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# **Bone char effects on soil: sequential fractionations and XANES spectroscopy**

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22 **Abstract.** Acceptability of novel bone char fertilizers depends on their P release but  
23 reactions at bone char surfaces and impacts to soil P speciation are insufficiently known. By  
24 sequential fractionation and synchrotron-based X-ray absorption near edge structure  
25 (XANES) spectroscopy we investigated whether and how the chemical composition of bone  
26 char particles has been altered in soil and, consequently, has affected the P speciation of  
27 amended soils. Therefore, two different kinds of bone char particles (BC, produced by  
28 pyrolysis of degreased animal bone chips at 800 °C and BC<sup>plus</sup>, a BC enriched with reduced  
29 sulfur compounds) were manually separated from the soil at the end of two different  
30 experiments: “incubation-leaching” and “ryegrass cultivation”. Sequential P-fractionation of  
31 amended soils showed P-enrichment in all fractions as compared to the control. The most P  
32 increase between all treatments significantly occurred in the NaOH-P and resin-P fractions in  
33 response to BC<sup>plus</sup> application in both, incubation-leaching and ryegrass cultivation  
34 experiments. This increase of the readily available P fraction in BC<sup>plus</sup> treated soils was  
35 confirmed by linear combination fitting (LCF) analysis on P K-edge XANES spectra of BCs  
36 particles and amended soils. The proportion of Ca-hydroxyapatite decreased, whereas the  
37 proportion of CaHPO<sub>4</sub> increased in BC<sup>plus</sup> particles after amended soils had been incubated  
38 and leached and cropped by ryegrass. Based on P XANES speciation as determined by LCF  
39 analysis, the proportion of inorganic Ca(H<sub>2</sub>PO<sub>4</sub>)<sub>2</sub> increased in amended soils after BC<sup>plus</sup>  
40 application. These results indicated that soil amendment with BC<sup>plus</sup> particles leads to elevated  
41 P concentration and maintains more soluble P species than BC particles even after 230-days  
42 of ryegrass cultivation.

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## 46 **1 Introduction**

47 At the global scale readily-available sources of phosphorus (P), a crucial macro-nutrient  
48 element for agricultural production, are being faced with scarcity and overpricing (Scholz et  
49 al., 2013; Van Vuuren et al., 2010). Environmental pollutions frequently arise from their  
50 impurities (Cd, U) (Hartley et al., 2013; Kratz et al., 2016), and from over-fertilization  
51 (Rubaek et al., 2013). Further problems are the quick formation of stable and inaccessible  
52 compounds that limit plant P uptake (Shen et al., 2011) and the low agronomic efficiency of  
53 not more than 15% of fertilizer P in the first year of application (Schnug et al., 2003). Many  
54 recent studies have targeted sustainable agriculture through improving P-availability from  
55 applied fertilizers (Delgado et al., 2002; Schröder et al., 2011), increasing P-uptake efficiency  
56 from organic and inorganic P-pools in the soil (Kaur and Reddy, 2014) and developing new  
57 technologies for P-recycling from human and animal waste (Siebers and Leinweber, 2013;  
58 Herzel et al., 2016). Particular attention has been paid to the oxidation process, e.g. by  
59 thiobacilli of elemental sulfur to sulfuric acid, in order to enhance the solubility of non water-  
60 soluble P from rock phosphates (Powers, 1923; Lee et al., 1987; Fan et al., 2002) or meat and  
61 bone ashes (Schnug et al., 2003).

62 As an economically and environmentally attractive example, pyrolyzed animal bone chips  
63 branded as “bone char” (BC), a slow-release apatite-based P-fertilizer, have been surface  
64 modified by sulfur (S) compounds to enhance its solubility in neutral to alkaline soils.  
65 Incubation-leaching and pot experiments confirmed that surface-modification was an effective  
66 approach in P-release promotion from BC fertilizer (Morshedizad et al., 2016; Zimmer, D.  
67 and Panten, K., personal communication). Such an "in situ" digestion of an apatitic phosphate  
68 with elemental S was first described by Fan et al. (2002; 2012).

69 Despite these attempts to raise dissolution and use-efficiency of BC in supplying P for crop  
70 requirements, a considerable fraction of applied BC-P to the soil remains insoluble in the

71 short term and is not taken up by plants over the entire cropping period. A detailed P  
72 speciation can clarify the fate of insoluble P from BC which has not been done before.

73 Chemical speciation is described as analytical identification of chemical species of defined  
74 elements and measuring their quantities in the system (Templeton et al., 2000). The precise  
75 characterization of various P species in the soil as a dynamic response to non-equilibrium  
76 conditions imposed by human activities such as fertilization can support a better  
77 understanding of reactivity, stability and particularly plant-accessibility of different P-forms  
78 and provide a basis for best management practices. Several techniques such as sequential  
79 fractionation (Dieter et al., 2010; Condrón and Newman, 2011), nuclear magnetic resonance  
80 (NMR) spectroscopy (Liu et al., 2009; Vestergren et al., 2012; Ahlgren et al., 2013), Raman  
81 spectroscopy (Lanfranco, 2003; Vogel et al., 2013) and chromatography coupled to mass  
82 spectroscopy (De Brabandere et al., 2008; Paraskova et al., 2015) have been developed for P  
83 speciation analysis in soil and sediments. Each one of these techniques can offer specific  
84 advantages and disadvantages depending on phase and complexity of sample matrixes (Kruse  
85 et al., 2015). Complementarily, X-ray absorption near edge structure (XANES) spectroscopy  
86 is well-suited for identification of various P species through fingerprinting of molecular  
87 structures in solid and heterogeneous mediums based on fine feature and position of  
88 absorbing edges (Kelly et al., 2008; Kizewski et al., 2011). The advantages of XANES  
89 spectroscopy for soil samples make it a promising technique for direct and *in situ* P-speciation  
90 with no pretreatment and minimum sample manipulation (Toor et al., 2006; Kelly et al.,  
91 2008).

92 According to the best of our knowledge, no studies have characterized P-speciation changes  
93 in BC particles over non-equilibrium conditions in the soil system, and only few  
94 investigations have been reported on the P-release from BC and alteration in P-species of the  
95 soil. Siebers et al. (2013) investigated *K*-edge XANES spectroscopy on BC-incubated soil

96 samples and provided evidence that the increase in extractable Ca- and Mg-phosphate  
97 fractions were related to the contribution of hydroxyapatite (HAP) increase after BC  
98 application. Accordingly, the objective of this study was to provide practical information on  
99 the fate and alteration of P-species in BC and novel surface modified BC (BC<sup>plus</sup>) particles  
100 and their treated soils under incubation-leaching and ryegrass cultivation practices, using  
101 sequential P-fractionation and P-XANES spectroscopy.

102

## 103 **2 Materials and methods**

### 104 **2.1 Incubation-leaching experiment**

105 Two particle size fractions (1-2 and 2-4 mm) of bone chars (BC, produced by pyrolysis of  
106 degreased animal bone chips at 800°C and BC<sup>plus</sup> as a surface modified BC obtained by  
107 blending with reduced S-containing compounds composed of 60% elemental S, 30% calcium  
108 sulfate dehydrate and 10% methansulfonate (Zimmer et al. unpublished results of S X-ray  
109 absorption near-edge fine structure spectroscopy) in a commercial biogas desulfurization  
110 process; patent application DE 212012000046U1; [www.google.com/patents](http://www.google.com/patents/DE212012000046U1?cl=en&hl=de)  
111 [/DE212012000046U1?cl=en&hl=de](http://www.google.com/patents/DE212012000046U1?cl=en&hl=de)) were incubated with a silt loam soil. The soil was  
112 classified as *Dystric Cambisol* (FAO) with pH of 4.7 (measured in 0.01 mol L<sup>-1</sup> CaCl<sub>2</sub>  
113 solution) and total (digestion with HNO<sub>3</sub> and analyzed using ICP-OES; USEPA, 1997) and  
114 available (extracted by 1 mol L<sup>-1</sup> NH<sub>4</sub>NO<sub>3</sub> and analyzed using ICP-OES; He and Singh, 1993)  
115 P-contents of 1.6 g P kg<sup>-1</sup> and 14 mg P kg<sup>-1</sup>, respectively. The BC and BC<sup>plus</sup> contained total P  
116 of 149 and 123 g kg<sup>-1</sup>, total calcium (Ca) 185 and 265 g kg<sup>-1</sup>, total S 6 and 199 g kg<sup>-1</sup> and had  
117 average of pH<sub>CaCl2</sub> values 7.8 and 4.9, respectively.

118 The BCs were added to 30 g of air dry soil (< 2 mm) at the levels of 0 mg P kg<sup>-1</sup> soil (control)  
119 and 500 mg P kg<sup>-1</sup> soil in five replicates. The soil and BCs mixture was homogenized and  
120 packed into glass columns with 10 cm length and inner diameter of 2 cm. A P-free filter (MN

121 616 G; Macherey-Nagel GmbH & Co., KG Düren, Germany) was placed at the bottom of  
122 each column to avoid any particle losses. The amended soils were incubated for 70 days at 20  
123 °C in the dark and constant soil moisture between 60-70% of soil water holding capacity.  
124 During the incubation period, the soil columns were leached with three pore volumes of  
125 deionized water added by a droplet irrigation simulator system. The leaching process was  
126 repeated in five steps, each one after 1, 5, 13, 34 and 70 days. The P-concentrations in  
127 collected leachates were measured using inductively coupled plasma-optical emission  
128 spectrometry (ICP-OES). Outcomes of the leaching experiment were described in  
129 Morshedizad and Leinweber (2017). After the incubation-leaching experiment, the treated soil  
130 samples were carefully removed from the glass columns, air dried and BCs particles were  
131 manually separated from the soils very gently. The BCs particles were delicately washed with  
132 deionized water to remove adhered soil particles, allowed to dry completely at ambient  
133 conditions and finely ground for further analyses.

