



543 **Table 3: Chronological order of the incubated factor combinations. Two different factor combinations with their**
 544 **respective repetitions ($n = 3$) were placed together for each weekly incubation course (cf. Fig. 1). The factors were**
 545 **combined by (1) soil (loamy sand: LS, clayey silt: CS), (2) amount (kg) of applied N from digestate per ha and (3) WFPS**
 546 **(%).**

Week	Factor combination 1	Factor combination 2
1	LS - 0 N - 35%	LS - 0 N - 55%
2	LS - 0 N - 75%	LS - 160 N - 35%
3	LS - 160 N - 55%	LS - 160 N - 75%
4	LS - 320 N - 35%	LS - 320 N - 55%
5	LS - 320 N - 75%	CS - 0 N - 35%
6	CS - 0 N - 55%	CS - 0 N - 75%
7	CS - 160 N - 35%	CS - 160 N - 55%
8	CS - 160 N - 75%	CS - 320 N - 35%
9	CS - 320 N - 55%	CS - 320 N - 75%

547



548 **Table 4: Mean recovered DOC values from soils after incubation with standard deviations in brackets for the respective**
 549 **treatments differing in amount of applied biogas digestate (BD) and water-filled pore space (WFPS).**

	kg digestate-N ha ⁻¹	WFPS (%)	mg DOC (kg soil) ⁻¹	
			Loamy sand	Clayey silt
0		35	41.4 (2.7)	18.9 (1.1)
		55	38.6 (3.1)	19.8 (1.4)
		75	43.7 (1.4)	19.0 (1.8)
160		35	197.4 (20.7)	n.a.
		55	190.5 (19.3)	68.3 (12.7)
		75	362.2 (40.0)	63.2 (9.6)
320		35	316.8 (25.3)	358.1 (26.3)
		55	312.5 (14.3)	94.8 (13.6)
		75	500.1 (33.4)	105.9 (14.8)

550 n.a.: data not available



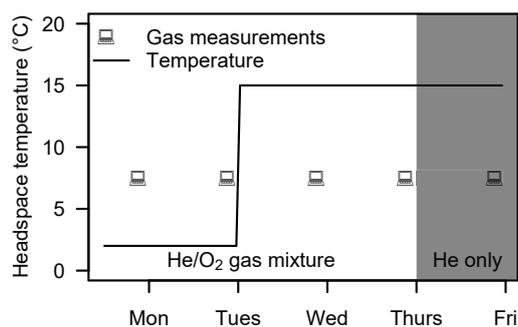
551 Table 5: Significant fixed effects with *P*-values (*t*-test) of the linear mixed models for estimated fluxes of N₂, N₂O, N₂/(N₂+N₂O) product ratio and CO₂ in aerobic He-O₂ atmosphere
 552 with soil type, water-filled pore space (WFPS), amount of digestate, temperature, NO₃⁻ content of soil after incubation as well as fluxes of N₂O and CO₂ as possible independent
 553 variables. The respective vessels in the incubation system were set as random effect, which was always significant.

554

Fixed effects							
Response	Soil type	WFPS	Digestate amount	Temperature	NO ₃ ⁻ soil post	N ₂ O flux	CO ₂ flux
N ₂	<0.001	<0.001	†	†	†	<0.001	†
N ₂ O	†	<0.001	<0.001	<0.001	<0.001	*	<0.001
N ₂ /(N ₂ +N ₂ O)	<0.001	<0.001	†	†	†	*	†
CO ₂	<0.001	†	<0.001	<0.001	†	†	*

