

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21

Bone char effects on soil: sequential fractionations and XANES spectroscopy

Mohsen Morshedizad¹, Kerstin Panten², Wantana Klysubun³, and Peter Leinweber¹

¹Soil Science, Faculty of Agricultural and Environmental Sciences, University of Rostock, Justus-von-Liebig Weg 6, 18051 Rostock, Germany

²Institute for Crop and Soil Science, Julius Kühn Institute, Bundesallee 50, D-38116 Braunschweig, Germany

³Synchrotron Light Research Institute, 111 University Avenue, Muang District, Nakhon Ratchasima 30000, Thailand

Correspondence to: Peter Leinweber (peter.leinweber@uni-rostock.de)

22 **Abstract.** Acceptability of novel bone char fertilizers depends on their P release but
23 reactions at bone char surfaces and impacts to soil P speciation are insufficiently known. By
24 sequential fractionation and synchrotron-based X-ray absorption near edge structure
25 (XANES) spectroscopy we investigated whether and how the chemical composition of bone
26 char particles has been altered in soil and, consequently, has affected the P speciation of
27 amended soils. Therefore, two different kinds of bone char particles (BC, produced by
28 pyrolysis of degreased animal bone chips at 800 °C and BC^{plus}, a BC enriched with reduced
29 sulfur compounds) were manually separated from the soil at the end of two different
30 experiments: “incubation-leaching” and “ryegrass cultivation”. Sequential P-fractionation of
31 amended soils showed P-enrichment in all fractions as compared to the control. The most P
32 increase between all treatments significantly occurred in the NaOH-P and resin-P fractions in
33 response to BC^{plus} application in both, incubation-leaching and ryegrass cultivation
34 experiments. This increase of the readily available P fraction in BC^{plus} treated soils was
35 confirmed by linear combination fitting (LCF) analysis on P K-edge XANES spectra of BCs
36 particles and amended soils. The proportion of Ca-hydroxyapatite decreased, whereas the
37 proportion of CaHPO₄ increased in BC^{plus} particles after amended soils had been incubated
38 and leached and cropped by ryegrass. Based on P XANES speciation as determined by LCF
39 analysis, the proportion of inorganic Ca(H₂PO₄)₂ increased in amended soils after BC^{plus}
40 application. These results indicated that soil amendment with BC^{plus} particles leads to elevated
41 P concentration and maintains more soluble P species than BC particles even after 230-days
42 of ryegrass cultivation.

43

44

45

46 **1 Introduction**

47 At the global scale readily-available sources of phosphorus (P), a crucial macro-nutrient
48 element for agricultural production, are being faced with scarcity and overpricing (Scholz et
49 al., 2013; Van Vuuren et al., 2010). Environmental pollutions frequently arise from their
50 impurities (Cd, U) (Hartley et al., 2013; Kratz et al., 2016), and from over-fertilization
51 (Rubaek et al., 2013). Further problems are the quick formation of stable and inaccessible
52 compounds that limit plant P uptake (Shen et al., 2011) and the low agronomic efficiency of
53 not more than 15% of fertilizer P in the first year of application (Schnug et al., 2003). Many
54 recent studies have targeted sustainable agriculture through improving P-availability from
55 applied fertilizers (Delgado et al., 2002; Schröder et al., 2011), increasing P-uptake efficiency
56 from organic and inorganic P-pools in the soil (Fan et al., 2002; Kaur and Reddy, 2014) and
57 developing new technologies for P-recycling from human and animal waste (Siebers and
58 Leinweber, 2013; Herzel et al., 2016).

59 As an economically and environmentally attractive example, pyrolyzed animal bone chips
60 branded as “bone char” (BC), a slow-release apatite-based P-fertilizer, have been surface
61 modified by sulfur (S) compounds to enhance its solubility in neutral to alkaline soils.
62 Incubation-leaching and pot experiments confirmed that surface-modification was an effective
63 approach in P-release promotion from BC fertilizer (Morshedizad et al., 2016; Zimmer, D.
64 and Panten, K., personal communication). Despite these attempts to raise dissolution and use-
65 efficiency of BC in supplying P for crop requirements, a considerable fraction of applied BC-
66 P to the soil remains insoluble in the short term and is not taken up by plants over the entire
67 cropping period. A detailed P speciation can clarify the fate of insoluble P from BC which has
68 not been done before.

69 Chemical speciation is described as analytical identification of chemical species of defined
70 elements and measuring their quantities in the system (Templeton et al., 2000). The precise

71 characterization of various P species in the soil as a dynamic response to non-equilibrium
72 conditions imposed by human activities such as fertilization can support a better
73 understanding of reactivity, stability and particularly plant-accessibility of different P-forms
74 and provide a basis for best management practices. Several techniques such as sequential
75 fractionation (Dieter et al., 2010; Condon and Newman, 2011), nuclear magnetic resonance
76 (NMR) spectroscopy (Liu et al., 2009; Vestergren et al., 2012; Ahlgren et al., 2013), Raman
77 spectroscopy (Lanfranco, 2003; Vogel et al., 2013) and chromatography coupled to mass
78 spectroscopy (De Brabandere et al., 2008; Paraskova et al., 2015) have been developed for P
79 speciation analysis in soil and sediments. Each one of these techniques can offer specific
80 advantages and disadvantages depending on phase and complexity of sample matrixes (Kruse
81 et al., 2015). Complementarily, X-ray absorption near edge structure (XANES) spectroscopy
82 is well-suited for identification of various P species through fingerprinting of molecular
83 structures in solid and heterogeneous mediums based on fine feature and position of
84 absorbing edges (Kelly et al., 2008; Kizewski et al., 2011). The advantages of XANES
85 spectroscopy for soil samples make it a promising technique for direct and *in situ* P-speciation
86 with no pretreatment and minimum sample manipulation (Toor et al., 2006; Kelly et al.,
87 2008).

88 According to the best of our knowledge, no studies have characterized P-speciation changes
89 in BC particles over non-equilibrium conditions in the soil system, and only few
90 investigations have been reported on the P-release from BC and alteration in P-species of the
91 soil. Siebers et al. (2013) investigated K-edge XANES spectroscopy on BC-incubated soil
92 samples and provided evidence that the increase in extractable Ca- and Mg-phosphate
93 fractions were related to the contribution of hydroxyapatite (HAP) increase after BC
94 application. Accordingly, the objective of this study was to provide practical information on
95 the fate and alteration of P-species in BC and novel surface modified BC (BC^{plus}) particles

96 and their treated soils under incubation-leaching and ryegrass cultivation practices, using
97 sequential P-fractionation and P-XANES spectroscopy.

98

99 **2 Materials and methods**

100 **2.1 Incubation-leaching experiment**

101 Two particle size fractions (1-2 and 2-4 mm) of bone chars (BC, produced by pyrolysis of
102 degreased animal bone chips at 800°C and BC^{plus} as a surface modified BC obtained by
103 blending with reduced S-containing compounds composed of 60 % elemental S, 30 %
104 calcium sulfate dehydrate and 10 % methansulfonate (Zimmer et al. unpublished results of S
105 X-ray absorption near-edge fine structure spectroscopy) in a commercial biogas
106 desulfurization process; patent application DE 212012000046U1; [www.google.com/patents](http://www.google.com/patents/DE212012000046U1?cl=en&hl=de)
107 [/DE212012000046U1?cl=en&hl=de](http://www.google.com/patents/DE212012000046U1?cl=en&hl=de)) were incubated with a silt loam soil. The soil was
108 classified as *Dystric Cambisol* (FAO) with pH of 4.7 (measured in 0.01 mol L⁻¹ CaCl₂
109 solution) and total (digestion with HNO₃ and analyzed using ICP-OES; USEPA, 1997) and
110 available (extracted by 1 mol L⁻¹ NH₄NO₃ and analyzed using ICP-OES; He and Singh, 1993)
111 P-contents of 1.6 g P kg⁻¹ and 14 mg P kg⁻¹, respectively. The BC and BC^{plus} contained total P
112 of 149 and 123 g kg⁻¹, total calcium (Ca) 185 and 265 g kg⁻¹, total S 6 and 199 g kg⁻¹ and had
113 average of pH_{CaCl2} values 7.8 and 4.9, respectively.

114 The BCs were added to 30 g of air dry soil (< 2 mm) at the levels of 0 mg P kg⁻¹ soil (control)
115 and 500 mg P kg⁻¹ soil in five replicates. The soil and BCs mixture was homogenized and
116 packed into glass columns with 10 cm length and inner diameter of 2 cm. A P-free filter (MN
117 616 G; Macherey-Nagel GmbH & Co., KG Düren, Germany) was placed at the bottom of
118 each column to avoid any particle losses. The amended soils were incubated for 70 days at 20
119 °C in the dark and constant soil moisture between 60-70 % of soil water holding capacity.
120 During the incubation period, the soil columns were leached with three pore volumes of

121 deionized water added by a droplet irrigation simulator system. The leaching process was
122 repeated in five steps, each one after 1, 5, 13, 34 and 70 days. The P-concentrations in
123 collected leachates were measured using inductively coupled plasma-optical emission
124 spectrometry (ICP-OES). Outcomes of the leaching experiment were described in
125 Morshedizad and Leinweber (2017). After the incubation-leaching experiment, the treated soil
126 samples were carefully removed from the glass columns, air dried and BCs particles were
127 manually separated from the soils very gently. The BCs particles were delicately washed with
128 deionized water to remove adhered soil particles, allowed to dry completely at ambient
129 conditions and finely ground for further analyses.