134

## 135 **2.2 Pot experiment with annual ryegrass**

136 The same BC and BC<sup>plus</sup> as described for the incubation-leaching experiment were used in  
137 original sizes (mostly between 1 to 5 mm) for P fertilization of annual ryegrass in a pot  
138 experiment. The experiment was set-up using an acidic sandy silt soil with available P-content  
139 of 24.2 mg P kg<sup>-1</sup> and pH of 5.2. The pot experiment was set up by adding BC and BC<sup>plus</sup> at  
140 the levels of 0 mg P kg<sup>-1</sup> (control) and 280 mg P kg<sup>-1</sup> into the 6 kg of the soil dry matter in  
141 each pot and in four replicates arranged in a complete randomized block. After four weeks of  
142 incubation at field capacity water content and ambient temperature conditions, 30 seeds of  
143 annual ryegrass per each pot were sown on 13<sup>th</sup> of May 2016. The experiment was conducted  
144 in a glasshouse under ambient air and temperature conditions and the soil moisture was  
145 maintained at field capacity during the whole experiment. All other essential nutrients were

146 sequentially added at sufficient levels before seeding and after each six cuts of ryegrass  
147 between 23<sup>rd</sup> of June and 3<sup>rd</sup> of November 2016. Finally, after the last harvest (7<sup>th</sup>), plant parts  
148 (shoots and roots) were dried at 60 °C and BCs particles were manually separated from the  
149 soils as they could be detected visually by their size and dark color very gently, using  
150 tweezers. Then these particles were washed delicately with deionized water to remove  
151 attached soil particles, allowed to dry completely at ambient conditions and finely ground to  
152 fine powders for further analyses.

153

### 154 **2.3 Sequential phosphorus fractionation**

155 Soil samples were sequentially extracted based on chemical solubility in order according to a  
156 modified Hedley et al. (1982) procedure. After BC particles detachment, duplicate 0.5 g fine-  
157 ground and air-dried soil samples were weighed into 50-mL centrifuge tubes. In summary,  
158 chemical P fractionation includes the following steps:

159 1) The mobile and readily available P fraction was extracted with resin strips (saturated in 0.5  
160 M NaHCO<sub>3</sub>) after 18 hours end-over-end shaking in 30 mL deionized water. The resin strips  
161 were separated from solids/solution and washed using 50 mL of 1 M HCl to remove absorbed  
162 P. The soil suspension was centrifuged at 2500 × g for 20 minutes and the supernatant was  
163 decanted.

164 2) Next, the labile inorganic and organic fractions weakly absorbed to mineral surfaces and  
165 some microbial P were extracted by 30 mL of 0.5 M NaHCO<sub>3</sub>, 18 hours end-over-end shaking  
166 and centrifugation at 2500 × g for 20 minutes. The supernatant was filtered (Whatman no. 42  
167 filter) and collected for measurements.

168 3) The inorganic P adsorbed and bound to Al- and Fe-oxide minerals and organic P from  
169 humic substances were extracted using 30 mL of 0.1 M NaOH solution and repeating the  
170 second step procedure as described above.

171 4) The relatively insoluble fraction of P bound to Ca and Mg minerals and apatite was  
172 extracted by 30 mL of 1 M HCl in the same way as for the previous steps.

173 Total P concentrations ( $P_t$ ) and inorganic P ( $P_i$ ) in all extracts were measured by ICP-OES and  
174 colorimetrically (molybdenum blue method; Murphy and Riley, 1962), respectively. The  
175 organic P ( $P_o$ ) concentrations were calculated by  $P_t - P_i$ .

176

## 177 **2.4 Phosphorus K-edge XANES analysis**

178 The XANES data collection for characterizing P-species in all soil samples and BCs particles  
179 was acquired at the Synchrotron Light Research Institute (SLRI) in Nakhon Ratchasima,  
180 Thailand, on the beamline 8 (BL8) of the electron storage ring with a covering photon energy  
181 from 1.25 to 10 KeV, electron energy operated at 1.2 GeV and beam current of 80-150 mA  
182 (Klysubun et al., 2012). The P K-edge XANES spectra were collected from dried and very  
183 finely ground treated soils and particulate BCs samples which had been diluted to P  
184 concentrations  $< 10 \text{ mg P kg}^{-1}$  with  $\text{SiO}_2$  powder (to eliminate self absorption effects; Prietzel  
185 et al., 2013), again ground in agate stone mini-mortar and spread uniformly as a thin layer on  
186 P-free kapton tape (Lanmar Inc., Northbrook, IL, USA). Data collection was operated in  
187 standard conditions comprising the energy calibration by standard pure elemental P and  
188 allocating the reference energy ( $E_0$ ) at 2145.5 eV using the maximum peak of spectrum. All  
189 spectra were recorded at photon energies between 2045.5 and 2495.5 eV in step sizes of 5 eV  
190 (2045.5 to 2105.5 eV and 2245.5 to 2495.5 eV), 1 eV (2105.5 to 2135.5 eV and 2195.5 to  
191 2245.5 eV) and 0.25 eV (2135.5 to 2195.5 eV) with a 13-channel germanium detector in  
192 fluorescence mode. At least three scans were collected and averaged for each sample.

193 The P-XANES spectra were normalized and after merging replicates, a linear combination  
194 fitting (LCF) was performed using the ATHENA software package (Ravel and Newville,  
195 2005). All XANES spectral data were baseline corrected in the pre-edge region between



196 2115–2145 eV and normalized in the post-edge region of 2190–2215 eV. The same ranges  
197 were used for the reference P *K*-edge XANES spectra to achieve consistency in the following  
198 fitting analysis (Prietz et al. 2016). To achieve the best compatible set of references with  
199 each specified sample spectrum, LCF analysis was performed in the energy range between -  
200 20 eV and +30 eV relative to the  $E_0$  using the combinatorics function of ATHENA software  
201 to attain all possible binary, ternary and at most quaternary combinations between all nineteen  
202 P reference spectra. The following set of reference P *K*-edge XANES spectra, all recorded in  
203 SLRI under the same adjustments by Werner and Prietz (2015) and Prietz et al. (2016),  
204 were used for fitting and calculations; Ca-, Al- and Fe-phytate, non-crystalline and crystalline  
205  $\text{AlPO}_4$ , non-crystalline and crystalline  $\text{FePO}_4 \cdot 2\text{H}_2\text{O}$ , Ca-hydroxyapatite ( $\text{Ca}_5(\text{OH})(\text{PO}_4)_3$ ),  
206 inositol hexakisphosphate (IHP), ferrihydrite-IHP, montmorillonite-Al-IHP, soil organic  
207 matter (SOM)-Al-IHP, ferrihydrite-orthophosphate, montmorillonite-Al-orthophosphate,  
208 SOM-Al-orthophosphate, bohemite-IHP, bohemite-orthophosphate,  $\text{CaHPO}_4$ ,  $\text{Ca}(\text{H}_2\text{PO}_4)_2$   
209 and  $\text{MgHPO}_4$ . To select the best possible combination fit between the sample spectrum and  
210 the P-reference spectra, the lowest reduced chi value ( $\chi^2$ ) and *R*-factor were chosen.

211

## 212 **3 Results**

### 213 **3.1 Effect of BCs on sequentially extracted P after incubation-leaching**

214 After 70 days of incubation-leaching, the sequential P-fractionation of amended soils showed  
215 variations in the amount and distribution of various P-fractions between different treatments  
216 (Table 1). For all treatments, NaOH extracted the majority of fractionated P (62.4 to 66.5% of  
217 total fractionated P), followed by the labile P fraction ( $\text{NaHCO}_3$ , 19.2 to 20.0%), HCl-P (9.5  
218 to 13.9%) and the readily available P (resin strips, 4.1 to 4.8%). The BCs addition increased  
219 the total soil P pools although the difference was significant only for the  $\text{BC}^{\text{plus}}_{1-2\text{ mm}}$  and  $\text{BC}_{1-}$   
220  $_{2\text{ mm}}$  treatments. The largest increase in total fractionated  $P_t$  ( $\text{resin-}P_t + \text{NaHCO}_3\text{-}P_t + \text{NaOH-}P_t$

221 + HCl-P<sub>i</sub>) occurred in BC<sup>plus</sup><sub>1-2 mm</sub> (133.8 mg P kg<sup>-1</sup> soil) followed by BC<sub>1-2 mm</sub> (118.6 mg P  
222 kg<sup>-1</sup> soil), BC<sub>2-4 mm</sub> (67.1 mg P kg<sup>-1</sup> soil) and BC<sup>plus</sup><sub>2-4 mm</sub> (35.7 mg P kg<sup>-1</sup> soil), compared to  
223 the control soil.

224

225

### INSERT Table 1 HERE

226

227 The proportion of P-enrichment in each fraction varied between different treatments in the  
228 order NaOH-P > HCl-P > NaHCO<sub>3</sub>-P > resin-P for BC<sup>plus</sup><sub>1-2 mm</sub> and BC<sup>plus</sup><sub>2-4 mm</sub>. For the BC<sub>1-  
229 2 mm</sub> and BC<sub>2-4 mm</sub> treatment the order was HCl-P > NaOH-P > NaHCO<sub>3</sub>-P > resin-P. In all  
230 treatments, the P<sub>i</sub> proportions in each of the P fractions were greater than the P<sub>o</sub> proportions.  
231 Compared to the control soil, the most P<sub>i</sub> increase was observed in NaOH-P<sub>i</sub> and resin-P<sub>i</sub> in  
232 response to BC<sup>plus</sup><sub>1-2 mm</sub> application (Table 1). Moreover, after 70 days of incubation-leaching,  
233 soil pH increased in BC treatments whereas BC<sup>plus</sup> amendments had an acidifying effect. Soil  
234 pH levels of BC<sub>1-2 mm</sub> and BC<sub>2-4 mm</sub> increased by 0.07 and 0.05 units and decreased for BC<sup>plus</sup><sub>1-  
235 2 mm</sub> and BC<sup>plus</sup><sub>2-4 mm</sub> treatments by 0.21 and 0.15 units compared to unamended control soil  
236 (pH = 5.06).