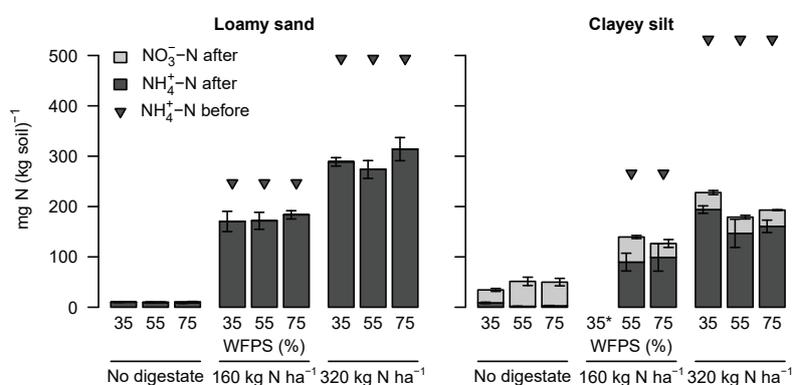
555 † Variable eliminated during stepwise regression selection

556 * Variable was not included into original regression



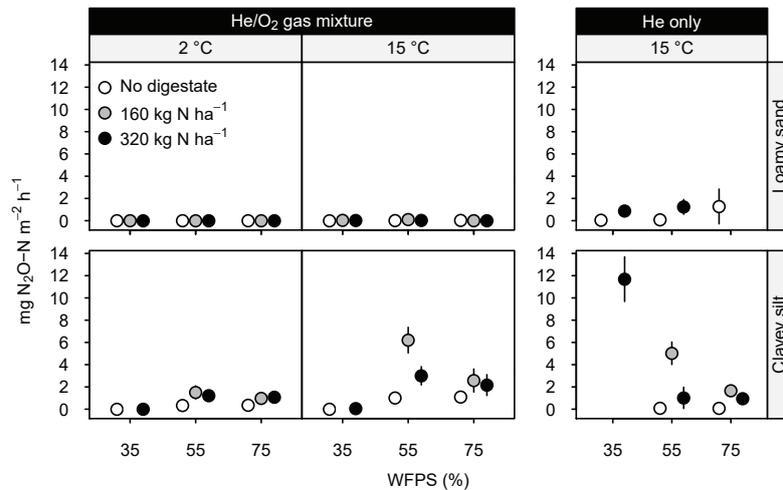
557 **Figure 1: Course of incubation and gas measurements with respect to atmosphere and temperature of the headspace**
558 **after two days of pre-incubation at 2 °C in He/O₂ gas mixture. Gas concentrations of the headspace were determined**
559 **on five consecutive days, i.e. Monday to Friday in the morning. After the first two measurement days, the headspace**
560 **temperature was increased from 2 to 15 °C. Additionally, after the fourth measurement day, the aerobic Helium/oxygen**
561 **gas mixture in the headspace was replaced by a pure Helium atmosphere.**

562

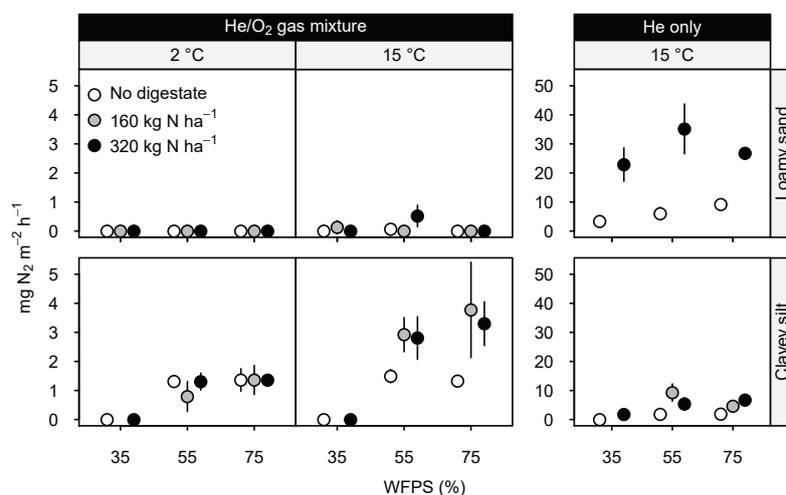


563 **Fig. 2: Ammonium and nitrate contents from loamy sand and clayey silt after incubation with different water-filled**
 564 **pore spaces (WFPS, %) and amounts of digestate (kg N ha⁻¹). Error bars denote standard deviations. In general, the**
 565 **ammonium content increased with digestate application with lower amounts detected in the clayey silt. Nitrate was**
 566 **found almost exclusively in the latter soil. For comparison, calculated amounts of ammonium applied with biogas**
 567 **digestate are shown by triangles. One treatment (*) was omitted from all analyses due to technical reasons.**