130

131 **2.2 Pot experiment with annual ryegrass**

132 The same BC and BC^{plus} as described for the incubation-leaching experiment were used in
133 original sizes (mostly between 1 to 5 mm) for P fertilization of annual ryegrass in a pot
134 experiment. The experiment was set-up using an acidic sandy silt soil with available P-content
135 of 24.2 mg P kg⁻¹ and pH of 5.2. The pot experiment was set up by adding BC and BC^{plus} at
136 the levels of 0 mg P kg⁻¹ (control) and 280 mg P kg⁻¹ into the 6 kg of the soil dry matter in
137 each pot and in four replicates arranged in a complete randomized block. After four weeks of
138 incubation at field capacity water content and ambient temperature conditions, 30 seeds of
139 annual ryegrass per each pot were sown on 13th of May 2016. The experiment was conducted
140 in a glasshouse under ambient air and temperature conditions and the soil moisture was
141 maintained at field capacity during the whole experiment. All other essential nutrients were
142 sequentially added at sufficient levels before seeding and after each six cuts of ryegrass
143 between 23rd of June and 3rd of November 2016. Finally, after the last harvest (7th), plant parts
144 (shoots and roots) were dried at 60 °C and BCs particles were manually separated from the
145 soils as they could be detected visually by their size and dark color very gently, using

146 tweezers. Then these particles were washed delicately with deionized water to remove
147 attached soil particles, allowed to dry completely at ambient conditions and finely ground to
148 fine powders for further analyses.

149

150 **2.3 Sequential phosphorus fractionation**

151 Soil samples were sequentially extracted based on chemical solubility in order according to a
152 modified Hedley et al. (1982) procedure. After BC particles detachment, duplicate 0.5 g fine-
153 ground and air-dried soil samples were weighed into 50-mL centrifuge tubes. In summary,
154 chemical P fractionation includes the following steps:

155 1) The mobile and readily available P fraction was extracted with resin strips (saturated in 0.5
156 M NaHCO₃) after 18 hours end-over-end shaking in 30 mL deionized water. The resin strips
157 were separated from solids/solution and washed using 50 mL of 1 M HCl to remove absorbed
158 P. The soil suspension was centrifuged at 2500 × g for 20 minutes and the supernatant was
159 decanted.

160 2) Next, the labile inorganic and organic fractions weakly absorbed to mineral surfaces and
161 some microbial P were extracted by 30 mL of 0.5 M NaHCO₃, 18 hours end-over-end shaking
162 and centrifugation at 2500 × g for 20 minutes. The supernatant was filtered ([Whatman no. 42](#)
163 [filter](#)) and collected for measurements.

164 3) The inorganic P adsorbed and bound to Al- and Fe-oxide minerals and organic P from
165 humic substances were extracted using 30 mL of 0.1 M NaOH solution and repeating the
166 second step procedure as described above.

167 4) The relatively insoluble fraction of P bound to Ca and Mg minerals and apatite was
168 extracted by 30 mL of 1 M HCl in the same way as for the previous steps.

169 Total P concentrations (P_t) and inorganic P (P_i) in all extracts were measured by ICP-OES and
170 colorimetrically (molybdenum blue method; Murphy and Riley, 1962), respectively. The
171 organic P (P_o) concentrations were calculated by $P_t - P_i$.

172

173 **2.4 Phosphorus K-edge XANES analysis**

174 The XANES data collection for characterizing P-species in all soil samples and BCs particles
175 was acquired at the Synchrotron Light Research Institute (SLRI) in Nakhon Ratchasima,
176 Thailand, on the beamline 8 (BL8) of the electron storage ring with a covering photon energy
177 from 1.25 to 10 KeV, electron energy operated at 1.2 GeV and beam current of 80-150 mA
178 (Klysubun et al., 2012). [The P K-edge XANES spectra were collected from dried and very](#)
179 [finely ground treated soils and particulate BCs samples](#) which had been diluted to P
180 concentrations $< 10 \text{ mg P kg}^{-1}$ with SiO_2 powder (to eliminate self absorption effects; Prietzel
181 et al., 2013), again ground in agate stone mini-mortar and spread uniformly as a thin layer on
182 P-free kapton tape (Lanmar Inc., Northbrook, IL, USA). Data collection was operated in
183 standard conditions comprising the energy calibration by standard pure elemental P and
184 allocating the reference energy (E_0) at 2145.5 eV using the maximum peak of spectrum. All
185 spectra were recorded at photon energies between 2045.5 and 2495.5 eV in step sizes of 5 eV
186 (2045.5 to 2105.5 eV and 2245.5 to 2495.5 eV), 1 eV (2105.5 to 2135.5 eV and 2195.5 to
187 2245.5 eV) and 0.25 eV (2135.5 to 2195.5 eV) with a 13-channel germanium detector [in](#)
188 [fluorescence mode](#). At least three scans were collected and averaged for each sample.

189 The P-XANES spectra were normalized and after merging replicates, a linear combination
190 fitting (LCF) was performed using the ATHENA software package (Ravel and Newville,
191 2005). [All XANES spectral data were baseline corrected in the pre-edge region between](#)
192 [2115–2145 eV and normalized in the post-edge region of 2190–2215 eV similar to those were](#)
193 [used for the reference P K-edge XANES spectra \(Prietzel et al. 2016\) to achieve consistency](#)

194 in the following fitting analysis. To achieve the best compatible set of references with each
195 specified sample spectrum, LCF analysis was performed in the energy range between -20 eV
196 and +30 eV relative to the E_0 using the combinatorics function of ATHENA software to attain
197 all possible binary, ternary and at most quaternary combinations between all nineteen P
198 reference spectra. The following set of reference P *K*-edge XANES spectra, all recorded in
199 SLRI under the same adjustments by Werner and Prietzel (2015) and Prietzel et al. (2016),
200 were used for fitting and calculations; Ca-, Al- and Fe-phytate, non-crystalline and crystalline
201 $AlPO_4$, non-crystalline and crystalline $FePO_4 \cdot 2H_2O$, Ca-hydroxyapatite ($Ca_5(OH)(PO_4)_3$),
202 inositol hexakisphosphate (IHP), ferrihydrite-IHP, montmorillonite-Al-IHP, soil organic
203 matter (SOM)-Al-IHP, ferrihydrite-orthophosphate, montmorillonite-Al-orthophosphate,
204 SOM-Al-orthophosphate, bohemite-IHP, bohemite-orthophosphate, $CaHPO_4$, $Ca(H_2PO_4)_2$
205 and $MgHPO_4$. To select the best possible combination fit between the sample spectrum and
206 the P-reference spectra, the lowest reduced chi value (χ^2) and *R*-factor were chosen.

207

208 **3 Results**

209 **3.1 Effect of BCs on sequentially extracted P after incubation-leaching**

210 After 70 days of incubation-leaching, the sequential P-fractionation of amended soils showed
211 variations in the amount and distribution of various P-fractions between different treatments
212 (Table 1). For all treatments, NaOH extracted the majority of fractionated P (62.4 to 66.5 %
213 of total fractionated P), followed by the labile P fraction ($NaHCO_3$, 19.2 to 20.0 %), HCl-P
214 (9.5 to 13.9 %) and the readily available P (resin strips, 4.1 to 4.8 %). The BCs addition
215 increased the total soil P pools although the difference was significant only for the $BC^{plus}_{1-2\text{ mm}}$
216 and $BC_{1-2\text{ mm}}$ treatments. The largest increase in total fractionated P_t (resin- P_t + $NaHCO_3$ - P_t +
217 $NaOH$ - P_t + HCl- P_t) occurred in $BC^{plus}_{1-2\text{ mm}}$ (133.8 mg P kg^{-1} soil) followed by $BC_{1-2\text{ mm}}$

218 (118.6 mg P kg⁻¹ soil), BC_{2-4 mm} (67.1 mg P kg⁻¹ soil) and BC^{plus}_{2-4 mm} (35.7 mg P kg⁻¹ soil),
219 compared to the control soil.

220

221

INSERT Table 1 HERE

222

223 The proportion of P-enrichment in each fraction varied between different treatments in the
224 order NaOH-P > HCl-P > NaHCO₃-P > resin-P for BC^{plus}_{1-2 mm} and BC^{plus}_{2-4 mm}. For the BC<sub>1-
225 2 mm</sub> and BC_{2-4 mm} treatment the order was HCl-P > NaOH-P > NaHCO₃-P > resin-P. In all
226 treatments, the P_i proportions in each of the P fractions were greater than the P_o proportions.
227 Compared to the control soil, the most P_i increase was observed in NaOH-P_i and resin-P_i in
228 response to BC^{plus}_{1-2 mm} application (Table 1). Moreover, after 70 days of incubation-leaching,
229 soil pH increased in BC treatments whereas BC^{plus} amendments had an acidifying effect. Soil
230 pH levels of BC_{1-2 mm} and BC_{2-4 mm} increased by 0.07 and 0.05 units and decreased for BC^{plus}<sub>1-
231 2 mm</sub> and BC^{plus}_{2-4 mm} treatments by 0.21 and 0.15 units compared to unamended control soil
232 (pH = 5.06).