237

### 238 **3.2 Effect of BCs on sequentially extracted P after ryegrass cropping**

239 Sequentially extracted P fractions in soil varied between different treatments after 230 days of  
240 ryegrass cropping (Table 1). In all treatments (control, BC and BC<sup>plus</sup>), NaOH-P was the  
241 largest P pool mainly associated with Al- and Fe-oxide minerals and humic substances (65.0  
242 to 67.5% of total fractionated P) followed by the NaHCO<sub>3</sub>-P (18.2 to 19.0%), HCl-P (11.5 to  
243 12.6%) and resin-P (2.2 to 3.4%) fractions. Enrichments of P fractions in BC<sup>plus</sup> treatments  
244 were more pronounced than in treated soils with BC particles. In this treatment the  
245 concentrations of readily available and labile inorganic P fractions were insignificantly

246 smaller than in the control. Additionally, a significant increase in P concentration was  
247 obtained only in resin-P<sub>i</sub> and NaOH-P<sub>i</sub> fractions of the BC<sup>plus</sup> treated soil (Table 1). The  
248 maximum increase of total fractionated P was obtained in BC<sup>plus</sup> treatment (37.6 mg P kg<sup>-1</sup>  
249 soil). In comparison to incubation-leaching results, a similar sequence was observed for the  
250 order of increasing magnitude of P fractions in response to BC and BC<sup>plus</sup> amendments  
251 (NaOH-P > NaHCO<sub>3</sub>-P > HCl-P > resin-P). However, for BC treatment, the total P extracted  
252 by resin strips was lowered in comparison with the control. In the control and BC treatments,  
253 P<sub>o</sub> was the predominant form in NaOH-P and NaHCO<sub>3</sub>-P fractions while for BC<sup>plus</sup> it was  
254 only in the NaOH-P fraction. Each P fraction was highest under BC<sup>plus</sup> application, except for  
255 NaHCO<sub>3</sub>-P<sub>o</sub> of BC treatment.

256 Separately, the effect of BC and BC<sup>plus</sup> application on ryegrass yield parameters was  
257 examined in the 230-days pot experiment. The results indicated that P uptake, ryegrass yield  
258 and apparent nutrient recovery efficiency (ANR) of BC<sup>plus</sup> treatments exceeded that of BC and  
259 control treatments and increased to values comparable with triple super phosphate (TSP)  
260 fertilizer (Zimmer, D. and Panten, K., personal communication). Addition of BC and BC<sup>plus</sup>  
261 did not significantly change the bulk soil pH, although local acidification around BC<sup>plus</sup>  
262 particles (pH 4.9; Morshedizad and Leinweber, 2017) probably can lower soil pH in small  
263 scale areas compared to BC treatments (pH about 8).

264

### 265 **3.3 XANES analysis of BCs particles**

266 All spectra from BCs were characterized by an intense white-line peak, post-edge position  
267 and without a distinct pre-edge which corresponded to calcium phosphate compounds  
268 including Ca-hydroxyapatite, dicalcium phosphate (CaHPO<sub>4</sub>) and Ca-phytate (Fig. 1). The P  
269 K-edge XANES results indicated no obvious alterations in spectral features of BCs particles  
270 after the incubation-leaching experiment. After 70-days of incubation-leaching, the BC

271 spectra were shifted towards Ca-hydroxyapatite, and this was more pronounced for the 2-4  
272 mm than for the 1-2 mm BC-particles. The opposite trend was the case for BC<sup>plus</sup> particles  
273 where the white-line signal intensity decreased after incubation-leaching period and the post-  
274 edge of spectra tended more to dicalcium phosphate. This effect was stronger for BC<sup>plus</sup>-  
275 particle size reduction from 2-4 mm to 1-2 mm.

276

277

**INSERT Figure 1 HERE**

278

279 To quantify the P speciation of BC and BC<sup>plus</sup> particles, LCF analyses using all possible  
280 combinations were performed on all P *K*-edge XANES spectra (Table 2). The fitting results  
281 indicated that untreated BC and BC<sup>plus</sup> particles before the experiment contained on average  
282 61 and 60% Ca-hydroxyapatite, 22 and 30% CaHPO<sub>4</sub> and 18 and 10% Ca-phytate. After 70-  
283 days of incubation-leaching, the proportion of Ca-hydroxyapatite increased to the average of  
284 80% in BC while it remained unchanged in BC<sup>plus</sup> particles. The CaHPO<sub>4</sub> proportion increased  
285 in BC<sup>plus</sup> particles to the average of 34% whereas the lower content was assigned in the  
286 spectra of BC particles accounting for 10% of total P species. Moreover, the Ca-phytate  
287 proportion decreased slightly in BC and BC<sup>plus</sup> particles from about 18 and 10% to averages  
288 of 11 and 7%, respectively.

289

290

**INSERT Table 2 HERE**

291

292 Spectra of BC and BC<sup>plus</sup> particles, before and after 230-days of ryegrass cultivation, were  
293 characterized by a sharp white-line followed by a shoulder and then a post-edge feature  
294 between 2160 and 2175 eV which was divided into two peaks (Fig. 2). These features were  
295 most similar to P *K*-edge XANES spectra of Ca-hydroxyapatite, CaHPO<sub>4</sub> and Ca-phytate

296 standard compounds. Treated BC particles had a white-line with higher intensity which  
297 appeared more similar to the Ca-hydroxyapatite spectrum. In contrast, BC<sup>plus</sup> particles under  
298 ryegrass cultivation showed a weaker white-line exhibiting the shoulder and post-edge feature  
299 more comparable to the CaHPO<sub>4</sub> spectrum.

300

301

**INSERT Figure 2 HERE**

302

303 Some differences in proportions of P species observed between BCs particles before and after  
304 the cropping period in the ryegrass pot experiment are presented in Table 3. The LCF results  
305 revealed overall contributions of 63 and 70% Ca-hydroxyapatite, 29 and 29% CaHPO<sub>4</sub> and 8  
306 and 1% Ca-phytate in the original BC and BC<sup>plus</sup>, respectively. After the cropping period, the  
307 percentage of Ca-hydroxyapatite was increased in BC particles. In the BC<sup>plus</sup> treatment, the  
308 percentage of CaHPO<sub>4</sub> increased from 29 to 43, while the percentage of Ca-hydroxyapatite  
309 was reduced from 70 to 49%. The Ca-phytate proportion remained unchanged in BC particles  
310 while that of BC<sup>plus</sup> increased from 1 to 8% after the ryegrass cultivation period.

311

312

**INSERT Table 3 HERE**

313

### 314 **3.4 XANES analysis of soil samples**

315 The P *K*-edge XANES spectra of soil samples from the incubation-leaching experiment  
316 showed two dominant features including 1) a strong white-line lacking pre-edge and shoulder  
317 and 2) a tailed post-edge feature (Fig. 3). The most similarity to these features was seen in  
318 XANES spectra of amorphous AlPO<sub>4</sub>, FePO<sub>4</sub> and SOM-Al-IHP compounds. Distinct  
319 differences appeared between the control and treated soil with BC<sup>plus</sup>, not with BC treatments.  
320 This was reflected by slightly lower intensities of both white-line and post-edge features.

321

322

**INSERT Figure 3 HERE**

323

324 The P species of treated soils in the incubation-leaching experiment were determined by LCF  
325 analysis to select at most four reference compounds in combinatorics of all possible fitting  
326 combinations (Table 4). The fitting results indicated that P in the control soil and BC  
327 treatments occurred dominantly as  $\text{AlPO}_4$  amorphous ( $\approx 40\%$ ),  $\text{FePO}_4$  ( $\approx 30\%$ ) and SOM-Al-  
328 IHP ( $\approx 20\%$ ) compounds. In  $\text{BC}^{\text{plus}}$  treated soils, the average proportion of amorphous  $\text{AlPO}_4$   
329 decreased to 26% and instead,  $\text{Ca}(\text{H}_2\text{PO}_4)_2$  was identified with an average of 25% which did  
330 not appear in the control and BC treatments. The LCF results showed that the soil treated with  
331  $\text{BC}^{\text{plus}}$  had no detectable Ca-hydroxyapatite which was found in the control and BC  
332 treatments.

333

334

**INSERT Table 4 HERE**

335

336 The XANES spectra recorded from treated soil samples in the ryegrass pot experiment  
337 showed the presence of an intense white-line in the energy range of 2152 to 2158 eV and a  
338 stretched post-edge feature approximately from 2165 to 2178 eV (Fig. 4). Decreases in white-  
339 line and post-edge intensities of the soil samples appeared as an effect of  $\text{BC}^{\text{plus}}$  application.  
340 Visual inspection of P *K*-edge spectra revealed no indication of specific alteration in spectral  
341 features in response to the BC treatment.

342

343

**INSERT Figure 4 HERE**

344

345 Amorphous  $\text{AlPO}_4$  was identified by LCF analysis as dominant component ( $\approx 35\%$ ) in all  
346 treated soil samples from the ryegrass pot experiment (Table 5). The second major P form in  
347 the control soil was IHP (29%) followed by Ca-phytate (27%), with the latter also as  
348 pronounced as that observed for BC and  $\text{BC}^{\text{plus}}$  treatments. All treated soils varied in  
349 proportions of free or bound IHP forms. The  $\text{Mont-Al-PO}_4$  and  $\text{Ca}(\text{H}_2\text{PO}_4)_2$  compounds were  
350 only assigned in the control and  $\text{BC}^{\text{plus}}$  treatments, respectively.

351

352

**INSERT Table 5 HERE**

353

## 354 **4 Discussion**

### 355 **4.1 P availability as revealed by sequential fractionation**

356 The sequence of P distribution between sequentially extracted P-fractions was in accordance  
357 with findings by many studies (Cross and Schlesinger, 1995; McDowell and Stewart, 2006;  
358 Hashimoto and Watanabe, 2014), reflecting the general status of different P-pools in acidic  
359 soils. The results indicated that the largest P proportion was found in the NaOH fraction  
360 reflecting P fixed to Fe- and Al-oxides as followed by the  $\text{NaHCO}_3$ -P fraction assigned to  
361 weakly P absorbed on crystalline Fe- and Al-oxides or surface of minerals. Guo et al. (2000)  
362 reported that the NaOH-P fraction may support the labile  $\text{NaHCO}_3$ -P fraction as a buffering  
363 P-pool in highly weathered and acidic soils. According to soil pH values (4.7 and 5.2), the  
364 larger proportions of  $\text{NaHCO}_3$ -P even than HCl-P can be explained by the abundance and  
365 surface loadings of Fe- and Al-oxides that support the electrostatic binding of phosphate ions  
366 and a scarcity of Ca and Mg minerals or soluble ions. As expected, the lowest P proportions  
367 were found in the mobile and readily available P fraction extracted by resin strips in  
368 agreement to many comparable studies (Cross and Schlesinger, 1995; Bauchemin et al., 2003;  
369 Sharpley et al., 2004; Siebers et al., 2013). Among the two soils which were used in the two

370 different experiments, the largest proportions of inorganic P were achieved in the soil after  
371 incubation-leaching experiment, while the organic P forms were considerably more abundant  
372 in the soil samples after ryegrass cultivation (Table 1). These differences may be due to the  
373 microbial activities in the rhizosphere of grasses and transformation of  $P_i$  to more stable  $P_o$   
374 fractions during longer plant cultivation period (230 days) than in the non-cropped  
375 incubation-leaching experiment (70 days).