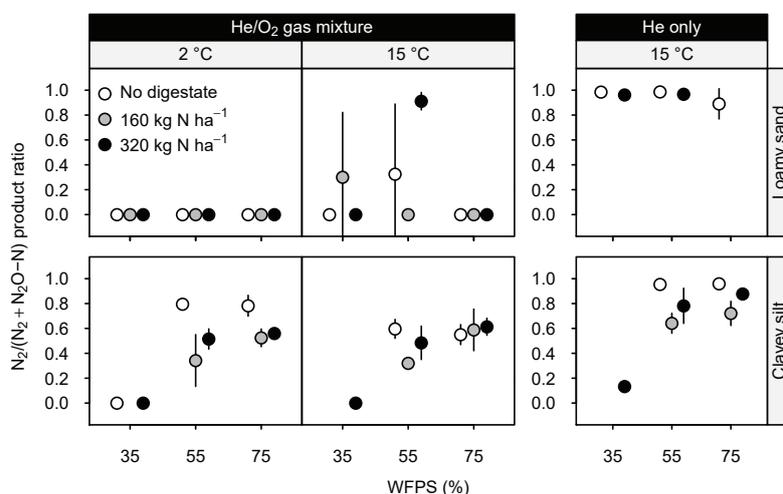
568



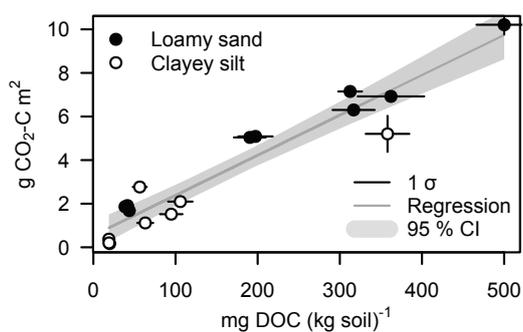
569 **Fig. 3:** Mean N_2O fluxes ($\text{mg N m}^{-2} \text{h}^{-1}$) from a loamy sand and a clayey silt incubated under different water-filled pore
 570 spaces (WFPS, %) with different amounts of digestate (kg N ha^{-1}). Shown are values measured on the second and fourth
 571 day of incubation (i.e. after two days with 2 and 15 °C, respectively) under aerobic He- O_2 atmosphere as well as day
 572 five (15 °C) under anaerobic He atmosphere. Error bars show standard deviations; if bars are not visible, they are
 573 smaller than the symbols of the means. Under aerobic atmosphere, N_2O fluxes from loamy sand were negligible, while
 574 fluxes from clayey silt showed an increase with temperature, especially with higher WFPS and intermediate amounts
 575 of digestate. Under anaerobic atmosphere, mean fluxes from loamy sand increased slightly, but significantly (Tukey's
 576 HSD, $p < 0.05$). The fluxes from clayey silt showed no significant differences (Tukey's HSD, $p < 0.05$) compared to the
 577 day before, with the exception of 35% WFPS, where mean flux increased strongly in the treatment with 320 kg digestate-
 578 N ha^{-1} .



579 Fig. 4: Mean N₂ fluxes (mg m⁻² h⁻¹) from a loamy sand and a clayey silt incubated under different water-filled pore
 580 spaces (WFPS, %) with different amounts of digestate (kg N ha⁻¹). Shown are values measured on the second and fourth
 581 day of incubation (i.e. after two days with 2 and 15 °C, respectively) under aerobic He-O₂ atmosphere as well as day
 582 five (15 °C) under anaerobic He atmosphere. Error bars show standard deviations; if bars are not visible, they are
 583 smaller than the symbols of the means. Under aerobic atmosphere, N₂ fluxes from loamy sand were zero or rather
 584 negligible, while fluxes from clayey silt show a distinct increase with WFPS and higher fluxes at 15 °C. Under anaerobic
 585 atmosphere, mean fluxes from loamy sand increased by orders of magnitude, while the fluxes from clayey silt increased
 586 as well, but more gently compared to the sand.