233

234 3.2 Effect of BCs on sequentially extracted P after ryegrass cropping

235 Sequentially extracted P fractions in soil varied between different treatments after 230 days of
236 ryegrass cropping (Table 1). In all treatments (control, BC and BC^{plus}), NaOH-P was the
237 largest P pool mainly associated with Al- and Fe-oxide minerals and humic substances (65.0
238 to 67.5 % of total fractionated P) followed by the NaHCO₃-P (18.2 to 19.0 %), HCl-P (11.5 to
239 12.6 %) and resin-P (2.2 to 3.4 %) fractions. Enrichments of P fractions in BC^{plus} treatments
240 were more pronounced than in treated soils with BC particles. In this treatment the
241 concentrations of readily available and labile inorganic P fractions were insignificantly
242 smaller than in the control. Additionally, a significant increase in P concentration was

243 obtained only in resin-P_i and NaOH-P_i fractions of the BC^{plus} treated soil (Table 1). The
244 maximum increase of total fractionated P was obtained in BC^{plus} treatment (37.6 mg P kg⁻¹
245 soil). In comparison to incubation-leaching results, a similar sequence was observed for the
246 order of increasing magnitude of P fractions in response to BC and BC^{plus} amendments
247 (NaOH-P > NaHCO₃-P > HCl-P > resin-P). However, for BC treatment, the total P extracted
248 by resin strips was lowered in comparison with the control. In the control and BC treatments,
249 P_o was the predominant form in NaOH-P and NaHCO₃-P fractions while for BC^{plus} it was
250 only in the NaOH-P fraction. Each P fraction was highest under BC^{plus} application, except for
251 NaHCO₃-P_o of BC treatment.

252 Separately, the effect of BC and BC^{plus} application on ryegrass yield parameters was
253 examined in the 230-days pot experiment. The results indicated that P uptake, ryegrass yield
254 and apparent nutrient recovery efficiency (ANR) of BC^{plus} treatments exceeded that of BC and
255 control treatments and increased to values comparable with triple super phosphate (TSP)
256 fertilizer (Zimmer, D. and Panten, K., personal communication). Addition of BC and BC^{plus}
257 did not significantly change the bulk soil pH, although local acidification around BC^{plus}
258 particles (pH 4.9; Morshedizad and Leinweber, 2017) probably can lower soil pH in small
259 scale areas compared to BC treatments (pH about 8).

260

261 **3.3 XANES analysis of BCs particles**

262 All spectra from BCs were characterized by an intense white-line peak, post-edge position
263 and without a distinct pre-edge which corresponded to calcium phosphate compounds
264 including Ca-hydroxyapatite, dicalcium phosphate (CaHPO₄) and Ca-phytate (Fig. 1). The P
265 K-edge XANES results indicated no obvious alterations in spectral features of BCs particles
266 after the incubation-leaching experiment. After 70-days of incubation-leaching, the BC
267 spectra were shifted towards Ca-hydroxyapatite, and this was more pronounced for the 2-4

268 mm than for the 1-2 mm BC-particles. The opposite trend was the case for BC^{plus} particles
269 where the white-line signal intensity decreased after incubation-leaching period and the post-
270 edge of spectra tended more to dicalcium phosphate. This effect was stronger for BC^{plus}-
271 particle size reduction from 2-4 mm to 1-2 mm.

272

273 **INSERT Figure 1 HERE**

274

275 To quantify the P speciation of BC and BC^{plus} particles, LCF analyses using all possible
276 combinations were performed on all P *K*-edge XANES spectra (Table 2). The fitting results
277 indicated that untreated BC and BC^{plus} particles before the experiment contained on average
278 61 and 60 % Ca-hydroxyapatite, 22 and 30 % CaHPO₄ and 18 and 10 % Ca-phytate. After 70-
279 days of incubation-leaching, the proportion of Ca-hydroxyapatite increased to the average of
280 80% in BC while it remained unchanged in BC^{plus} particles. The CaHPO₄ proportion increased
281 in BC^{plus} particles to the average of 34 % whereas the lower content was assigned in the
282 spectra of BC particles accounting for 10 % of total P species. Moreover, the Ca-phytate
283 proportion decreased slightly in BC and BC^{plus} particles from about 18 and 10% to averages
284 of 11 and 7 %, respectively.

285

286 **INSERT Table 2 HERE**

287

288 Spectra of BC and BC^{plus} particles, before and after 230-days of ryegrass cultivation, were
289 characterized by a sharp white-line followed by a shoulder and then a post-edge feature
290 between 2160 and 2175 eV which was divided into two peaks (Fig. 2). These features were
291 most similar to P *K*-edge XANES spectra of Ca-hydroxyapatite, CaHPO₄ and Ca-phytate
292 standard compounds. Treated BC particles had a white-line with higher intensity which

293 appeared more similar to the Ca-hydroxyapatite spectrum. In contrast, BC^{plus} particles under
294 ryegrass cultivation showed a weaker white-line exhibiting the shoulder and post-edge feature
295 more comparable to the CaHPO₄ spectrum.

296

297 **INSERT Figure 2 HERE**

298

299 Some differences in proportions of P species observed between BCs particles before and after
300 the cropping [period in the ryegrass pot](#) experiment are presented in Table 3. The LCF results
301 revealed overall contributions of 63 and 70 % Ca-hydroxyapatite, 29 and 29% CaHPO₄ and 8
302 and 1 % Ca-phytate in the original BC and BC^{plus}, respectively. After the cropping period, the
303 percentage of Ca-hydroxyapatite was increased in BC particles. In the BC^{plus} treatment, the
304 percentage of CaHPO₄ increased from 29 to 43, while the percentage of Ca-hydroxyapatite
305 was reduced from 70 to 49 %. The Ca-phytate proportion remained unchanged in BC particles
306 while that of BC^{plus} increased from 1 to 8 % after the ryegrass cultivation period.

307

308 **INSERT Table 3 HERE**

309

310 **3.4 XANES analysis of soil samples**

311 The P *K*-edge XANES spectra of soil samples from the incubation-leaching experiment
312 showed two dominant features including 1) a strong white-line lacking pre-edge and shoulder
313 and 2) a tailed post-edge feature (Fig. 3). The most similarity to these features was seen in
314 XANES spectra of amorphous AlPO₄, FePO₄ and SOM-Al-IHP compounds. Distinct
315 differences appeared between the control and treated soil with BC^{plus}, not with BC treatments.
316 This was reflected by slightly lower intensities of both white-line and post-edge features.

317

318

INSERT Figure 3 HERE

319

320 The P species of treated soils in the incubation-leaching experiment were determined by LCF
321 analysis to select at most four reference compounds in combinatorics of all possible fitting
322 combinations (Table 4). The fitting results indicated that P in the control soil and BC
323 treatments occurred dominantly as AlPO_4 amorphous ($\approx 40\%$), FePO_4 ($\approx 30\%$) and SOM-
324 Al-IHP ($\approx 20\%$) compounds. In BC^{plus} treated soils, the average proportion of amorphous
325 AlPO_4 decreased to 26 % and instead, $\text{Ca}(\text{H}_2\text{PO}_4)_2$ was identified with an average of 25 %
326 which did not appear in the control and BC treatments. The LCF results showed that the soil
327 treated with BC^{plus} had no detectable Ca-hydroxyapatite which was found in the control and
328 BC treatments.

329

INSERT Table 4 HERE

331

332 The XANES spectra recorded from treated soil samples in the ryegrass pot experiment
333 showed the presence of an intense white-line in the energy range of 2152 to 2158 eV and a
334 stretched post-edge feature approximately from 2165 to 2178 eV (Fig. 4). Decreases in white-
335 line and post-edge intensities of the soil samples appeared as an effect of BC^{plus} application.
336 Visual inspection of P *K*-edge spectra revealed no indication of specific alteration in spectral
337 features in response to the BC treatment.

338

INSERT Figure 4 HERE

340

341 Amorphous AlPO_4 was identified by LCF analysis as dominant component ($\approx 35\%$) in all
342 treated soil samples from the ryegrass pot experiment (Table 5). The second major P form in

343 the control soil was IHP (29 %) followed by Ca-phytate (27 %), with the latter also as
344 pronounced as that observed for BC and BC^{plus} treatments. All treated soils varied in
345 proportions of free or bound IHP forms. The Mont-Al-PO₄ and Ca(H₂PO₄)₂ compounds were
346 only assigned in the control and BC^{plus} treatments, respectively.

347

348

INSERT Table 5 HERE

349

350 **4 Discussion**

351 **4.1 P availability as revealed by sequential fractionation**

352 The sequence of P distribution between sequentially extracted P-fractions was in accordance
353 with findings by many studies (Cross and Schlesinger, 1995; McDowell and Stewart, 2006;
354 Hashimoto and Watanabe, 2014), reflecting the general status of different P-pools in acidic
355 soils. The results indicated that the largest P proportion was found in the NaOH fraction
356 reflecting P fixed to Fe- and Al-oxides as followed by the NaHCO₃-P fraction assigned to
357 weakly P absorbed on crystalline Fe- and Al-oxides or surface of minerals. Guo et al. (2000)
358 reported that the NaOH-P fraction may support the labile NaHCO₃-P fraction as a buffering
359 P-pool in highly weathered and acidic soils. According to soil pH values (4.7 and 5.2), the
360 larger proportions of NaHCO₃-P even than HCl-P can be explained by the abundance and
361 surface loadings of Fe- and Al-oxides that support the electrostatic binding of phosphate ions
362 and a scarcity of Ca and Mg minerals or soluble ions. As expected, the lowest P proportions
363 were found in the mobile and readily available P fraction extracted by resin strips in
364 agreement to many comparable studies (Cross and Schlesinger, 1995; Bauchemin et al., 2003;
365 Sharpley et al., 2004; Siebers et al., 2013). Among the two soils which were used in the two
366 different experiments, the largest proportions of inorganic P were achieved in the soil after
367 incubation-leaching experiment, while the organic P forms were considerably more abundant

368 in the soil samples after ryegrass cultivation (Table 1). These differences may be due to the
369 microbial activities in the rhizosphere of grasses and transformation of P_i to more stable P_o
370 fractions during longer plant cultivation period (230 days) than in the non-cropped
371 incubation-leaching experiment (70 days).