376 In general, all P fraction concentrations were elevated by adding BC and  $BC^{plus}$  particles  
377 which appeared to follow the same pattern in both soils under two different experimental  
378 conditions. However, significant differences were found only between the control and  $BC^{plus}$   
379 treated soils (1-2 mm in the incubation-leaching experiment) for the resin-P and NaOH-P  
380 fractions. Since the BCs particles were separated from the soils before chemical analysis, it  
381 was expected that partly dissolved BCs would have a limited impact on different P-fractions  
382 rather than totally ground and mixed BCs. This is consistent with the study of Siebers et al.  
383 (2013) according to which the BC application ( $<90 \mu\text{m}$  BC thoroughly mixed to soil)  
384 significantly increased the insoluble P proportion ( $\text{H}_2\text{SO}_4\text{-P}$ ). Additionally, our study  
385 confirmed previous findings concerning the effect of particle sizes on the P release from BCs  
386 (Morshedizad and Leinweber, 2017) and consequently the P status of treated soils (Ma and  
387 Matsunaka, 2013). Sequentially extracted P contents increased with decreasing size of BC  
388 particles whereby  $BC^{plus}$  treatments appeared more dependent on particle size than BC  
389 treatments. The results of sequential P fractionation of  $BC^{plus}$  treatments in the incubation-  
390 leaching experiment indicated that the P increase was more pronounced for P fixed to Al- and  
391 Fe-oxides (NaOH-P) than other fractions, whereas for BC treatments the largest increase  
392 occurred in P bound to Ca and Mg minerals (HCl-P). It seems that local pH changes in soil  
393 associated with BC and  $BC^{plus}$  amendments could eventually lead to a different distribution of  
394 released P into differently soluble or insoluble P-pools which are generally controlled by pH



395 (Arai and Sparks, 2007). However, due to lower fertilization level and longer period of  
396 experiment in ryegrass cultivation compared to incubation-leaching, it appears that the  
397 chemical equilibrium has been established in the soil (no significant change in bulk soil pH)  
398 and, accordingly, the soil P fractions were altered minimal.

399

#### 400 **4.2 P speciation of BCs particles by XANES**

401 The predominance of Ca-hydroxyapatite in BCs as evidenced by P *K*-edge XANES analysis  
402 is consistent with findings reported by previous studies (Warren et al., 2009; Siebers et al.,  
403 2013). The mineral phase of bone consists mainly of hydroxyapatite, and its contribution to  
404 bone and bone char compositions depends on species and the age of animals (Wu et al., 2003)  
405 and carbonization temperature and residence time (Novotny et al., 2012). Bone crystallinity  
406 might be improved through structural modifications on poorly crystalline fresh bone samples  
407 (such as mineral maturity over periods of time or intensive carbonization) which can also  
408 result in increased proportions of hydroxyapatite and accordingly a decrease in P solubility  
409 (Novotny et al., 2012). Based on LC fittings, the second major component of BCs particles  
410 was CaHPO<sub>4</sub>, in good agreement with the results of Rajendran et al. (2013) who indicated the  
411 heated bones at 400 °C contained some more soluble phosphates such as CaHPO<sub>4</sub> and  
412 CaH<sub>2</sub>PO<sub>4</sub> in addition to the hydroxyapatite fraction. The authors reported that spectra of  
413 calcined bone samples at 700 °C had white-line at 2154 eV and two post-edge peaks at 2162  
414 eV and 2169 eV with no pre-edge peaks and appeared similar to CaHPO<sub>4</sub> and CaH<sub>2</sub>PO<sub>4</sub>  
415 spectra. Our LCF also assigned Ca-phytate in BCs samples which seems to be controversial  
416 as a component of animal bone materials. The P *K*-edge spectrum of Ca-phytate is very  
417 similar to other Ca-bound P compounds with a distinct white-line and lack of a pre-edge  
418 feature, although it is likely distinguishable due to specific shape of white-line tailing and  
419 absence of post-edge signal at 2164 eV (Prietz et al., 2016). Moreover, some inaccuracies in

420 LCF estimations have to be considered because of 1) uncertainty in speciation of organic P  
421 forms by *K*-edge XANES, 2) lack of reference compounds representing all P forms in BCs  
422 and 3) smaller Ca-phytate proportions than proposed 10 to 15% of  $P_t$  as detection limit for  
423 reliable XANES fittings (Beauchemin et al., 2003). Therefore, the P proportions assigned to  
424 Ca-phytate also could originate from a range of other CaP compounds.

425 In both experiments, incubation-leaching and ryegrass cropping, changes in proportions of  
426 Ca-hydroxyapatite and  $CaHPO_4$  in BC particles followed an opposite trend than in  $BC^{plus}$   
427 particles. After placement of BC particles in the soil, Ca-phosphate seemed to be released  
428 gradually over time which provides a locally lime-saturated condition. Due to elevated pH  
429 surrounding the BC particles, dissolved P can be resorbed to maintain solubility and the Ca-P  
430 equilibrium constant which likely resulted in a decreased proportion of soluble  $CaHPO_4$  and  
431 possibly the transformation into the relatively insoluble Ca-hydroxyapatite fraction. In  
432 contrast, if  $BC^{plus}$  particles were applied to soils, larger proportions of  $CaHPO_4$  at the expense  
433 of Ca-hydroxyapatite could be explained by soil acidification through the microbial oxidation  
434 of released S (Lee, et al. 1987; Fan et al., 2002). This effect was more pronounced over the  
435 longer time period in the ryegrass cropping pot experiment, favoring a greater  $CaHPO_4$  than  
436 Ca-hydroxyapatite fraction. This implies that  $BC^{plus}$  can actively supply P with predominance  
437 of soluble over insoluble P forms in the long-term and, thus, meet crop requirements.

438

### 439 **4.3 P speciation of treated soils by XANES**

440 Differences between characteristics of two soils, dissimilar mechanisms of incubation-  
441 leaching and plant uptake besides different experiment time durations complicate the joint  
442 interpretation of the P-XANES data. In unfertilized soil of the incubation-leaching  
443 experiment, the proportions of P species followed the order  $AlPO_4 > FePO_4 > SOM-Al-IHP >$   
444 Ca-hydroxyapatite which did not vary despite partial changes in some proportions after

445 application of both size fractions of BC particles. In general, these results concur with the  
446 findings by Siebers et al. (2013) that Ca-hydroxyapatite proportion was slightly increased by  
447 BC application. This could be attributed to irreversibly mixing finely-ground BC to the soil  
448 samples whereas in the present experiments the BC particles were separated from the soils  
449 before P speciation. Furthermore, these XANES data (Table 4 and Table 5) are in agreement  
450 with sequential P fractionation results (Table 1) which indicated the dominance of inorganic  
451 over organic P forms and showed the P fractions almost unchanged after BC application.  
452 Implications of low solubility of BC particles observed in this work are consistent with  
453 previous studies showing a slow release P from BCs (Warren et al., 2009; Siebers et al., 2013;  
454 Morshedizad et al., 2016). Besides reducing the  $\text{AlPO}_4$  and Ca-hydroxyapatite proportions,  
455  $\text{BC}^{\text{plus}}$  particles introduced highly soluble  $\text{Ca}(\text{H}_2\text{PO}_4)_2$  to soils in the incubation-leaching  
456 experiment. These results imply that considerable changes in P speciation were more  
457 attributed to pH reductions and, accordingly, leaching out solubilized P-forms compared with  
458 P enrichment by  $\text{BC}^{\text{plus}}$  dissolution. This is supported by results from a previous publication in  
459 which two particle sizes of  $\text{BC}^{\text{plus}}$  gave a significant rise in the leached P-concentration after  
460 1, 5, 13, 34 and 70 days of incubation along with reductions in soil pH (Morshedizad and  
461 Leinweber, 2017). This is in line with Sato et al. (2005) who found that increasing soil pH in  
462 a naturally acidic soil (pH = 4.32) was an effective approach to minimize P leaching, while  
463 pH decrease resulted in transformation of stable to soluble and more leachable P species.  
464 Regarding the XANES results of the ryegrass cultivation experiment (Table 5), the effect of  
465  $\text{BC}^{\text{plus}}$  treatment can be explained better. In the control soil, the presence of  $\text{AlPO}_4$  and  
466 increasing abundance of organic P-forms (Ca-phytate and IHP compounds) (Table 5) were  
467 consistent with the appearance of NaOH-P and HCl-P fractions by sequential extraction  
468 (Table 1). In the BC treatment the proportions of  $\text{AlPO}_4$  and Ca-phytate did not change  
469 compared to the control but the contribution of organic P increased by Mont-Al-IHP

470 formation. The stability of different P fractions can be favored by the pH effect (Gustafsson et  
471 al., 2012) likewise the dependence of BC particles solubility on the soil pH (Siebers et al.,  
472 2013). In agreement with incubation-leaching results (Table 4),  $\text{Ca}(\text{H}_2\text{PO}_4)_2$  was detected as a  
473 result of  $\text{BC}^{\text{plus}}$  amendment even though similar proportions of  $\text{AlPO}_4$  and Ca-phytate were  
474 observed between the control and  $\text{BC}^{\text{plus}}$  treatment. However, the data in Table 5 on the  
475 presence/absence of  $\text{Ca}(\text{H}_2\text{PO}_4)_2$  in soils of ryegrass experiment may have been influenced by  
476 small proportions (<10-15%; reliable detection limit by XANES, Beauchemin et al., 2003) of  
477 other simple calcium phosphates that have a spectrum similar to the one of  $\text{Ca}(\text{H}_2\text{PO}_4)_2$  in  
478 LCF analysis. The results of sequential P fractionation and XANES analyses on treatments in  
479 the two different experiments presented here demonstrated that surface modification of BC  
480 particles effectively improved soluble P fractions in  $\text{BC}^{\text{plus}}$  particles and, consequently, in  
481 amended soils.