587 Fig. 5: Mean $N_2/(N_2 + N_2O-N)$ product ratio from a loamy sand and a clayey silt incubated under different water-filled
 588 pore spaces (WFPS, %) with different amounts of digestate ($kg\ N\ ha^{-1}$). Shown are values measured on the second and
 589 fourth day of incubation (i.e. after two days with 2 and 15 °C, respectively) under aerobic He-O₂ atmosphere as well as
 590 day five (15 °C) under anaerobic He atmosphere. Error bars show standard deviations; if bars are not visible, they are
 591 smaller than the symbols of the means. For the loamy sand, there was a clear distinction of the ratios between aerobic
 592 and anaerobic atmospheres: while the ratios tended to 0 in the former, they tended to 1 in the latter, irrespectively of
 593 temperature or amount of digestate. For the clayey silt, ratios increased with WFPS and were highest from the
 594 unamended treatments under both the aerobic and the anaerobic atmospheres.



595 **Fig. 6: Regression between DOC recoveries (mg per 100 g soil) after the incubation and the respective cumulated CO₂**
596 **emissions (g C m⁻²) during the period of aerobic headspace with their standard deviations and confidence interval**
597 **(95%). If error bars are not visible, they are smaller than the symbols of the means. Both soils showed increasing**
598 **emissions with increasing soil DOC contents as well a good regression fit ($R^2 = 0.91$, $p < 0.001$).**



599 **Table A1: Mean N₂O-N fluxes with standard deviations in mg m⁻² h⁻¹ from the loamy sand and the clayey silt, treated**
 600 **with different water-filled pore spaces (WFPS, %), amounts of digestate (kg N ha⁻¹) as well as different temperature**
 601 **regimes (°C) under aerobic (He-O₂) and anaerobic (He) atmosphere. Column 'Day' denotes the consecutive measuring**
 602 **days of the respective incubation cycle. Different letters after fluxes indicate significant differences (Tukey's HSD, *p* <**
 603 **0.05) within each soil and measuring day. Zeros as last digits were omitted.**

Day	Atmosphere	Temperature (°C)	WFPS (%)	kg N ha ⁻¹	mg N ₂ O-N m ⁻² h ⁻¹	
					Loamy sand	Clayey silt
1	He-O ₂	2	35	0	0 ± 0	0 ± 0 c
1	He-O ₂	2	35	160	0 ± 0	NA
1	He-O ₂	2	35	320	0 ± 0	0 ± 0 c
1	He-O ₂	2	55	0	0 ± 0	0.3 ± 0.1 c
1	He-O ₂	2	55	160	0 ± 0	1.7 ± 0.4 a
1	He-O ₂	2	55	320	0 ± 0	1.1 ± 0.1 b
1	He-O ₂	2	75	0	0 ± 0	0.4 ± 0.1 c
1	He-O ₂	2	75	160	0 ± 0	1 ± 0.1 b
1	He-O ₂	2	75	320	0 ± 0	1 ± 0.2 b
2	He-O ₂	2	35	0	0 ± 0	0 ± 0 d
2	He-O ₂	2	35	160	0 ± 0	NA
2	He-O ₂	2	35	320	0 ± 0	0 ± 0 cd
2	He-O ₂	2	55	0	0 ± 0	0.3 ± 0.1 bc
2	He-O ₂	2	55	160	0 ± 0	1.5 ± 0.6 a
2	He-O ₂	2	55	320	0 ± 0	1.2 ± 0.2 a
2	He-O ₂	2	75	0	0 ± 0	0.4 ± 0.1 bc
2	He-O ₂	2	75	160	0 ± 0	1 ± 0.1 ab
2	He-O ₂	2	75	320	0 ± 0	1.1 ± 0.2 a
3	He-O ₂	15	35	0	0 ± 0 cd	0 ± 0 c
3	He-O ₂	15	35	160	0 ± 0 abc	NA
3	He-O ₂	15	35	320	0 ± 0 ab	0 ± 0 c
3	He-O ₂	15	55	0	0 ± 0 bcd	0.8 ± 0.2 c
3	He-O ₂	15	55	160	0 ± 0 bcd	7.1 ± 0.9 a
3	He-O ₂	15	55	320	0 ± 0 a	3.5 ± 0.7 b
3	He-O ₂	15	75	0	0 ± 0 ab	0.8 ± 0.2 c
3	He-O ₂	15	75	160	0 ± 0 d	3.2 ± 0.7 b
3	He-O ₂	15	75	320	0 ± 0 cd	3 ± 0.9 b
4	He-O ₂	15	35	0	0 ± 0 b	0 ± 0 c
4	He-O ₂	15	35	160	0 ± 0 ab	NA
4	He-O ₂	15	35	320	0 ± 0 ab	0.1 ± 0.1 c
4	He-O ₂	15	55	0	0 ± 0 b	1 ± 0.2 bc
4	He-O ₂	15	55	160	0.1 ± 0.1 a	6.2 ± 1.1 a
4	He-O ₂	15	55	320	0 ± 0 ab	3 ± 0.8 b