372 In general, all P fraction concentrations were elevated by adding BC and BC^{plus} particles
373 which appeared to follow the same pattern in both soils under two different experimental
374 conditions. However, significant differences were found only between the control and BC^{plus}
375 treated soils (1-2 mm in the incubation-leaching experiment) for the resin-P and NaOH-P
376 fractions. Since the BCs particles were separated from the soils before chemical analysis, it
377 was expected that partly dissolved BCs would have a limited impact on different P-fractions
378 rather than totally ground and mixed BCs. This is consistent with the study of Siebers et al.
379 (2013) according to which the BC application ($<90 \mu\text{m}$ BC thoroughly mixed to soil)
380 significantly increased the insoluble P proportion ($\text{H}_2\text{SO}_4\text{-P}$). Additionally, our study
381 confirmed previous findings concerning the effect of particle sizes on the P release from BCs
382 (Morshedizad and Leinweber, 2017) and consequently the P status of treated soils (Ma and
383 Matsunaka, 2013). Sequentially extracted P contents increased with decreasing size of BC
384 particles whereby BC^{plus} treatments appeared more dependent on particle size than BC
385 treatments. [The results of sequential P fractionation of \$BC^{plus}\$ treatments in the incubation-](#)
386 [leaching experiment indicated that](#) the P increase was more pronounced for P fixed to Al- and
387 Fe-oxides (NaOH-P) than other fractions, whereas for BC treatments the largest increase
388 occurred in P bound to Ca and Mg minerals (HCl-P). It seems that local pH changes in soil
389 associated with BC and BC^{plus} amendments could eventually lead to a different distribution of
390 released P into differently soluble or insoluble P-pools which are generally controlled by pH
391 (Arai and Sparks, 2007). [However, due to lower fertilization level and longer period of](#)
392 [experiment in ryegrass cultivation compared to incubation-leaching, it appears that the](#)

393 chemical equilibrium has been established in the soil (no significant change in bulk soil pH)
394 and, accordingly, the soil P fractions were altered minimal.

395

396 **4.2 P speciation of BCs particles by XANES**

397 The predominance of Ca-hydroxyapatite in BCs as evidenced by P *K*-edge XANES analysis
398 is consistent with findings reported by previous studies (Warren et al., 2009; Siebers et al.,
399 2013). The mineral phase of bone consists mainly of hydroxyapatite, and its contribution to
400 bone and bone char compositions depends on species and the age of animals (Wu et al., 2003)
401 and carbonization temperature and residence time (Novotny et al., 2012). Bone crystallinity
402 might be improved through structural modifications on poorly crystalline fresh bone samples
403 (such as mineral maturity over periods of time or intensive carbonization) which can also
404 result in increased proportions of hydroxyapatite and accordingly a decrease in P solubility
405 (Novotny et al., 2012). Based on LC fittings, the second major component of BCs particles
406 was CaHPO₄, in good agreement with the results of Rajendran et al. (2013) who indicated the
407 heated bones at 400 °C contained some more soluble phosphates such as CaHPO₄ and
408 CaH₂PO₄ in addition to the hydroxyapatite fraction. The authors reported that spectra of
409 calcined bone samples at 700 °C had white-line at 2154 eV and two post-edge peaks at 2162
410 eV and 2169 eV with no pre-edge peaks and appeared similar to CaHPO₄ and CaH₂PO₄
411 spectra. Our LCF also assigned Ca-phytate in BCs samples which seems to be controversial
412 as a component of animal bone materials. The P *K*-edge spectrum of Ca-phytate is very
413 similar to other Ca-bound P compounds with a distinct white-line and lack of a pre-edge
414 feature, although it is likely distinguishable due to specific shape of white-line tailing and
415 absence of post-edge signal at 2164 eV (Prietz et al., 2016). Moreover, some inaccuracies in
416 LCF estimations have to be considered because of 1) uncertainty in speciation of organic P
417 forms by *K*-edge XANES, 2) lack of reference compounds representing all P forms in BCs

418 and 3) smaller Ca-phytate proportions than proposed 10 to 15 % of P_t as detection limit for
419 reliable XANES fittings (Beauchemin et al., 2003). Therefore, the P proportions assigned to
420 Ca-phytate also could originate from a range of other CaP compounds.

421 In both experiments, incubation-leaching and ryegrass cropping, changes in proportions of
422 Ca-hydroxyapatite and $CaHPO_4$ in BC particles followed an opposite trend than in BC^{plus}
423 particles. After placement of BC particles in the soil, Ca-phosphate seemed to be released
424 gradually over time which provides a locally lime-saturated condition. Due to elevated pH
425 surrounding the BC particles, dissolved P can be resorbed to maintain solubility and the Ca-P
426 equilibrium constant which likely resulted in a decreased proportion of soluble $CaHPO_4$ and
427 possibly the transformation into the relatively insoluble Ca-hydroxyapatite fraction. In
428 contrast, if BC^{plus} particles were applied to soils, larger proportions of $CaHPO_4$ at the expense
429 of Ca-hydroxyapatite could be explained by soil acidification through the microbial oxidation
430 of released S. This effect was more pronounced over the longer time period in the ryegrass
431 cropping pot experiment, favoring a greater $CaHPO_4$ than Ca-hydroxyapatite fraction. This
432 implies that BC^{plus} can actively supply P with predominance of soluble over insoluble P forms
433 in the long-term and, thus, meet crop requirements.

434

435 **4.3 P speciation of treated soils by XANES**

436 Differences between characteristics of two soils, dissimilar mechanisms of incubation-
437 leaching and plant uptake besides different experiment time durations complicate the joint
438 interpretation of the P-XANES data. In unfertilized soil of the incubation-leaching
439 experiment, the proportions of P species followed the order $AlPO_4 > FePO_4 > SOM-Al-IHP >$
440 Ca-hydroxyapatite which did not vary despite partial changes in some proportions after
441 application of both size fractions of BC particles. In general, these results concur with the
442 findings by Siebers et al. (2013) that Ca-hydroxyapatite proportion was slightly increased by

443 BC application. This could be attributed to irreversibly mixing finely-ground BC to the soil
444 samples whereas in the present experiments the BC particles were separated from the soils
445 before P speciation. Furthermore, these XANES data (Table 4 and Table 5) are in agreement
446 with sequential P fractionation results (Table 1) which indicated the dominance of inorganic
447 over organic P forms and showed the P fractions almost unchanged after BC application.
448 Implications of low solubility of BC particles observed in this work are consistent with
449 previous studies showing a slow release P from BCs (Warren et al., 2009; Siebers et al., 2013;
450 Morshedizad et al., 2016). Besides reducing the AlPO_4 and Ca-hydroxyapatite proportions,
451 BC^{plus} particles introduced highly soluble $\text{Ca}(\text{H}_2\text{PO}_4)_2$ to soils in the incubation-leaching
452 experiment. These results imply that considerable changes in P speciation were more
453 attributed to pH reductions and, accordingly, leaching out solubilized P-forms compared with
454 P enrichment by BC^{plus} dissolution. This is supported by results from a previous publication in
455 which two particle sizes of BC^{plus} gave a significant rise in the leached P-concentration after
456 1, 5, 13, 34 and 70 days of incubation along with reductions in soil pH (Morshedizad and
457 Leinweber, 2017). This is in line with Sato et al. (2005) who found that increasing soil pH in
458 a naturally acidic soil (pH = 4.32) was an effective approach to minimize P leaching, while
459 pH decrease resulted in transformation of stable to soluble and more leachable P species.
460 Regarding the XANES results of the ryegrass cultivation experiment (Table 5), the effect of
461 BC^{plus} treatment can be explained better. In the control soil, the presence of AlPO_4 and
462 increasing abundance of organic P-forms (Ca-phytate and IHP compounds) (Table 5) were
463 consistent with the appearance of NaOH-P and HCl-P fractions by sequential extraction
464 (Table 1). In the BC treatment the proportions of AlPO_4 and Ca-phytate did not change
465 compared to the control but the contribution of organic P increased by Mont-Al-IHP
466 formation. The stability of different P fractions can be favored by the pH effect (Gustafsson et
467 al., 2012) likewise the dependence of BC particles solubility on the soil pH (Siebers et al.,

468 2013). In agreement with incubation-leaching results (Table 4), $\text{Ca}(\text{H}_2\text{PO}_4)_2$ was detected as a
469 result of BC^{plus} amendment even though similar proportions of AlPO_4 and Ca-phytate were
470 observed between the control and BC^{plus} treatment. The results of sequential P fractionation
471 and XANES analyses [on treatments in the two different experiments](#) presented here
472 demonstrated that surface modification of BC particles effectively improved soluble P
473 fractions in BC^{plus} particles and, consequently, in amended soils.

474

475 **5 Conclusions**

476 In the present study, the P speciation by a sequential P fractionation and P K-edge XANES
477 spectroscopy revealed the noticeable alteration in the P-pools of treated soil samples. Results
478 of incubation-leaching and ryegrass cultivation experiments indicated that BC^{plus} produced by
479 surface-modification of BC [through addition of S compounds](#) provided more soluble and
480 plant-available P than non-modified BC during the growth season. The S oxidation and
481 thereby the soil pH decrease seems to stimulate the P release from BC^{plus} particles. The P K-
482 edge XANES analysis of BC^{plus} particles revealed more soluble CaHPO_4 than in BC particles
483 at the expense of Ca-hydroxyapatite. This was associated with $\text{Ca}(\text{H}_2\text{PO}_4)_2$ addition to
484 amended soils, as identified by sequential P fractionation and XANES analyses. Future
485 studies will be directed to validate the beneficial effects of BC^{plus} at the field scale with
486 different soils and to optimize the surface modification of BCs.

487

488 **Data availability.** All compiled data of this study are published in figures and tables.
489 Detailed primary data including the incubation-leaching and ryegrass cultivation results and
490 the acquired XANES spectra will be saved and published in the BonaRes Data Center
491 (<https://www.bonares.de/research-data>) and get the BonaRes DOI prefix (10.20387).

