

1 **Dear Dr. Rumpel, we thank you for the effort that has gone into evaluating our article.**
2 **Your suggestions greatly helped to improve the quality of the manuscript. Please find**
3 **below our replies to the individual comments.**

4

- 5 • The manuscript should be carefully proofread, e.g. in the first sentence it should be
6 '...determine many crucial soil properties and the cycling of carbon' ; l.26
7 'derived' should be 'derivated' ; P.2, l. 4 : 'understanding of SOM formation' ; P.6, l.
8 27 : 'are relatively small components' should be replaced by 'contributing little to', P.
9 8, l.7 : 'aromatic matter' should be replaced by 'aromatic OM compounds' ; P.12, l.11
10 : 'on' should be deleted.

11

12 **Answer: We proofread the manuscript and corrected these sentences.**

13

- 14 • In the two sections giving the arguments for contradictory theories for stabilisation of
15 aromatic C, authors should discuss all literature in terms of positive arguments for
16 these research lines.

17

18 **Answer: At first, we planned to write a comprehensive literature review on the topic,**
19 **but then decided to write the manuscript in a more condensed manner, because for**
20 **many of the individual arguments that we discuss, review articles have been published**
21 **during the last years.**

22 **For instance, Kleber et al. (2015) recently presented a comprehensive literature review**
23 **and discussion on importance of sorption of DOM to mineral surfaces for the long-term**
24 **preservation of organic matter in soil. Thevenot et al. (2010) summed up the large body**
25 **of literature (about 150 articles) on distribution of lignin phenols in mineral soils. In the**
26 **revised manuscript, we now clearly indicate the cases, in which we refer to data and**
27 **conclusions discussed in review articles.**

28 **In other cases, however, when no recent comprehensive review articles are available, we**
29 **now present a list of relevant studies supporting the argumentation. This resulted in the**
30 **addition of about 40 references to the manuscript.**

31

- 32 • In the 'Dissolved phase line of evidence', the third paragraph on the importance of
33 root origin of dissolved aromatics should be the third argument. In this paragraph it

1 should be stated that all these arguments were obtained for acid forest soils under
2 temperate climate or through laboratory analyses. Therefore, this line of evidence is
3 difficult to use as a general argument.
4

5 **Answer: Done. We now present the paragraph as the third argument supporting the**
6 **`dissolved phase line of evidence`. In addition, we outline that the currently available**
7 **data on release of aromatic DOM components is limited to laboratory experiments and**
8 **refers to root decomposition in acid temperate forest soils. We agree, knowledge about**
9 **how root decomposition contributes to DOM in mineral soil is currently very limited.**

- 10
- 11 • In the second section 'solid line of evidence', the contradictory argument mentioned in
12 the last sentence (P.7, l.30-32) should be deleted, as it introduces a confusion to the
13 reader.
14

15 **Answer: Done.**

- 16
- 17 • P. 8, l.29: the last sentence should be deleted, as it is a general statement. Instead it
18 could be good to add a transition sentence introducing NMR as a method to quantify
19 aromatic compounds.
20

21 **Answer: Done. The sentence ` Our knowledge about how much lignin is `hidden`**
22 **(Hernes et al., 2013) in soil is still insufficient` has been deleted. In addition, we revised**
23 **the first sentence of the paragraph on NMR. It now reads: ` Solid-state CPMAS ¹³C-**
24 **NMR has been widely used in the last decades to study the composition of SOM.**
25 **Whether the results are quantitative has been subject of an intensive debate`.**

- 26
- 27 • P.9 : Concerning the analytical bias introduced by HF treatment, the study by
28 Eusterhues et al. (2007) , OG should be considered. It shows that the composition of
29 HF soluble C is different for different soil types – thus not providing evidence of
30 selective loss of specific C species.
31

32 **Answer: We discuss the article of Eusterhues in the revised manuscript. In our opinion,**
33 **the comparison of NMR spectra from untreated and HF-treated soil comes with**
34 **uncertainties. In the untreated soil, part of the SOM attached to mineral surfaces might**

1 have been invisible due to the proximity to paramagnetic material, while in the HF-
2 treated soil, the same SOM might have become removed during the treatment, thus, was
3 no more detectable. Moreover, the data presented in Eusterhues et al. (2007) suggest
4 that not all of the minerals were removed by the HF-treatment, presumably as the
5 minerals were protected by OM coatings. In topsoil, only 20-30% of the mineral-
6 associated OC was removed. Hence, it was not possible to gain any information about
7 the composition of the other 70-80%.