482

## 483 **5 Conclusions**

484 In the present study, the P speciation by a sequential P fractionation and P K-edge XANES  
485 spectroscopy revealed the noticeable alteration in the P-pools of treated soil samples. Results  
486 of incubation-leaching and ryegrass cultivation experiments indicated that  $\text{BC}^{\text{plus}}$  produced by  
487 surface-modification of BC through addition of S compounds provided more soluble and  
488 plant-available P than non-modified BC during the growth season. The S oxidation and  
489 thereby the soil pH decrease seems to stimulate the P release from  $\text{BC}^{\text{plus}}$  particles. The P K-  
490 edge XANES analyses of  $\text{BC}^{\text{plus}}$  particles revealed more soluble  $\text{CaHPO}_4$  than in BC particles  
491 at the expense of Ca-hydroxyapatite. This was associated with the addition  $\text{Ca}(\text{H}_2\text{PO}_4)_2$  or  
492 similar simple Ca-P-compounds to amended soils, as indicated by sequential P fractionation  
493 and XANES analyses. Future studies will be directed to validate the beneficial effects of  
494  $\text{BC}^{\text{plus}}$  at the field scale with different soils and to optimize the surface modification of BCs.

495

496 **Data availability.** All compiled data of this study are published in figures and tables.  
497 Detailed primary data including the incubation-leaching and ryegrass cultivation results and  
498 the acquired XANES spectra will be saved and published in the BonaRes Data Center  
499 (<https://www.bonares.de/research-data>) and get the BonaRes DOI prefix (10.20387).

500

501 **Competing interests.** The authors declare that they have no conflict of interest.

502

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512

## 513 **References**

514 Ahlgren, J., Djodjic, F., Börjesson, G., and Mattsson, L.: Identification and quantification of  
515 organic phosphorus forms in soils from fertility experiments, *Soil Use Manag.*, 29,  
516 24–35, doi:10.1111/sum.12014, 2013.

517 Arai, Y. and Sparks, D. L.: Phosphate reaction dynamics in soil and soil components: a  
518 multiscale approach, *Adv. Agron.*, 94, 135–179, doi:10.1016/S0065-2113(06)94003-6,  
519 2007.

520 Beauchemin, S., Hesterberg, D., Chou, J., Beauchemin, M., Simard, R. R., and Sayers, D.E.:  
521 Speciation of phosphorus in phosphorus-enriched agricultural soils using X-ray  
522 absorption near-edge structure spectroscopy and chemical fractionation, *J. Environ.*  
523 *Qual.*, 32, 1809–1819, doi:10.2134/jeq2003.1809, 2003.

524 Condon, L. M. and Newman, S.: Revisiting the fundamentals of phosphorus fractionation of  
525 sediments and soils, *J. Soil Sediment*, 11, 830–840, doi:10.1007/s11368-011-0363-2,  
526 2011.

527 Cross, A. F. and Schlesinger, W. H.: A literature-review and evaluation of the Hedley  
528 fractionation - applications to the biochemical cycle of soil-phosphorus in natural  
529 ecosystems, *Geoderma*, 64, 197–214, doi:10.1016/0016-7061(94)00023-4, 1995.

530 De Brabandere, H., Forsgard, N., Israelsson, L., Petterson, J., Rydin, E., Waldebäk, M., and  
531 Sjöerg, P. J.: Screening for organic phosphorus compounds in aquatic sediments by  
532 liquid chromatography coupled to ICP-AES and ESI-MS/MS, *Anal. Chem.*, 80, 6689–  
533 6697, doi:10.1021/ac8006335, 2008.

534 Delgado, A., Madrid, A., Kassem, S., Andreu, L., and Campillo, M. C.: Phosphorus fertilizer  
535 recovery from calcareous soils amended with humic and fulvic acids, *Plant Soil*, 245,  
536 277–286, doi:10.1023/A:1020445710584, 2002.

537 Dieter, D., Elsenbeer, H., and Turner, B. L.: Phosphorus fractionation in lowland tropical  
538 rainforest soils in central Panama, *Catena*, 82, 118–125,  
539 doi:10.1016/j.catena.2010.05.010, 2010.

540 Fan, X., Habib, L., Fleckenstein, J., Haneklaus, S., and Schnug, E.: "In situ digestion" a  
541 concept to manage soil phosphate in organic farming, In: *Proc. of the 13th Intern.*  
542 *Fertilizer Symposium*, Tokat, Turkey, 10-13 June 2002. 219–228, 2002.

543 Fan, X., Schnug, E., Haneklaus, S., and Li, Y.: In situ digestion of rock phosphates to  
544 mobilize plant-available phosphate for organic farming, *Commun. Soil Sci. Plant*, 43,  
545 2191–2201, doi:10.1080/00103624.2012.708073, 2012.

546 Guo, F., Yost, R. S., Hue, N. V., Evensen, C. I., and Silva, J. A.: Changes in phosphorus  
547 fractions in soils under intensive plant growth, *Soil Sci. Soc. Am. J.*, 64, 1681–1689,  
548 doi:10.2136/sssaj2000.6451681x, 2000.

549 Gustafsson, J. P., Mwamila, L. B., and Kergoat, K.: The pH dependence of phosphate  
550 sorption and desorption in Swedish agricultural soils, *Geoderma*, 189–190, 304–311,  
551 doi:10.1016/j.geoderma.2012.05.014, 2012.

552 Hartley, T. N., Macdonald, A. J., McGrath, S. P., and Zhao, F. J.: Historical arsenic  
553 contamination of soil due to long-term phosphate fertiliser applications, *Environ.*  
554 *Pollut.*, 180, 259–264, doi:10.1016/j.envpol.2013.05.034, 2013.

555 Hashimoto, Y. and Watanabe, Y.: Combined applications of chemical fractionation, solution  
556 <sup>31</sup>P-NMR and P K-edge XANES to determine phosphorus speciation in soils formed  
557 on serpentine landscapes, *Geoderma*, 230, 143–150, doi:  
558 10.1016/j.geoderma.2014.04.001, 2014.

559 He, Q. B., and Singh, B. R.: Plant availability of cadmium in soils: I. Extractable cadmium in  
560 newly and long-term cultivated soils, *Acta. Agric. Scand. Sect. B*, 43, 134–141,  
561 doi:10.1080/09064719309411231, 1993.

562 Hedley, M. J., Stewart, J. W. B., and Chauhan, B. S.: Changes in organic and organic soil  
563 phosphorus fractions induced by cultivation practices and laboratory incubations, *Soil*  
564 *Sci. Soc. Am. J.*, 46, 970–976, doi:10.2136/sssaj1982.03615995004600050017x,  
565 1982.

566 Herzl, H., Krüger, O., Hermann, L., and Adam, C.: Sewage sludge ash - A promising  
567 secondary phosphorus source for fertilizer production, *Sci. Total Environ.*, 542, 1136–  
568 1143, doi:10.1016/j.scitotenv.2015.08.059, 2016.

569 Kaur, G. and Reddy, M. S.: Role of phosphate-solubilizing bacteria in improving the soil  
570 fertility and crop productivity in organic farming, *Arch. Agron. Soil Sci.*, 60, 549–564,  
571 doi:10.1080/03650340.2013.817667, 2014.

572 Kelly, S., Hesterberg, D., and Ravel, B.: Analysis of soils and minerals using x-ray absorption  
573 spectroscopy, p. 387–463. In A.L. Ulery and R. Drees (ed.) *Methods of soil analysis.*  
574 *Part 5. Mineralogical methods*, SSSA, Madison, WI., 2008.

575 Kizewski, F., Liu, Y. T., Morris, A., and Hesterberg, D.: Spectroscopic approaches for  
576 phosphorus speciation in soils and other environmental systems, *J. Environ. Qual.*, 40,  
577 751–766, doi:10.2134/jeq2010.0169, 2011.

578 Klysubun, W., Sombunchoo, P., Deenan, W., and Kongmark, C.: Performance and status of  
579 beamline BL8 at SLRI for X-ray absorption spectroscopy, *J. Synchrotron Rad.*, 19,  
580 930–936, doi:10.1107/S0909049512040381, 2012.

581 Kratz, S., Schick, J., and Schnug E.: Trace elements in rock phosphates and P containing  
582 mineral and organo-mineral fertilizers sold in Germany, *Sci. Total Environ.*, 542,  
583 1013–1019, doi:10.1016/j.scitotenv.2015.08.046, 2016.

584 Kruse, J., Abraham, M., Amelung, W., Baum, C., Bol, R., Kuhn, O., Lewandowski, H.,  
585 Niederberger, J., Oelmann, Y., Ruger, C., Santner, J., Siebers, M., Siebers, N., Spohn,  
586 M., Vestergren, J., Vogts, A., and Leinweber, P.: Innovative methods in soil  
587 phosphorus research: a review, *J. Plant Nutr. Soil Sci.*, 178, 43–88,  
588 doi:10.1002/jpln.201400327, 2015.

589 Lanfranco, A. M., Schofield, P. F., Murphy, P. J., Hodson, M. E., Mosselmans, J. F. W., and  
590 Valsami-Jones, E.: Characterization and identification of mixed-metal phosphates in



591 soils: the application of Raman spectroscopy, *Mineral. Mag.*, 67, 1299–1316,  
592 doi:10.1180/0026461036760166, 2003.

593 Lee, A., Watkinson, J. H., Orbell, G., Bagyaraj, J., and Lauren, D. R.: Factors influencing  
594 dissolution of phosphate rock and oxidation of elemental sulphur in some New  
595 Zealand soils, *New Zeal. J. Agr. Res.*, 30, 373–385, doi:10.1080/00288233.1987.  
596 10421898, 1987.

597 Liu, J. Y., Wang, H., Yang, H. J., Ma, Y. J., and Cai, O. C.: Detection of phosphorus species  
598 in sediments of artificial landscape lakes in China by fractionation and phosphorus-31  
599 nuclear magnetic resonance spectroscopy, *Environ. Pollut.*, 157, 49–56,  
600 doi:10.1016/j.envpol.2008.07.031, 2009.

601 Ma, Y. L. and Matsunaka, T.: Biochar derived from dairy cattle carcasses as an alternative  
602 source of phosphorus and amendment for soil acidity, *Soil Sci. Plant Nutr.*, 59,  
603 628–641, doi:10.1080/00380768.2013.806205, 2013.

604 McDowell, R. W. and Stewart, I.: The phosphorus composition of contrasting soils in  
605 pastoral, native and forest management in Otago, New Zealand: Sequential extraction  
606 and <sup>31</sup>P NMR, *Geoderma*, 130, 176–189, doi:10.1016/j.geoderma.2005.01.020, 2006.

607 Morshedizad, M. and Leinweber, P.: Leaching of phosphorus and cadmium in soils amended  
608 with different bone chars, *CLEAN – Soil, Air, Water*, 45 (8),  
609 doi:10.1002/clen.201600635, 2017.