492

493 **Competing interests.** The authors declare that they have no conflict of interest.

494

495 **Acknowledgments.** We are grateful to the technician team and all staff at the Beamline 8
496 of SLRI for their help and assistance associated to XANES spectra acquisitions. The authors
497 are appreciative to Prof. J. Prietzel (Department of Soil Science, Technical University of
498 Munich) for providing the P reference spectra and invaluable comments that improved our
499 research. M. Morshedizad acknowledges a Ph.D. grant from the Federal State of
500 Mecklenburg, Western Pommerania in Germany and travel expenses from the Leibniz
501 Science Campus Phosphorus Research Rostock. Parts of this work were funded by the
502 German Federal Ministry of Education and Research (BMBF) through the BonaRes-Project
503 InnoSoilPhos (No. 031A558).

504

505 **References**

506 Ahlgren, J., Djodjic, F., Börjesson, G., and Mattsson, L.: Identification and quantification of
507 organic phosphorus forms in soils from fertility experiments, *Soil Use Manag.*, 29,
508 24–35, doi:10.1111/sum.12014, 2013.

509 Arai, Y. and Sparks, D. L.: Phosphate reaction dynamics in soil and soil components: a
510 multiscale approach, *Adv. Agron.*, 94, 135–179, doi:10.1016/S0065-2113(06)94003-6,
511 2007.

512 Beauchemin, S., Hesterberg, D., Chou, J., Beauchemin, M., Simard, R. R., and Sayers, D.E.:
513 Speciation of phosphorus in phosphorus-enriched agricultural soils using X-ray
514 absorption near-edge structure spectroscopy and chemical fractionation, *J. Environ.*
515 *Qual.*, 32, 1809–1819, doi:10.2134/jeq2003.1809, 2003.

516 Condron, L. M. and Newman, S.: Revisiting the fundamentals of phosphorus fractionation of
517 sediments and soils, *J. Soil Sediment*, 11, 830–840, doi:10.1007/s11368-011-0363-2,
518 2011.

519 Cross, A. F. and Schlesinger, W. H.: A literature-review and evaluation of the Hedley
520 fractionation - applications to the biochemical cycle of soil-phosphorus in natural
521 ecosystems, *Geoderma*, 64, 197–214, doi:10.1016/0016-7061(94)00023-4, 1995.

522 De Brabandere, H., Forsgard, N., Israelsson, L., Petterson, J., Rydin, E., Waldebäk, M., and
523 Sjöerg, P. J.: Screening for organic phosphorus compounds in aquatic sediments by
524 liquid chromatography coupled to ICP-AES and ESI-MS/MS, *Anal. Chem.*, 80, 6689–
525 6697, doi:10.1021/ac8006335, 2008.

526 Delgado, A., Madrid, A., Kassem, S., Andreu, L., and Campillo, M. C.: Phosphorus fertilizer
527 recovery from calcareous soils amended with humic and fulvic acids, *Plant Soil*, 245,
528 277–286, doi:10.1023/A:1020445710584, 2002.

529 Dieter, D., Elsenbeer, H., and Turner, B. L.: Phosphorus fractionation in lowland tropical
530 rainforest soils in central Panama, *Catena*, 82, 118–125,
531 doi:10.1016/j.catena.2010.05.010, 2010.

532 [Fan, X., Habib, L., Fleckenstein, J., Haneklaus, S., and Schnug, E.: "In situ digestion" a](#)
533 [concept to manage soil phosphate in organic farming, In: Proc. of the 13th Intern.](#)
534 [Fertilizer Symposium, Tokat, Turkey, 10-13 June 2002. 219–228, 2003.](#)

535 Guo, F., Yost, R. S., Hue, N. V., Evensen, C. I., and Silva, J. A.: Changes in phosphorus
536 fractions in soils under intensive plant growth, *Soil Sci. Soc. Am. J.*, 64, 1681–1689,
537 doi:10.2136/sssaj2000.6451681x, 2000.

538 Gustafsson, J. P., Mwamila, L. B., and Kergoat, K.: The pH dependence of phosphate
539 sorption and desorption in Swedish agricultural soils, *Geoderma*, 189–190, 304–311,
540 doi:10.1016/j.geoderma.2012.05.014, 2012.

541 Hartley, T. N., Macdonald, A. J., McGrath, S. P., and Zhao, F. J.: Historical arsenic
542 contamination of soil due to long-term phosphate fertiliser applications, *Environ.*
543 *Pollut.*, 180, 259–264, doi:10.1016/j.envpol.2013.05.034, 2013.

544 Hashimoto, Y. and Watanabe, Y.: Combined applications of chemical fractionation, solution
545 ³¹P-NMR and P K-edge XANES to determine phosphorus speciation in soils formed
546 on serpentine landscapes, *Geoderma*, 230, 143–150, doi:
547 10.1016/j.geoderma.2014.04.001, 2014.

548 [He, Q. B., and Singh, B. R.: Plant availability of cadmium in soils: I. Extractable cadmium in](#)
549 [newly and long-term cultivated soils, *Acta. Agric. Scand. Sect. B*, 43, 134–141,](#)
550 [doi:10.1080/09064719309411231, 1993.](#)

551 Hedley, M. J., Stewart, J. W. B., and Chauhan, B. S.: Changes in organic and organic soil
552 phosphorus fractions induced by cultivation practices and laboratory incubations, *Soil*
553 *Sci. Soc. Am. J.*, 46, 970–976, doi:10.2136/sssaj1982.03615995004600050017x,
554 1982.

555 Herzel, H., Krüger, O., Hermann, L., and Adam, C.: Sewage sludge ash - A promising
556 secondary phosphorus source for fertilizer production, *Sci. Total Environ.*, 542, 1136–
557 1143, doi:10.1016/j.scitotenv.2015.08.059, 2016.

558 Kaur, G. and Reddy, M. S.: Role of phosphate-solubilizing bacteria in improving the soil
559 fertility and crop productivity in organic farming, *Arch. Agron. Soil Sci.*, 60, 549–564,
560 doi:10.1080/03650340.2013.817667, 2014.

561 Kelly, S., Hesterberg, D., and Ravel, B.: Analysis of soils and minerals using x-ray absorption
562 spectroscopy, p. 387–463. In A.L. Ulery and R. Drees (ed.) *Methods of soil analysis.*
563 *Part 5. Mineralogical methods*, SSSA, Madison, WI., 2008.

564 Kizewski, F., Liu, Y. T., Morris, A., and Hesterberg, D.: Spectroscopic approaches for
565 phosphorus speciation in soils and other environmental systems, *J. Environ. Qual.*, 40,
566 751–766, doi:10.2134/jeq2010.0169, 2011.

567 Klysubun, W., Sombunchoo, P., Deenan, W., and Kongmark, C.: Performance and status of
568 beamline BL8 at SLRI for X-ray absorption spectroscopy, *J. Synchrotron Rad.*, 19,
569 930–936, doi:10.1107/S0909049512040381, 2012.

570 Kratz, S., Schick, J., and Schnug E.: Trace elements in rock phosphates and P containing
571 mineral and organo-mineral fertilizers sold in Germany, *Sci. Total Environ.*, 542,
572 1013–1019, doi:10.1016/j.scitotenv.2015.08.046, 2016.

573 Kruse, J., Abraham, M., Amelung, W., Baum, C., Bol, R., Kuhn, O., Lewandowski, H.,
574 Niederberger, J., Oelmann, Y., Ruger, C., Santner, J., Siebers, M., Siebers, N., Spohn,
575 M., Vestergren, J., Vogts, A., and Leinweber, P.: Innovative methods in soil
576 phosphorus research: a review, *J. Plant Nutr. Soil Sci.*, 178, 43–88,
577 doi:10.1002/jpln.201400327, 2015.

578 Lanfranco, A. M., Schofield, P. F., Murphy, P. J., Hodson, M. E., Mosselmans, J. F. W., and
579 Valsami-Jones, E.: Characterization and identification of mixed-metal phosphates in
580 soils: the application of Raman spectroscopy, *Mineral. Mag.*, 67, 1299–1316,
581 doi:10.1180/0026461036760166, 2003.

582 Liu, J. Y., Wang, H., Yang, H. J., Ma, Y. J., and Cai, O. C.: Detection of phosphorus species
583 in sediments of artificial landscape lakes in China by fractionation and phosphorus-31
584 nuclear magnetic resonance spectroscopy, *Environ. Pollut.*, 157, 49–56,
585 doi:10.1016/j.envpol.2008.07.031, 2009.

586 Ma, Y. L. and Matsunaka, T.: Biochar derived from dairy cattle carcasses as an alternative
587 source of phosphorus and amendment for soil acidity, *Soil Sci. Plant Nutr.*, 59,
588 628–641, doi:10.1080/00380768.2013.806205, 2013.

589 McDowell, R. W. and Stewart, I.: The phosphorus composition of contrasting soils in
590 pastoral, native and forest management in Otago, New Zealand: Sequential extraction
591 and ^{31}P NMR, *Geoderma*, 130, 176–189, doi:10.1016/j.geoderma.2005.01.020, 2006.

592 Morshedizad, M. and Leinweber, P.: Leaching of phosphorus and cadmium in soils amended
593 with different bone chars, *CLEAN – Soil, Air, Water*, 45 (8),
594 doi:10.1002/clen.201600635, 2017.

595 Morshedizad, M., Zimmer, D., and Leinweber, P.: Effect of bone chars on phosphorus-
596 cadmium-interactions as evaluated by three extraction procedures, *J. Plant Nutr. Soil*
597 *Sci.*, 179, 388–398, doi:10.1002/jpln.201500604, 2016.