4	He-O ₂	15	75	0	0 ± 0 ab	1.1 ± 0.3 bc
4	He-O ₂	15	75	160	0 ± 0 b	2.6 ± 1 b
4	He-O ₂	15	75	320	0 ± 0 b	2.2 ± 0.9 b
5	He	15	35	0	0.1 ± 0	NA
5	He	15	35	160	NA	NA
5	He	15	35	320	0.9 ± 0.1	11.7 ± 2 a
5	He	15	55	0	0.1 ± 0	0.1 ± 0 c
5	He	15	55	160	NA	5 ± 1 b
5	He	15	55	320	1.2 ± 0.7	1.4 ± 0.8 c
5	He	15	75	0	1.3 ± 1.6	0.1 ± 0 c
5	He	15	75	160	NA	1.7 ± 0.3 c
5	He	15	75	320	NA	1 ± 0.3 c

604



605 **Table A2: Mean N₂ fluxes with standard deviations in mg m⁻² h⁻¹ from the loamy sand and the clayey silt, treated with**
 606 **different water-filled pore spaces (WFPS, %), amounts of digestate (kg N ha⁻¹) as well as different temperature regimes**
 607 **(°C) under aerobic (He-O₂) and anaerobic (He) atmosphere. Column 'Day' denotes the consecutive measuring days of**
 608 **the respective incubation cycle. Different letters after fluxes indicate significant differences (Tukey's HSD, *p* < 0.05)**
 609 **within each soil and measuring day. Zeros as last digits were omitted.**