610 Morshedizad, M., Zimmer, D., and Leinweber, P.: Effect of bone chars on phosphorus-  
611 cadmium-interactions as evaluated by three extraction procedures, *J. Plant Nutr. Soil*  
612 *Sci.*, 179, 388–398, doi:10.1002/jpln.201500604, 2016.

613 Murphy, J. and Riley, J. P.: A modified single solution method for the determination of  
614 phosphate in natural waters, *Anal. Chim. Acta*, 27, 31–36, doi:10.1016/S0003-  
615 2670(00)88444-5, 1962.

616 Novotny, E. H., Auccaise, R., Velloso, M. H. R., Corrêa, J. C., Higarashi, M. M., Abreu, V.  
617 M. N., Rocha, J. D., and Kwapinski, W.: Characterization of phosphate structures in  
618 biochar from swine bones, *Pesq. Agropec. Bras.*, 47, 672–676, doi:10.1590/S0100-  
619 204X2012000500006, 2012.

620 Paraskova, J. V., Jorgensen, C., Reitzel, K. Pettersson, J., Rydin, E., and Sjoberg, P. J. R.:  
621 Speciation of inositol phosphates in lake sediments by ion-exchange chromatography  
622 coupled with mass spectrometry, inductively coupled plasma atomic emission  
623 spectroscopy, and  $^{31}\text{P}$  NMR spectroscopy, *Anal. Chem.*, 87, 2672–2677,  
624 doi:10.1021/ac5033484, 2015.

625 Powers, W. L.: Sulfur in relation to soil fertility, Station Bulletin 199 of the Oregon  
626 Agricultural College Experiment Station, 45 pp, 1923, [http://ir.library.oregonstate.edu](http://ir.library.oregonstate.edu/xmlui/bitstream/handle/1957/14537/StationBulletin199.pdf?sequence=1)  
627 [/xmlui/bitstream/handle/1957/14537/StationBulletin199.pdf?sequence=1](http://ir.library.oregonstate.edu/xmlui/bitstream/handle/1957/14537/StationBulletin199.pdf?sequence=1) [last access  
628 04.10.2017].

629 Prietzel, J., Dümig, A., Wu, Y., Zhou, J., and Klysubun, W.: Synchrotron-based P K-edge  
630 XANES spectroscopy reveals rapid changes of phosphorus speciation in the topsoil of  
631 two glacier foreland chronosequences, *Geochim. Cosmochim. Acta*, 108, 154–171,  
632 doi:10.1016/j.gca.2013.01.029, 2013.

633 Prietzel, J., Harrington, G., Häusler, W., Heister, K., Werner, F., and Klysubun, W.:  
634 Reference spectra of important adsorbed organic and inorganic phosphate binding  
635 forms for soil P speciation using synchrotron-based K-edge XANES spectroscopy, *J.*  
636 *Synchrotron Rad.*, 23, 532–544, doi:10.1107/S1600577515023085, 2016.

637 Rajendran, J., Gialanella, S., and Aswath, P. B.: XANES analysis of dried and calcined bones,  
638 *Mater. Sci. Eng. C*, 33, 3968–3979, doi:10.1016/j.msec.2013.05.038, 2013.

639 Ravel, B. and Newville, M.: Athena, Artemis, Hephaestus: data analysis for X-ray absorption  
640 spectroscopy using IFEFFIT, *J. Synchrotron Rad.*, 12, 537–541,  
641 doi:10.1107/S0909049505012719, 2005.

642 Rubaek, G. H., Kristensen, K., Olesen, S. E., Ostergaard, H. S., and Heckrath, G.: Phosphorus  
643 accumulation and spatial distribution in agricultural soils in Denmark, *Geoderma*, 209,  
644 241–250, doi:10.1016/j.geoderma.2013.06.022, 2013.

645 Sato, S., Solomon, D., Hyland, C., Ketterings, Q. M., and Lehmann, J.: Phosphorus speciation  
646 in manure and manure-amended soils using XANES spectroscopy, *Environ. Sci.*  
647 *Technol.*, 39, 7485–7491, doi:10.1021/es0503130, 2005.

648 Schnug, E., Haneklaus, S., Rogasik, J., and Gassner, A.: Utilisation of fertiliser P with special  
649 regard to organic farming, In: *Proc. of the 14th Intern. Symposium of Fertilizers*,  
650 Debrecen, Hungary, 22-25 June 2003. 11–29, 2003.

651 Scholz, R. W., Ulrich, A. E., Eilittä, M., and Roy, A.: Sustainable use of phosphorus: a finite  
652 resource, *Sci. Total Environ.*, 461–462, 799–803, doi:10.1016/j.scitotenv.2013.05.043,  
653 2013.

654 Schröder, J. J., Smit, A. L., Cordell, D., and Rosemarin, A.: Improved phosphorus use  
655 efficiency in agriculture: a key requirement for its sustainable use, *Chemosphere*, 84,  
656 822–831, doi:10.1016/j.chemosphere.2011.01.065, 2011.

657 Sharpley, A. N., McDowell, R. W., and Kleinman, P. J. A.: Amounts, forms and solubility of  
658 phosphorus in soils receiving manure, *Soil Sci. Soc. Am. J.*, 68, 2048–2054,  
659 doi:10.2136/sssaj2004.2048, 2004.

660 Shen, J. B., Yuan L. X., Zhang J. L., Li H. G., Bai Z. H., Chen, X. P., Zhang, W. F., and  
661 Zhang, F. S.: Phosphorus dynamics: from soil to plant, *Plant Physiol.*, 156, 997–1005,  
662 doi:10.1104/pp.111.175232, 2011.

663 Siebers, N. and Leinweber, P.: Bone Char - A clean and renewable fertilizer with cadmium  
664 immobilizing capacity, *J. Environ. Qual.*, 42, 405–411, doi:10.2134/jeq2012.0363,  
665 2013.

666 Siebers, N., Kruse, J., and Leinweber, P.: Speciation of phosphorus and cadmium in a  
667 contaminated soil amended with bone char: sequential fractionations and XANES  
668 spectroscopy, *Water Air Soil Pollut.*, 224, 1564–1577, doi:10.1007/s11270-013-1564-  
669 7, 2013.

670 Templeton, D. M., Ariese, F., Cornelis, R., Danielsson, L. G., Muntau, H., Van Leewen, H.  
671 P., and Lobinski, R.: Guidelines for the terms related to chemical speciation and  
672 fractionation of elements. Definitions, structural aspects, and methodological  
673 approaches (IUPAC Recommendations 2000), *Pure Appl. Chem.*, 72, 1453–1470,  
674 doi:10.1351/pac200072081453, 2000.

675 Toor, G. S., Hunger, S., Peak, J. D., Sims, J. T., and Sparks, D. L.: Advances in the  
676 characterization of phosphorus in organic wastes: Environmental and agronomic  
677 applications, *Adv. Agron.*, 89, 1–72, doi:10.1016/S0065-2113(05)89001-7, 2006.

678 USEPA. Method 3051a: Microwave assisted acid dissolution of sediments, sludges, soils, and  
679 oils, 2nd ed. U.S. Gov. Print. Office, Washington, DC, 1997.

680 Van Vuuren, D. P., Bouwman, A. F., and Beusen, A. H. W.: Phosphorus demand for the  
681 1970–2100 period: A scenario analysis of resource depletion, *Globe Environ. Change*,  
682 20, 428–439, doi:10.1016/j.gloenvcha.2010.04.004, 2010.

683 Vestergren, J., Vincent, A. G., Jansson, M., Persson, P., Ilstedt, I., Gröbner, G., Giesler, R.,  
684 and Schleucher, J.: High-resolution characterization of organic phosphorus in soil  
685 extracts using 2D  $^1\text{H-P}^{31}$  NMR correlation spectroscopy, *Environ. Sci. Technol.*, 46,  
686 3950–3956, doi:10.1021/es204016h, 2012.

687 Vogel, C., Adam, C., Sekine, R., Schiller, T., Lipiec, E., and McNaughton, D.: Determination  
688 of phosphorus fertilizer soil reactions by Raman and synchrotron infrared  
689 microspectroscopy, *Appl. Spectrosc.*, 67, 1165–1170, doi:10.1366/13-07056, 2013.

690 Warren, G. P., Robinson, J. S., and Someus, E.: Dissolution of phosphorus from animal bone  
691 char in 12 soils, *Nutr. Cycling Agroecosyst.*, 84, 167–178, doi:10.1007/s10705-008-  
692 9235-6, 2009.

693 Werner, F. and Prietzel, J.: Standard protocol and quality assessment of soil phosphorus  
694 speciation by P *K*-edge XANES spectroscopy, *Environ. Sci. Technol.*, 49, 10521–  
695 10528, doi:10.1021/acs.est.5b03096, 2015.

696 Wu, Y., Ackerman, J. L., Strawich, E. S, Rey, C., Kim, H-M., and Glimcher, M. J.: Phosphate  
697 ions in bone: identification of a calcium–organic phosphate complex by <sup>31</sup>P solid-state  
698 NMR spectroscopy at early stages of mineralization, *Calcif. Tissue Int.*, 72, 610–26,  
699 doi:10.1007/s00223-002-1068-8, 2003.

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712 **Table 1.** Distribution of inorganic P (P<sub>i</sub>), organic P (P<sub>o</sub>) and total P (P<sub>t</sub>) concentrations (mg P kg<sup>-1</sup> soil)  
 713 of sequentially extracted P-fractions in the soils as affected by different treatments (treated with two  
 714 particle size fractions (1-2 and 2-4 mm) and original sizes of BC and BC<sup>plus</sup> or unfertilized soils  
 715 (control) after incubation-leaching and ryegrass cultivation experiments.