8

- 9 • P.9 : the Bloch decay technique may lead in some cases to underestimation of O-alkyl
10 C, whereas the work of Knicker shows nicely that aromatic C, if not very condensed
11 can be detected by CPMAS.

12

13 **Answer:** We agree that aromatic C can be detected in soil with the CPMAS method.
14 However, it is likely that its contribution is underestimated by the common approach of
15 integrating and comparing peak areas of different regions of the spectra. Evidence for
16 this can be found in literature. We added a more comprehensive discussion on this issue
17 to the revised manuscript. It reads as follows: “The common approach used to quantify
18 the relative contribution of different C types to SOM is to integrate and compare peak
19 areas of different spectra regions without considering any non-proportional signal
20 responses (e.g., Kögel-Knabner 2002). It has been shown that this approach
21 underestimates lignin vs. cellulose in ligno-cellulose isolated from wheat (Gauthier et al.,
22 2002). Also, peaks of methoxyl C often tend to be larger than those of aromatic C. Both,
23 methoxyl C and aromatic C derive from lignin, but lignin has much less methoxyl C
24 than aromatic C. Spectra of wood samples (e.g., Bonanomi et al. 2014), needle litter
25 (Preston and Trofymow, 2015) and grass litter (McKee et al., 2016) all show that
26 patterns of methoxyl signals being larger than those of aromatic and phenolic C
27 combined.”

28 Moreover, we added a sentence addressing problems related to the Bloch decay
29 technique. Our literature review showed that the commonly mentioned problem is the
30 generally low signal intensity.

31

- 32 • The discussion about pyrogenic C should be moved to the process understanding part.
33 The last sentence (P.9, 129-31) can be deleted.

1 **Answer: Done.**

2

- 3 • P.10, l.1 : a sentence is needed introducing the fact that process understanding may be
4 limited by several different aspects related to our conceptual and experimental
5 approaches.
6

7 **Answer: We added the following introductory sentence: “The contradictions outlined
8 herein might also suggest gaps in the understanding of SOM turnover processes. Here
9 we argue that, in particular, knowledge about the turnover of SOM at mineral surfaces
10 is insufficient. This is due to the yet uncertain quantitative composition of SOM. In
11 addition, prevailing conceptual ideas and paradigms have been questioned in recent
12 years.”**

13

- 14 • 4.2. Limits in process understanding : I think that this paragraph need to be better
15 structured – temporal aspect as discussed in the beginning should be separated from
16 spatial ones (probalby the last paragraph could be moved up a little); plant activity
17 effects should be separated from the processes depending on pedology
18

19 **Answer: We revised the first part of the chapter on process understanding in order to
20 improve the structure. First, we present an introductory paragraph (see above), which
21 argues that uncertainties about processing of sorbed OM represent a major knowledge
22 gap. We then discuss current findings and hypotheses about processing of sorbed OM
23 (i.e., the `cycling downwards` model of Kaiser and Kalbitz and the possible role of root
24 exudates for turnover of sorbed SOM). Thereafter we discuss recent findings on the nm-
25 scale distribution of SOM at mineral surfaces, which might have implications for the
26 understanding of the processes occurring at mineral surfaces.**

27

- 28 • P.11, l.31 : ‘and methodological problems ‘ should be added after ‘the contradictions’
29