	Atmosphere	Temperature (°C)	WFPS (%)	kg N ha ⁻¹	mg N ₂ m ⁻² h ⁻¹	
					Loamy sand	Clayey silt
1	He-O ₂	2	35	0	0 ± 0	0 ± 0 bc
1	He-O ₂	2	35	160	0 ± 0	NA
1	He-O ₂	2	35	320	0 ± 0	0.1 ± 0.1 bc
1	He-O ₂	2	55	0	0 ± 0	1.5 ± 0.3 a
1	He-O ₂	2	55	160	0 ± 0	1.5 ± 0.3 a
1	He-O ₂	2	55	320	0 ± 0	1.5 ± 0 a
1	He-O ₂	2	75	0	0 ± 0	1.2 ± 1.1 a
1	He-O ₂	2	75	160	0 ± 0	1.5 ± 0.2 a
1	He-O ₂	2	75	320	0 ± 0	1.7 ± 0.4 a
2	He-O ₂	2	35	0	0 ± 0	0 ± 0 c
2	He-O ₂	2	35	160	0 ± 0	NA
2	He-O ₂	2	35	320	0 ± 0	0 ± 0 c
2	He-O ₂	2	55	0	0 ± 0	1.3 ± 0.1 a
2	He-O ₂	2	55	160	0 ± 0	0.8 ± 0.5 b
2	He-O ₂	2	55	320	0 ± 0	1.3 ± 0.3 a
2	He-O ₂	2	75	0	0 ± 0	1.4 ± 0.4 a
2	He-O ₂	2	75	160	0 ± 0	1.4 ± 0.5 a
2	He-O ₂	2	75	320	0 ± 0	1.4 ± 0.1 a
3	He-O ₂	15	35	0	0 ± 0 b	0 ± 0 e
3	He-O ₂	15	35	160	0 ± 0 b	NA
3	He-O ₂	15	35	320	0.1 ± 0.1 ab	0 ± 0 e
3	He-O ₂	15	55	0	0 ± 0 b	1.8 ± 0.3 cd
3	He-O ₂	15	55	160	0 ± 0 b	2.3 ± 0.4 bc
3	He-O ₂	15	55	320	0 ± 0 b	2.5 ± 0.2 ab
3	He-O ₂	15	75	0	0.2 ± 0.3 a	1.5 ± 0.2 d
3	He-O ₂	15	75	160	0 ± 0 b	3 ± 0.9 a
3	He-O ₂	15	75	320	0 ± 0 b	2.6 ± 0.8 ab
4	He-O ₂	15	35	0	0 ± 0 b	0 ± 0 c
4	He-O ₂	15	35	160	0.1 ± 0.2 b	NA
4	He-O ₂	15	35	320	0 ± 0 b	0 ± 0 c
4	He-O ₂	15	55	0	0.1 ± 0.1 b	1.5 ± 0.2 b
4	He-O ₂	15	55	160	0 ± 0 b	2.9 ± 0.6 a
4	He-O ₂	15	55	320	0.5 ± 0.4 a	2.8 ± 0.7 a



4	He-O ₂	15	75	0	0 ± 0 b	1.3 ± 0.2 bc
4	He-O ₂	15	75	160	0 ± 0 b	3.8 ± 1.6 a
4	He-O ₂	15	75	320	0 ± 0 b	3.3 ± 0.8 a
5	He	15	35	0	3.3 ± 0.4 d	0 ± 0 c
5	He	15	35	160	NA	NA
5	He	15	35	320	22.9 ± 5.7 b	1.8 ± 0.1 c
5	He	15	55	0	6 ± 2.2 cd	1.8 ± 0.2
5	He	15	55	160	NA	9.5 ± 2.7 a
5	He	15	55	320	35.1 ± 8.6 a	5.1 ± 1.8 bc
5	He	15	75	0	9.2 ± 0.4 c	1.9 ± 0.1 c
5	He	15	75	160	NA	4.8 ± 1.6 bc
5	He	15	75	320	26.8 ± 1.1 b	6.7 ± 0.8 b

610



611 **Table A3: Mean CO₂-C fluxes with standard deviations in mg m⁻² h⁻¹ from the loamy sand and the clayey silt, treated**
 612 **with different water-filled pore spaces (WFPS, %), amounts of digestate (kg N ha⁻¹) as well as different temperature**
 613 **regimes (°C) under aerobic (He-O₂) and anaerobic (He) atmosphere. Column 'Day' denotes the consecutive measuring**
 614 **days of the respective incubation cycle. Different letters after fluxes indicate significant differences (Tukey's HSD, *p* <**
 615 **0.05) within each soil and measuring day. Zeros as last digits were omitted.**