Treatment	Resin-P			NaHCO <sub>3</sub> -P			NaOH-P			HCl-P		
	P <sub>i</sub>	P <sub>o</sub>	P <sub>t</sub>	P <sub>i</sub>	P <sub>o</sub>	P <sub>t</sub>	P <sub>i</sub>	P <sub>o</sub>	P <sub>t</sub>	P <sub>i</sub>	P <sub>o</sub>	P <sub>t</sub>
<b>Incubation-leaching</b>												
Control	47	5	52	160	99	259	565	294	859	113	10	123
BC <sub>1-2</sub>	56 <sup>NS</sup>	7 <sup>NS</sup>	63 <sup>NS</sup>	163 <sup>NS</sup>	108 <sup>NS</sup>	271 <sup>NS</sup>	578 <sup>NS</sup>	303 <sup>NS</sup>	881 <sup>NS</sup>	140 <sup>NS</sup>	56 <sup>NS</sup>	196 <sup>NS</sup>
BC <sub>2-4</sub>	50 <sup>NS</sup>	7 <sup>NS</sup>	57 <sup>NS</sup>	161 <sup>NS</sup>	105 <sup>NS</sup>	266 <sup>NS</sup>	574 <sup>NS</sup>	301 <sup>NS</sup>	875 <sup>NS</sup>	121 <sup>NS</sup>	40 <sup>NS</sup>	161 <sup>NS</sup>
BC <sup>plus</sup> <sub>1-2</sub>	61 <sup>*</sup>	7 <sup>NS</sup>	68 <sup>NS</sup>	172 <sup>NS</sup>	111 <sup>NS</sup>	283 <sup>NS</sup>	593 <sup>*</sup>	313 <sup>NS</sup>	906 <sup>*</sup>	131 <sup>NS</sup>	37 <sup>NS</sup>	170 <sup>NS</sup>
BC <sup>plus</sup> <sub>2-4</sub>	50 <sup>NS</sup>	7 <sup>NS</sup>	57 <sup>NS</sup>	160 <sup>NS</sup>	104 <sup>NS</sup>	264 <sup>NS</sup>	574 <sup>NS</sup>	298 <sup>NS</sup>	872 <sup>NS</sup>	115 <sup>NS</sup>	21 <sup>NS</sup>	135 <sup>NS</sup>
<b>Ryegrass cropping experiment</b>												
Control	4	4	8	25	27	52	75	121	196	28	5	33
BC	2 <sup>NS</sup>	5 <sup>NS</sup>	7 <sup>NS</sup>	24 <sup>NS</sup>	32 <sup>NS</sup>	56 <sup>NS</sup>	79 <sup>NS</sup>	125 <sup>NS</sup>	204 <sup>NS</sup>	30 <sup>NS</sup>	6 <sup>NS</sup>	36 <sup>NS</sup>
BC <sup>plus</sup>	6 <sup>*</sup>	5 <sup>NS</sup>	11 <sup>*</sup>	35 <sup>NS</sup>	27 <sup>NS</sup>	62 <sup>NS</sup>	85 <sup>**</sup>	128 <sup>NS</sup>	213 <sup>NS</sup>	34 <sup>NS</sup>	7 <sup>NS</sup>	41 <sup>NS</sup>

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 717 \* Significant at  $P < 0.05$ ; \*\* Significant at  $P < 0.01$ ; <sup>NS</sup> Non-significant difference (treatment vs. control; Tukey-Test)  
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731 **Table 2.** Results of linear combination fitting (LCF) conducted on P *K*-edge XANES spectra of bone  
 732 char (BC) and surface modified bone char (BC<sup>plus</sup>) particles before and after a 70-days incubation-  
 733 leaching period. [These best fits were achieved using all possible combinations with nineteen spectra of](#)  
 734 [P reference compounds.](#)

Reference compound	Before experiment				After 70 days incubation-leaching			
	BC		BC <sup>plus</sup>		BC		BC <sup>plus</sup>	
	1-2 mm	2-4 mm	1-2 mm	2-4 mm	1-2 mm	2-4 mm	1-2 mm	2-4 mm
Ca-hydroxyapatite (%)	58± 6	64±5	62± 5	58± 5	75± 4	85± 3	59± 5	60± 6
CaHPO <sub>4</sub> (%)	24± 5	19± 4	28± 4	32± 6	14± 3	5± 2	33± 4	35± 5
Ca-phytate (%)	18± 4	17± 4	10± 3	10± 3	11± 3	10± 2	8± 4	5± 4
<i>R</i> -factor	0.012	0.008	0.007	0.009	0.005	0.002	0.009	0.010

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753 **Table 3.** Results of linear combination fitting (LCF) conducted on P *K*-edge XANES spectra of bone  
 754 char (BC) and surface modified bone char (BC<sup>plus</sup>) particles before and after 230-days of ryegrass  
 755 cultivation in a pot experiment. [These best fits were achieved using all possible combinations with](#)  
 756 [nineteen spectra of P reference compounds.](#)

Reference compound	Before experiment		After 230 days ryegrass cultivation	
	BC	BC <sup>plus</sup>	BC	BC <sup>plus</sup>
Ca-hydroxyapatite (%)	63± 6	70± 4	75± 4	49± 8
CaHPO <sub>4</sub> (%)	29± 5	29± 3	17± 4	43± 6
Ca-phytate (%)	8± 4	1± 3	8± 3	8± 5
<i>R</i> -factor	0.012	0.005	0.006	0.018

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775 **Table 4.** Results of linear combination fitting (LCF) conducted on P *K*-edge XANES spectra of  
 776 unfertilized (control) and fertilized soils with bone char (BC) and surface modified bone char (BC<sup>plus</sup>)  
 777 particles in the 70-days incubation-leaching experiment. [These best fits were achieved using all](#)  
 778 [possible combinations with nineteen spectra of P reference compounds.](#)

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Reference compound	Control	BC treatment		BC <sup>plus</sup> treatment	
		1-2 mm	2-4 mm	1-2 mm	2-4 mm
Ca-hydroxyapatite (%)	8± 1	4± 1	8± 1	0	0
AlPO <sub>4</sub> amorphous (%)	42± 1	42± 2	40± 1	27± 1	24± 1
FePO <sub>4</sub> (%)	29± 1	31± 2	31± 1	27± 1	26± 1
SOM-Al-IHP (%)	21± 2	23± 4	21± 3	26± 2	21± 1
Ca(H <sub>2</sub> PO <sub>4</sub> ) <sub>2</sub> (%)	0	0	0	20± 1	29± 1
<i>R</i> -factor	0.0003	0.0007	0.0003	0.0005	0.0004

801 **Table 5.** Results of linear combination fitting (LCF) conducted on P *K*-edge XANES spectra of  
 802 unfertilized (control) and fertilized soils with bone char (BC) and surface modified bone char (BC<sup>plus</sup>)  
 803 particles under 230-days of ryegrass cultivation in a pot experiment. [These best fits were achieved](#)  
 804 [using all possible combinations with nineteen spectra of P reference compounds.](#)

Reference compound	Control	BC treatment	BC <sup>plus</sup> treatment
AlPO <sub>4</sub> amorphous (%)	35± 3	35± 3	34± 1
Ca-phytate (%)	27± 3	28± 3	27± 1
IHP (%)	29± 5	21± 7	0
Mont-Al-IHP (%)	0	16± 1	0
SOM-Al-IHP (%)	0	0	25± 2
Mont-Al-PO <sub>4</sub> (%)	9± 1	0	0
Ca(H <sub>2</sub> PO <sub>4</sub> ) <sub>2</sub> (%)	0	0	14± 2
<i>R</i> -factor	0.0006	0.0008	0.0006

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839 **Figure captions**

840 **Figure 1.** Normalized P *K*-edge XANES spectra of different BC and BC<sup>plus</sup> particle sizes (1-2 and 2-4  
841 mm) before (control) and after 70-days incubation-leaching experiment compared to the reference  
842 compounds selected using LCF method.

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844 **Figure 2.** P *K*-edge XANES spectra of BC and BC<sup>plus</sup> particles before (control) and after 230-days of  
845 ryegrass cultivation compared to the reference compounds selected by LCF method.

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847 **Figure 3.** P *K*-edge XANES spectra of unfertilized (control) and fertilized soils with BC and BC<sup>plus</sup>  
848 particles under 70-days incubation-leaching experiment compared to the reference compounds  
849 selected by LCF method.

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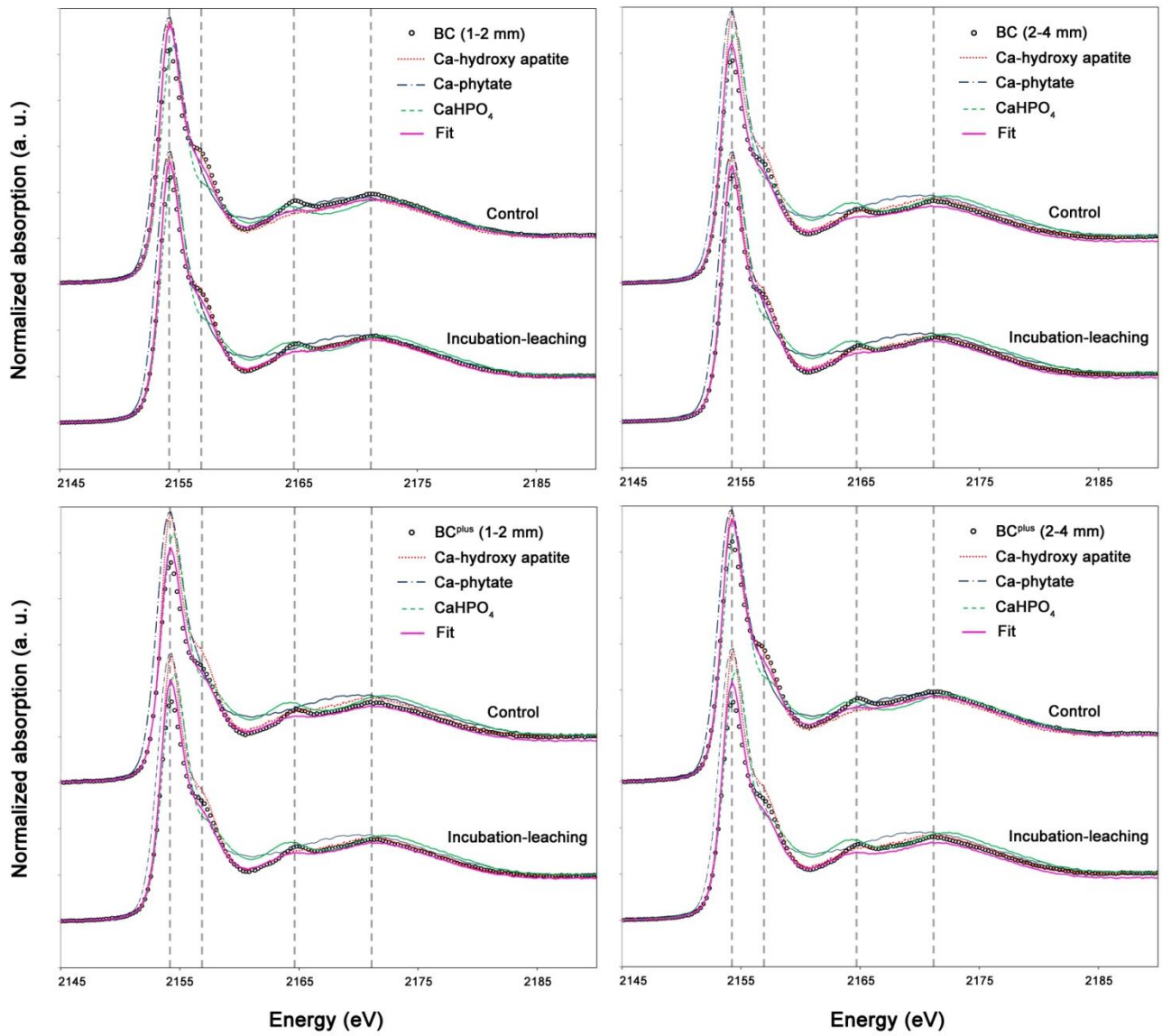
851 **Figure 4.** P *K*-edge XANES spectra of unfertilized (control) and fertilized soils with BC and BC<sup>plus</sup>  
852 particles under 230-days of ryegrass cultivation compared to the reference compounds selected by  
853 LCF method.