598 Murphy, J. and Riley, J. P.: A modified single solution method for the determination of
599 phosphate in natural waters, *Anal. Chim. Acta*, 27, 31–36, doi:10.1016/S0003-
600 2670(00)88444-5, 1962.

601 Novotny, E. H., Auccaise, R., Velloso, M. H. R., Corrêa, J. C., Higarashi, M. M., Abreu, V.
602 M. N., Rocha, J. D., and Kwapinski, W.: Characterization of phosphate structures in
603 biochar from swine bones, *Pesq. Agropec. Bras.*, 47, 672–676, doi:10.1590/S0100-
604 204X2012000500006, 2012.

605 Paraskova, J. V., Jorgensen, C., Reitzel, K. Pettersson, J., Rydin, E., and Sjoberg, P. J. R.:
606 Speciation of inositol phosphates in lake sediments by ion-exchange chromatography
607 coupled with mass spectrometry, inductively coupled plasma atomic emission
608 spectroscopy, and ^{31}P NMR spectroscopy, *Anal. Chem.*, 87, 2672–2677,
609 doi:10.1021/ac5033484, 2015.

610 Prietzel, J., Dümig, A., Wu, Y., Zhou, J., and Klysubun, W.: Synchrotron-based P K-edge
611 XANES spectroscopy reveals rapid changes of phosphorus speciation in the topsoil of
612 two glacier foreland chronosequences, *Geochim. Cosmochim. Acta*, 108, 154–171,
613 doi:10.1016/j.gca.2013.01.029, 2013.

614 Prietzel, J., Harrington, G., Häusler, W., Heister, K., Werner, F., and Klysubun, W.:
615 Reference spectra of important adsorbed organic and inorganic phosphate binding
616 forms for soil P speciation using synchrotron-based *K*-edge XANES spectroscopy, *J.*
617 *Synchrotron Rad.*, 23, 532–544, doi:10.1107/S1600577515023085, 2016.

618 Rajendran, J., Gialanella, S., and Aswath, P. B.: XANES analysis of dried and calcined bones,
619 *Mater. Sci. Eng. C*, 33, 3968–3979, doi:10.1016/j.msec.2013.05.038, 2013.

620 Ravel, B. and Newville, M.: Athena, Artemis, Hephaestus: data analysis for X-ray absorption
621 spectroscopy using IFEFFIT, *J. Synchrotron Rad.*, 12, 537–541,
622 doi:10.1107/S0909049505012719, 2005.

623 Rubaek, G. H., Kristensen, K., Olesen, S. E., Ostergaard, H. S., and Heckrath, G.: Phosphorus
624 accumulation and spatial distribution in agricultural soils in Denmark, *Geoderma*, 209,
625 241–250, doi:10.1016/j.geoderma.2013.06.022, 2013.

626 Sato, S., Solomon, D., Hyland, C., Ketterings, Q. M., and Lehmann, J.: Phosphorus speciation
627 in manure and manure-amended soils using XANES spectroscopy, *Environ. Sci.*
628 *Technol.*, 39, 7485–7491, doi:10.1021/es0503130, 2005.

629 [Schnug, E., Haneklaus, S., Rogasik, J., and Gassner, A.: Utilisation of fertiliser P with special
630 regard to organic farming, In: Proc. of the 14th Intern. Symposium of Fertilizers,
631 Debrecen, Hungary, 22-25 June 2003. 11–29, 2003.](#)

632 Scholz, R. W., Ulrich, A. E., Eilittä, M., and Roy, A.: Sustainable use of phosphorus: a finite
633 resource, *Sci. Total Environ.*, 461–462, 799–803, doi:10.1016/j.scitotenv.2013.05.043,
634 2013.

635 Schröder, J. J., Smit, A. L., Cordell, D., and Rosemarin, A.: Improved phosphorus use
636 efficiency in agriculture: a key requirement for its sustainable use, *Chemosphere*, 84,
637 822–831, doi:10.1016/j.chemosphere.2011.01.065, 2011.

638 Sharpley, A. N., McDowell, R. W., and Kleinman, P. J. A.: Amounts, forms and solubility of
639 phosphorus in soils receiving manure, *Soil Sci. Soc. Am. J.*, 68, 2048–2054,
640 doi:10.2136/sssaj2004.2048, 2004.

641 Shen, J. B., Yuan L. X., Zhang J. L., Li H. G., Bai Z. H., Chen, X. P., Zhang, W. F., and
642 Zhang, F. S.: Phosphorus dynamics: from soil to plant, *Plant Physiol.*, 156, 997–1005,
643 doi:10.1104/pp.111.175232, 2011.

644 Siebers, N. and Leinweber, P.: Bone Char - A clean and renewable fertilizer with cadmium
645 immobilizing capacity, *J. Environ. Qual.*, 42, 405–411, doi:10.2134/jeq2012.0363,
646 2013.

647 Siebers, N., Kruse, J., and Leinweber, P.: Speciation of phosphorus and cadmium in a
648 contaminated soil amended with bone char: sequential fractionations and XANES
649 spectroscopy, *Water Air Soil Pollut.*, 224, 1564–1577, doi:10.1007/s11270-013-1564-
650 7, 2013.

651 Templeton, D. M., Ariese, F., Cornelis, R., Danielsson, L. G., Muntau, H., Van Leewen, H.
652 P., and Lobinski, R.: Guidelines for the terms related to chemical speciation and
653 fractionation of elements. Definitions, structural aspects, and methodological
654 approaches (IUPAC Recommendations 2000), *Pure Appl. Chem.*, 72, 1453–1470,
655 doi:10.1351/pac200072081453, 2000.

656 Toor, G. S., Hunger, S., Peak, J. D., Sims, J. T., and Sparks, D. L.: Advances in the
657 characterization of phosphorus in organic wastes: Environmental and agronomic
658 applications, *Adv. Agron.*, 89, 1–72, doi:10.1016/S0065-2113(05)89001-7, 2006.

659 [USEPA. Method 3051a: Microwave assisted acid dissolution of sediments, sludges, soils, and](#)
660 [oils, 2nd ed. U.S. Gov. Print. Office, Washington, DC, 1997.](#)

661 Van Vuuren, D. P., Bouwman, A. F., and Beusen, A. H. W.: Phosphorus demand for the
662 1970–2100 period: A scenario analysis of resource depletion, *Globe Environ. Change*,
663 20, 428–439, doi:10.1016/j.gloenvcha.2010.04.004, 2010.

664 Vestergren, J., Vincent, A. G., Jansson, M., Persson, P., Ilstedt, I., Gröbner, G., Giesler, R.,
665 and Schleucher, J.: High-resolution characterization of organic phosphorus in soil
666 extracts using 2D ^1H - ^{31}P NMR correlation spectroscopy, *Environ. Sci. Technol.*, 46,
667 3950–3956, doi:10.1021/es204016h, 2012.

668 Vogel, C., Adam, C., Sekine, R., Schiller, T., Lipiec, E., and McNaughton, D.: Determination
669 of phosphorus fertilizer soil reactions by Raman and synchrotron infrared
670 microspectroscopy, *Appl. Spectrosc.*, 67, 1165–1170, doi:10.1366/13-07056, 2013.

671 Warren, G. P., Robinson, J. S., and Someus, E.: Dissolution of phosphorus from animal bone
672 char in 12 soils, *Nutr. Cycling Agroecosyst.*, 84, 167–178, doi:10.1007/s10705-008-
673 9235-6, 2009.

674 Werner, F. and Prietzel, J.: Standard protocol and quality assessment of soil phosphorus
675 speciation by P *K*-edge XANES spectroscopy, *Environ. Sci. Technol.*, 49, 10521–
676 10528, doi:10.1021/acs.est.5b03096, 2015.

677 Wu, Y., Ackerman, J. L., Strawich, E. S., Rey, C., Kim, H-M., and Glimcher, M. J.: Phosphate
678 ions in bone: identification of a calcium–organic phosphate complex by ^{31}P solid-state
679 NMR spectroscopy at early stages of mineralization, *Calcif. Tissue Int.*, 72, 610–26,
680 doi:10.1007/s00223-002-1068-8, 2003.

681

682

683

684

685

686 **Table 1.** Distribution of inorganic P (P_i), organic P (P_o) and total P (P_t) concentrations (mg P kg^{-1} soil)
687 of sequentially extracted P-fractions in the soils as affected by different treatments (treated with two
688 particle size fractions (1-2 and 2-4 mm) and original sizes of BC and BC^{plus} or unfertilized soils
689 (control) after incubation-leaching and ryegrass cultivation experiments.