Day	Atmosphere	Temperature (°C)	WFPS (%)	kg N ha ⁻¹	mg CO ₂ -C m ⁻² h ⁻¹	
					Loamy sand	Clayey silt
1	He-O ₂	2	35	0	6.8 ± 2.4 cd	0 ± 0 c
1	He-O ₂	2	35	160	22 ± 3.5 bcd	NA
1	He-O ₂	2	35	320	23.3 ± 9.3 bc	22.8 ± 2.8 ab
1	He-O ₂	2	55	0	6 ± 0.7 d	4.6 ± 7.9 bc
1	He-O ₂	2	55	160	34.4 ± 3.1 b	34.5 ± 11.6 a
1	He-O ₂	2	55	320	28 ± 3.2 b	15.9 ± 3.4 abc
1	He-O ₂	2	75	0	9.4 ± 1.4 cd	0 ± 0 c
1	He-O ₂	2	75	160	37.5 ± 6 b	15.5 ± 12.1 abc
1	He-O ₂	2	75	320	68.3 ± 12.1 a	24.5 ± 2.7 a
2	He-O ₂	2	35	0	9.8 ± 3.5 c	1.3 ± 1.4 b
2	He-O ₂	2	35	160	23 ± 3.9 bc	NA
2	He-O ₂	2	35	320	30.9 ± 2.2 b	22.2 ± 2.4 a
2	He-O ₂	2	55	0	8.7 ± 1.5 c	0.6 ± 1 b
2	He-O ₂	2	55	160	33.4 ± 0.9 b	27.6 ± 12.3 a
2	He-O ₂	2	55	320	35.9 ± 2.7 b	14.4 ± 1.9 ab
2	He-O ₂	2	75	0	8.3 ± 1.5 c	0 ± 0 b
2	He-O ₂	2	75	160	31.9 ± 3 b	13 ± 9.3 ab
2	He-O ₂	2	75	320	57.6 ± 14.8 a	18.3 ± 4 a
3	He-O ₂	15	35	0	42.5 ± 4.5 c	6.7 ± 0.7 b
3	He-O ₂	15	35	160	114.3 ± 12.2 b	NA
3	He-O ₂	15	35	320	149.5 ± 9.4 b	130.9 ± 105 a
3	He-O ₂	15	55	0	41.3 ± 3.5 c	3.2 ± 0.4 b
3	He-O ₂	15	55	160	108.7 ± 10.1 b	57.8 ± 12.2 bc
3	He-O ₂	15	55	320	162.1 ± 9.6 b	26.8 ± 0.7 bc
3	He-O ₂	15	75	0	44.1 ± 9.8 c	3.2 ± 0.7 b
3	He-O ₂	15	75	160	150.4 ± 19 b	26.4 ± 11.8 bc
3	He-O ₂	15	75	320	249.7 ± 53.5 a	35.3 ± 6 bc
4	He-O ₂	15	35	0	48.7 ± 6 c	15.1 ± 4.9 cd
4	He-O ₂	15	35	160	114.3 ± 6.4 b	NA
4	He-O ₂	15	35	320	156.9 ± 15.4 a	65.7 ± 2.2 a
4	He-O ₂	15	55	0	48 ± 3.4 c	4.2 ± 0.2 d
4	He-O ₂	15	55	160	109 ± 14.4 b	51.2 ± 15.1 ab
4	He-O ₂	15	55	320	177.7 ± 7.5 a	26.6 ± 2.3 cd



4	He-O ₂	15	75	0	34 ± 7.8 c	6.7 ± 4 d
4	He-O ₂	15	75	160	168.7 ± 0.4 a	22.1 ± 14.8 cd
4	He-O ₂	15	75	320	166.3 ± 23.1 a	34.1 ± 5.7 bc
5	He	15	35	0	11.2 ± 0.6 d	NA
5	He	15	35	160	54.8 ± 9.3 c	NA
5	He	15	35	320	149.3 ± 3.9 a	45.8 ± 2.1 a
5	He	15	55	0	13.6 ± 1.9 d	3.4 ± 0.6 c
5	He	15	55	160	55.2 ± 4.4 bc	32 ± 11.4 ab
5	He	15	55	320	164.5 ± 3.5 a	15.2 ± 10.7 bc
5	He	15	75	0	20.9 ± 2.3 d	3.6 ± 0.1 c
5	He	15	75	160	75 ± 7.3 b	20.6 ± 8.5 bc
5	He	15	75	320	NA	26.1 ± 2.6 ab

616