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859 Morshedizad et al., Figure 1

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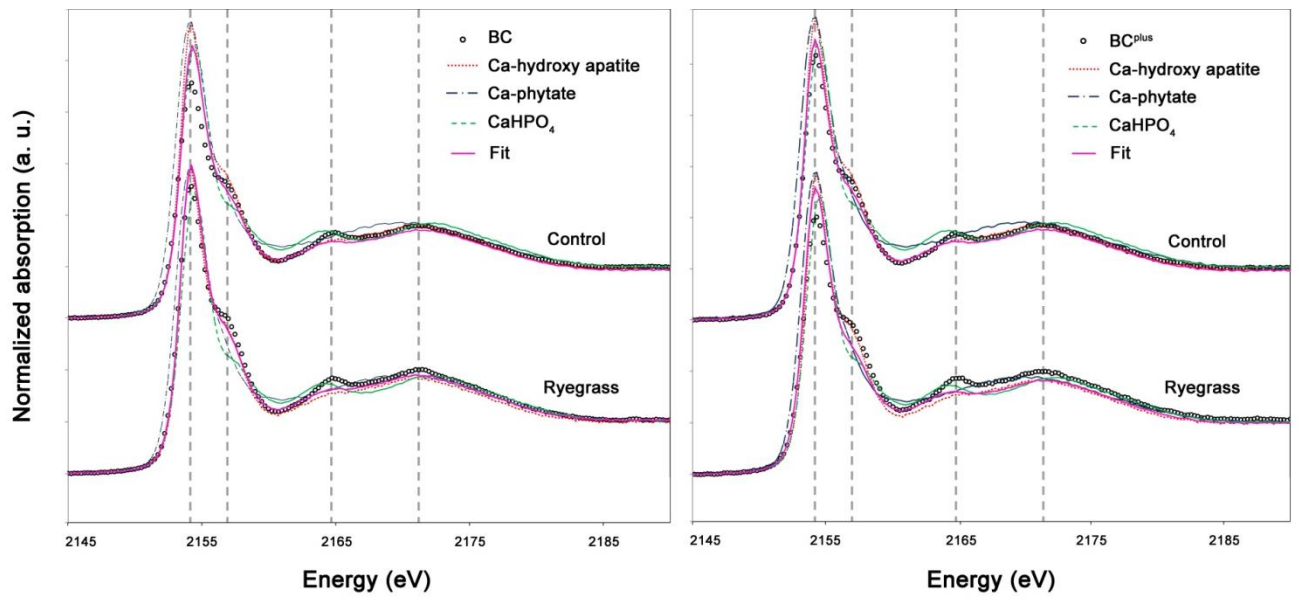
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868 Morshedizad et al., Figure 2

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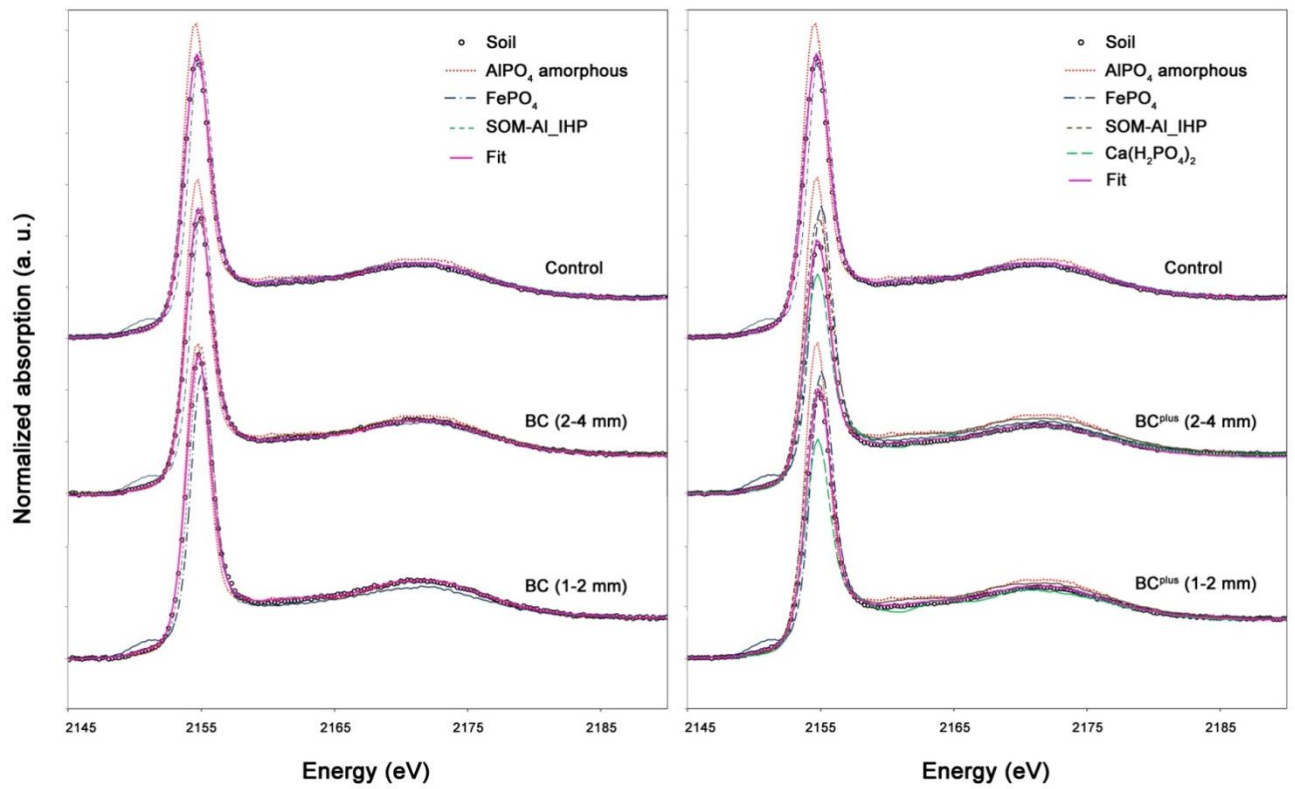
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876 Morshedizad et al., Figure 3

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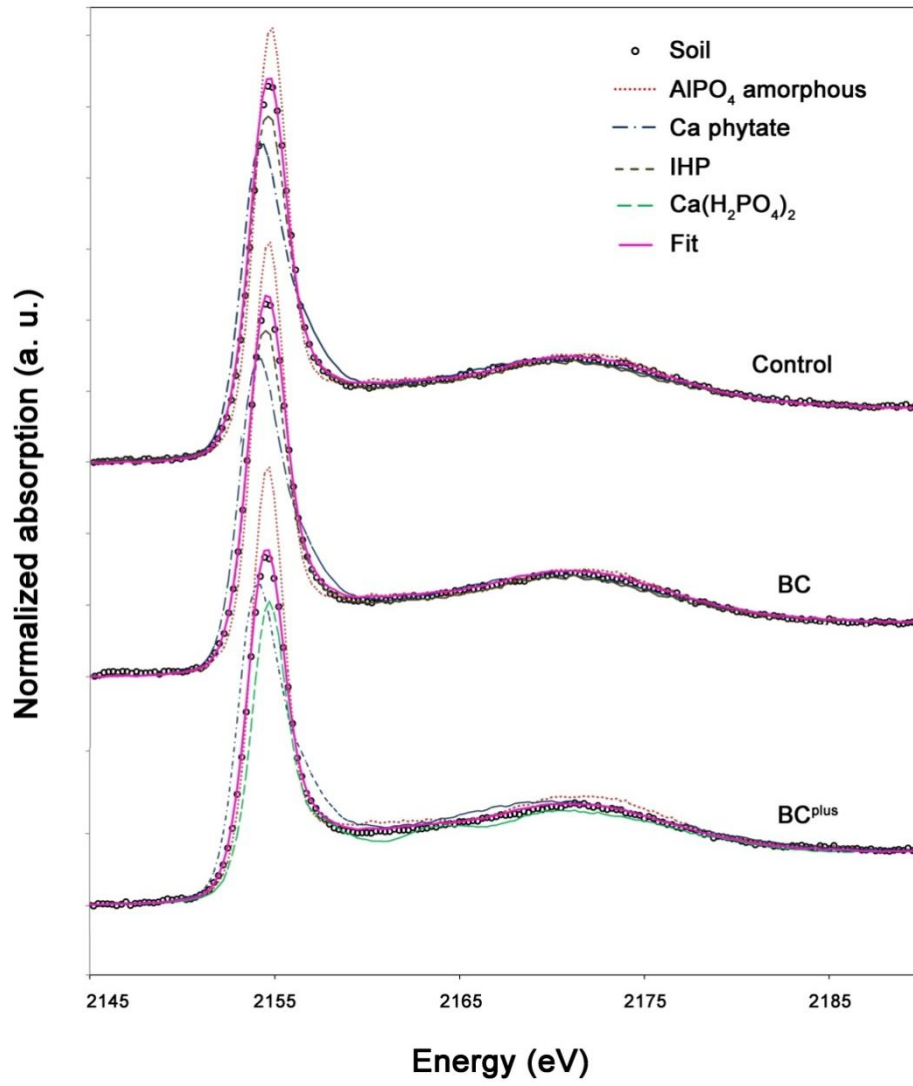
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890 Morshedizad et al., Figure 4

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