Treatment	Resin-P			NaHCO ₃ -P			NaOH-P			HCl-P		
	P _i	P _o	P _t	P _i	P _o	P _t	P _i	P _o	P _t	P _i	P _o	P _t
Incubation-leaching												
Control	47	5	52	160	99	259	565	294	859	113	10	123
BC ₁₋₂	56 ^{NS}	7 ^{NS}	63 ^{NS}	163 ^{NS}	108 ^{NS}	271 ^{NS}	578 ^{NS}	303 ^{NS}	881 ^{NS}	140 ^{NS}	56 ^{NS}	196 ^{NS}
BC ₂₋₄	50 ^{NS}	7 ^{NS}	57 ^{NS}	161 ^{NS}	105 ^{NS}	266 ^{NS}	574 ^{NS}	301 ^{NS}	875 ^{NS}	121 ^{NS}	40 ^{NS}	161 ^{NS}
BC ^{plus} ₁₋₂	61 [*]	7 ^{NS}	68 ^{NS}	172 ^{NS}	111 ^{NS}	283 ^{NS}	593 [*]	313 ^{NS}	906 [*]	131 ^{NS}	37 ^{NS}	170 ^{NS}
BC ^{plus} ₂₋₄	50 ^{NS}	7 ^{NS}	57 ^{NS}	160 ^{NS}	104 ^{NS}	264 ^{NS}	574 ^{NS}	298 ^{NS}	872 ^{NS}	115 ^{NS}	21 ^{NS}	135 ^{NS}
Ryegrass cropping experiment												
Control	4	4	8	25	27	52	75	121	196	28	5	33
BC	2 ^{NS}	5 ^{NS}	7 ^{NS}	24 ^{NS}	32 ^{NS}	56 ^{NS}	79 ^{NS}	125 ^{NS}	204 ^{NS}	30 ^{NS}	6 ^{NS}	36 ^{NS}
BC ^{plus}	6 [*]	5 ^{NS}	11 [*]	35 ^{NS}	27 ^{NS}	62 ^{NS}	85 ^{**}	128 ^{NS}	213 ^{NS}	34 ^{NS}	7 ^{NS}	41 ^{NS}

690
691 * Significant at $P < 0.05$; ** Significant at $P < 0.01$; ^{NS} Non-significant difference (treatment vs. control; Tukey-Test)
692

693

694

695

696

697

698

699

700

701

702

703

704

705 **Table 2.** Results of linear combination fitting (LCF) conducted on P *K*-edge XANES spectra of bone
 706 char (BC) and surface modified bone char (BC^{plus}) particles before and after a 70-days incubation-
 707 leaching period.

Reference compound	Before experiment				After 70 days incubation-leaching			
	BC		BC ^{plus}		BC		BC ^{plus}	
	1-2 mm	2-4 mm	1-2 mm	2-4 mm	1-2 mm	2-4 mm	1-2 mm	2-4 mm
Ca-hydroxyapatite (%)	58± 6	64±5	62± 5	58± 5	75± 4	85± 3	59± 5	60± 6
CaHPO ₄ (%)	24± 5	19± 4	28± 4	32± 6	14± 3	5± 2	33± 4	35± 5
Ca-phytate (%)	18± 4	17± 4	10± 3	10± 3	11± 3	10± 2	8± 4	5± 4
<i>R</i> -factor	0.012	0.008	0.007	0.009	0.005	0.002	0.009	0.010

708
709
710
711
712
713
714
715
716
717
718
719
720
721
722
723
724
725
726

727 **Table 3.** Results of linear combination fitting (LCF) conducted on P *K*-edge XANES spectra of bone
 728 char (BC) and surface modified bone char (BC^{plus}) particles before and after 230-days of ryegrass
 729 cultivation in a pot experiment.

Reference compound	Before experiment		After 230 days ryegrass cultivation	
	BC	BC ^{plus}	BC	BC ^{plus}
Ca-hydroxyapatite (%)	63± 6	70± 4	75± 4	49± 8
CaHPO ₄ (%)	29± 5	29± 3	17± 4	43± 6
Ca-phytate (%)	8± 4	1± 3	8± 3	8± 5
<i>R</i> -factor	0.012	0.005	0.006	0.018

730

731

732

733

734

735

736

737

738

739

740

741

742

743

744

745

746

747

748

749 **Table 4.** Results of linear combination fitting (LCF) conducted on P *K*-edge XANES spectra of
 750 unfertilized (control) and fertilized soils with bone char (BC) and surface modified bone char (BC^{plus})
 751 particles in the 70-days incubation-leaching experiment.

752

Reference compound	Control	BC treatment		BC ^{plus} treatment	
		1-2 mm	2-4 mm	1-2 mm	2-4 mm
Ca-hydroxyapatite (%)	8± 1	4± 1	8± 1	0	0
AlPO ₄ amorphous (%)	42± 1	42± 2	40± 1	27± 1	24± 1
FePO ₄ (%)	29± 1	31± 2	31± 1	27± 1	26± 1
SOM-Al-IHP (%)	21± 2	23± 4	21± 3	26± 2	21± 1
Ca(H ₂ PO ₄) ₂ (%)	0	0	0	20± 1	29± 1
<i>R</i> -factor	0.0003	0.0007	0.0003	0.0005	0.0004

758

759

760

761

762

763

764

765

766

767

768

769

770

771

772

773

774

775 **Table 5.** Results of linear combination fitting (LCF) conducted on P *K*-edge XANES spectra of
 776 unfertilized (control) and fertilized soils with bone char (BC) and surface modified bone char (BC^{plus})
 777 particles under 230-days of ryegrass cultivation in a pot experiment.

778	Reference compound	Control	BC treatment	BC ^{plus} treatment
779	AlPO ₄ amorphous (%)	35± 3	35± 3	34± 1
780	Ca-phytate (%)	27± 3	28± 3	27± 1
781	IHP (%)	29± 5	21± 7	0
782	Mont-Al-IHP (%)	0	16± 1	0
783	SOM-Al-IHP (%)	0	0	25± 2
784	Mont-Al-PO ₄ (%)	9± 1	0	0
785	Ca(H ₂ PO ₄) ₂ (%)	0	0	14± 2
786	<i>R</i> -factor	0.0006	0.0008	0.0006

795
796
797
798

799

800

801

802

803

804

805

806

807

808

809

810

811

812

813 **Figure captions**

814 **Figure 1.** Normalized P *K*-edge XANES spectra of different BC and BC^{plus} particle sizes (1-2 and 2-4
815 mm) before (control) and after 70-days incubation-leaching experiment compared to the reference
816 compounds selected using LCF method.

817

818 **Figure 2.** P *K*-edge XANES spectra of BC and BC^{plus} particles before (control) and after 230-days of
819 ryegrass cultivation compared to the reference compounds selected by LCF method.

820

821 **Figure 3.** P *K*-edge XANES spectra of unfertilized (control) and fertilized soils with BC and BC^{plus}
822 particles under 70-days incubation-leaching experiment compared to the reference compounds
823 selected by LCF method.

824

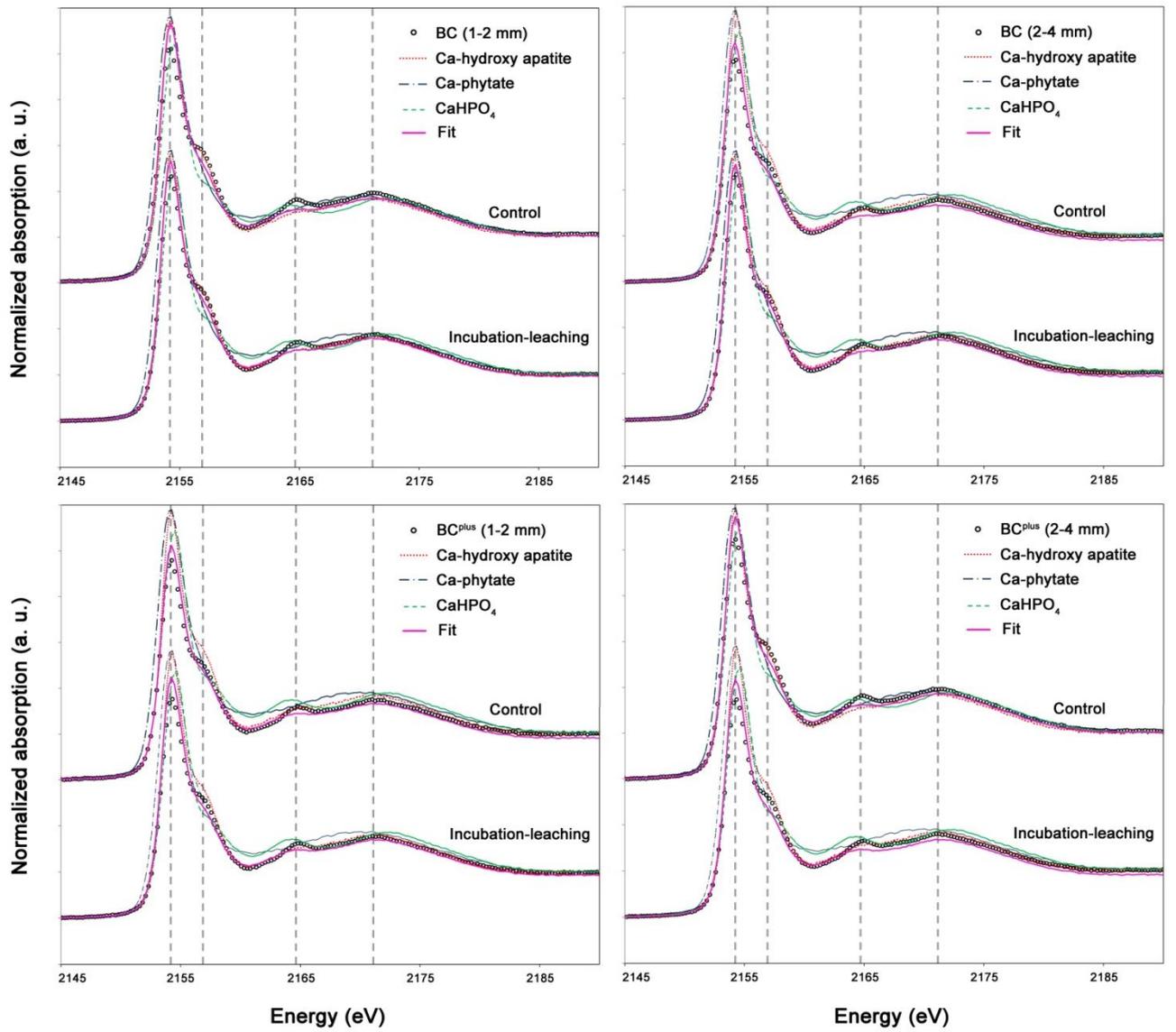
825 **Figure 4.** P *K*-edge XANES spectra of unfertilized (control) and fertilized soils with BC and BC^{plus}
826 particles under 230-days of ryegrass cultivation compared to the reference compounds selected by
827 LCF method.

828

829

830

831



832

833 Morshedizad et al., Figure 1

834

835

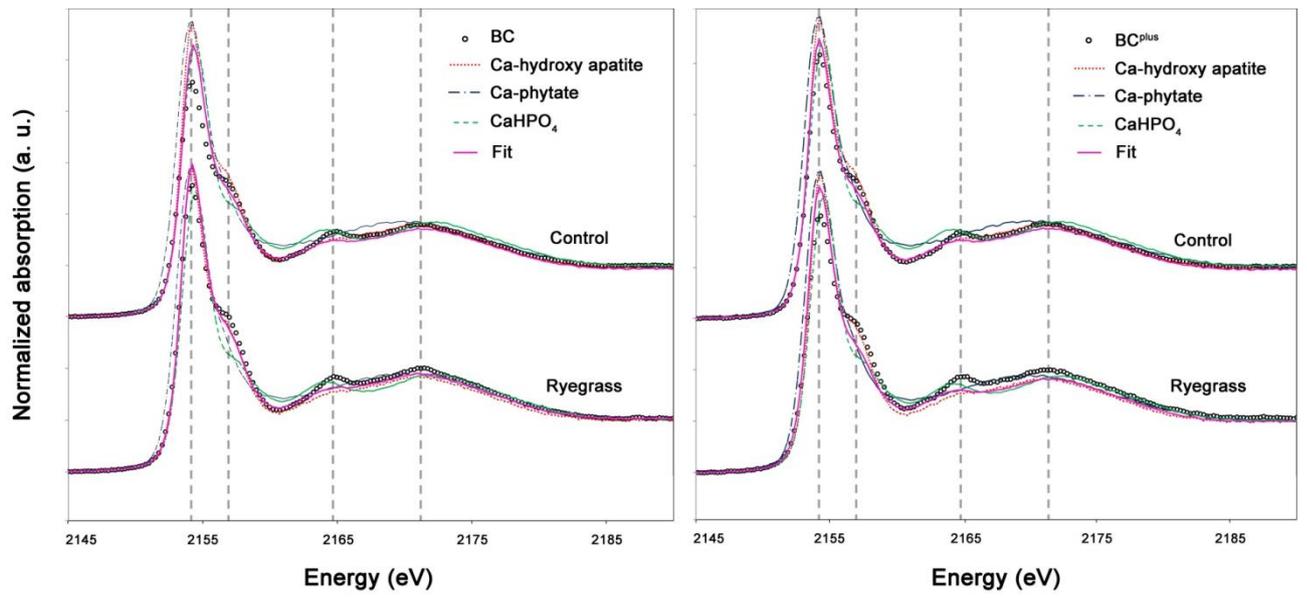
836

837

838

839

840



841

842 Morshedizad et al., Figure 2

843

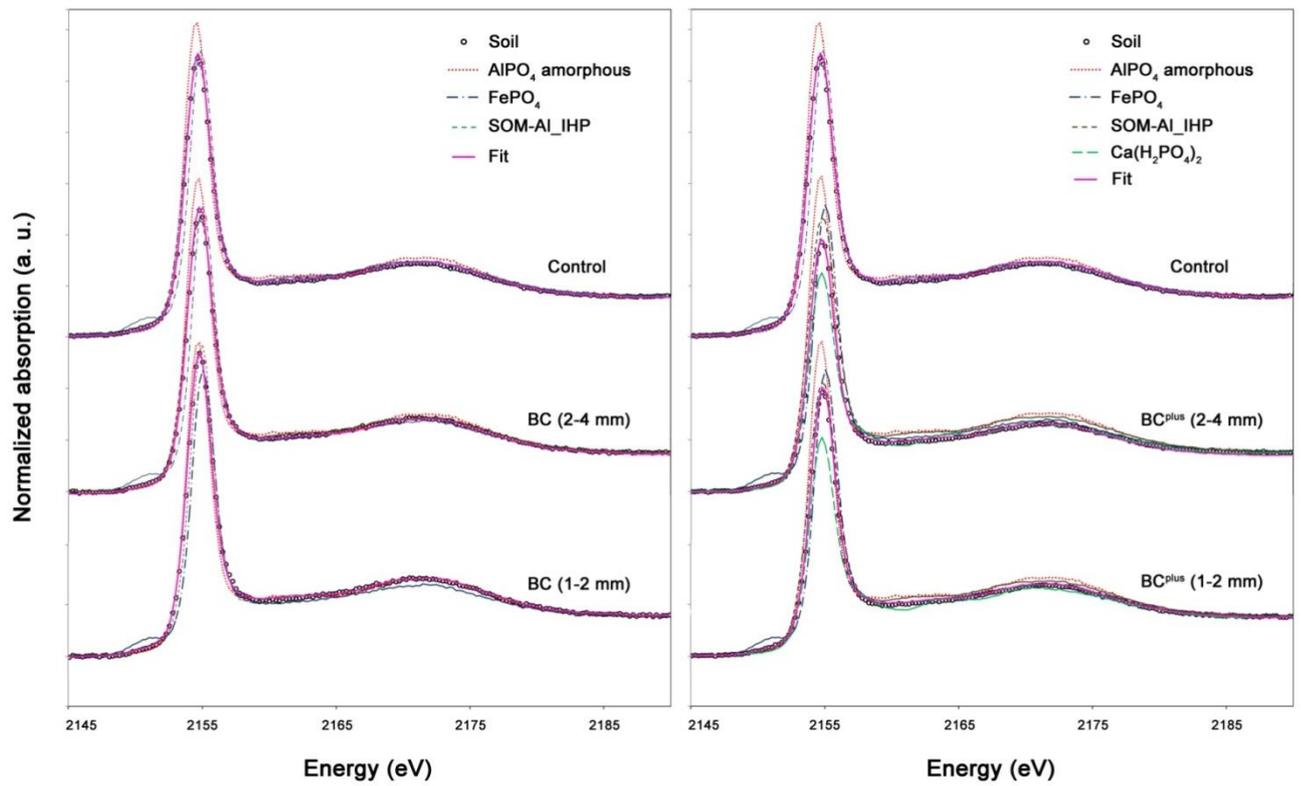
844

845

846

847

848



849

850 Morshedizad et al., Figure 3

851

852

853

854

855

856

857

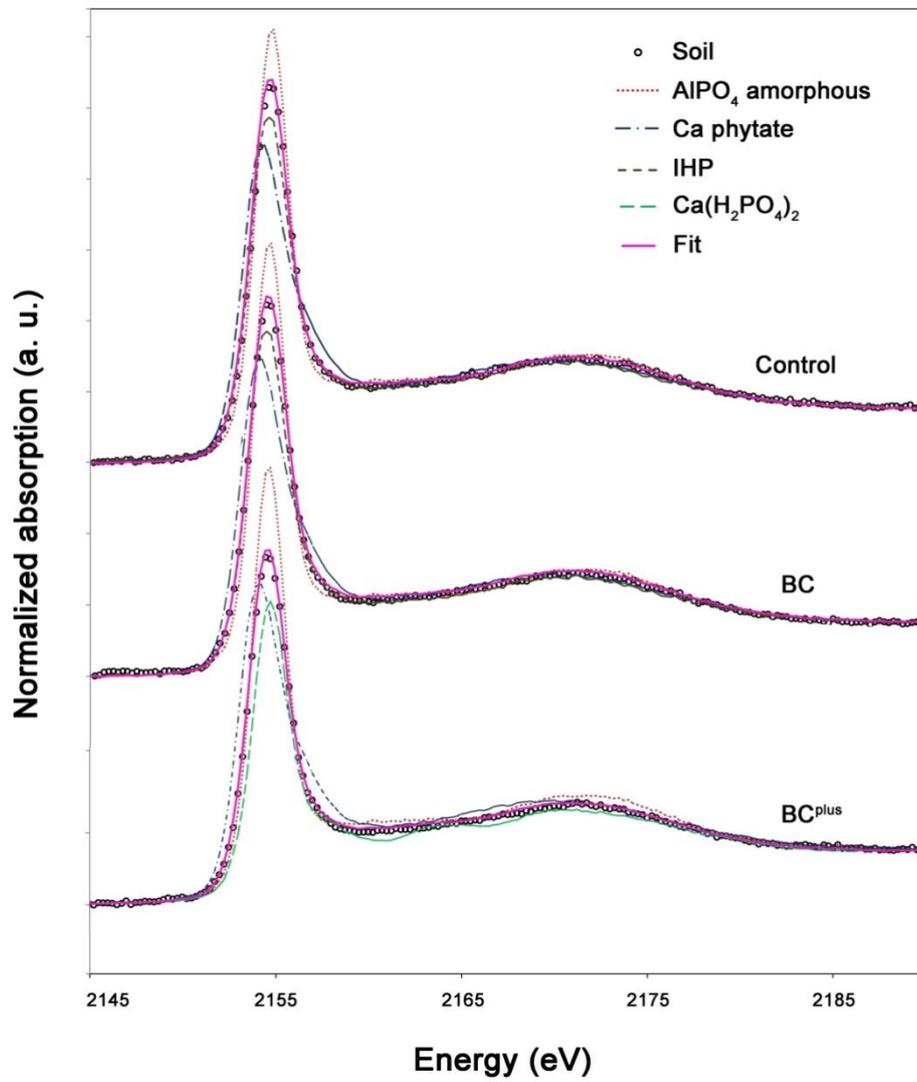
858

859

860

861

862



863

864 Morshedizad et al., Figure 4

865

866

867

868

869

870

871

872