29

30 **Answer: Done.**

31

- 1 • P.12/13 : in the research strategies it should be mentioned that the origin of aromatics in
2 water and soil should be elucidated and specific stabilisation and destabilisation
3 mechanisms studied (e.g. photooxidation ; binding of tannins to proteins....)
4

5 **Answer: We mention the problem that the origin of the aromatic DOM is oftentimes**
6 **uncertain in the first point of the research strategies. Moreover, we added the following**
7 **paragraph to the research strategies (second point): “The causes of the commonly**
8 **observed decreasing fluxes of aromatic DOM fluxes with depth of the mineral soil need**
9 **to be re-examined. Are they really mainly the result of sorption to mineral surfaces (as**
10 **proposed herein in chapter 2), or do other processes such as the binding of tannins to**
11 **proteins or mineralization also play a decisive role?”.**

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1 **Gone or just out of sight? The apparent disappearance of**
2 **aromatic litter components in soils**

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12

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14

15 **Abstract**

16 Uncertainties concerning stabilization of organic compounds in soil limit our basic
17 understanding on soil organic matter (SOM) formation and our ability to model and manage
18 effects of global change on SOM stocks. One controversially debated aspect is the
19 contribution of aromatic litter components, such as lignin and tannins, to stable SOM forms.
20 In the present opinion paper, we summarize and discuss the inconsistencies and propose
21 research options to clear them.

22 Lignin degradation takes place step-wise, starting with (i) depolymerisation, followed by (ii)
23 transformation of the water-soluble depolymerization products. The long-term fate of the
24 depolymerization products and other soluble aromatics, e.g., tannins, in the mineral soils is
25 still a mystery. Research on dissolved organic matter (DOM) composition and fluxes
26 indicates dissolved aromatics are important precursors of stable SOM attached to mineral
27 surfaces and persist in soils for centuries to millennia. Evidence comes from flux analyses in
28 soil profiles, biodegradation assays, and sorption experiments. In contrast, studies on
29 composition of mineral-associated SOM indicate the prevalence of non-aromatic microbial-

1 derived compounds. Other studies suggest the turnover of lignin in soil can be faster than the
2 turnover of bulk SOM. Mechanisms that can explain the apparent fast disappearance of lignin
3 in mineral soils are, however, not yet identified.

4 The contradictions might be explained by analytical problems. Commonly used methods
5 probably detect only a fraction of the aromatics stored in the mineral soil. Careful data
6 interpretation, critical assessment of analytical limitations, and combined studies on DOM
7 and solid-phase SOM could thus be ways to unveil the issues.

8

9 **1 Introduction**

10 Storage and quality of soil organic matter (SOM) determine many crucial soil properties and
11 the ~~eyeles-cycling~~ of carbon (C) and essential nutrients through ecosystems. The storage of
12 SOM is determined by plant litter inputs and decomposition processes. Decomposition of
13 SOM is a significant source of atmospheric CO₂, thus, a critical parameter in climate models
14 (Schlesinger and Andrews, 2000). Decomposition rates are sensitive to global change factors
15 such as temperature, precipitation, and land use. However, our ability to understand and
16 predict such responses is limited by uncertainties about pathways of organic matter
17 transformation in soil. In particular, the question as to why some SOM components persist in
18 soil for centuries (denoted as `stable SOM` from here on) while others turn over quickly is
19 still puzzling (Schmidt et al., 2011).

20 Recent research challenges traditional theories presuming that stable SOM results from
21 neoformation of complex humic polymers in soil (`humification`). Stable SOM rather seems
22 to be composed of relatively simple organic compounds that are protected against
23 biodegradation, e.g., because they are tightly bound to mineral surfaces (Schmidt et al., 2011;
24 Kleber et al., 2015). Herein, we hold to this view but argue that, despite extensive research in
25 the last years, the chemistry and source of compounds incorporated into stable SOM is still
26 largely uncertain. In particular, the importance of aromatic compounds derived from abundant
27 plant litter components, such as lignin and tannins, is controversially debated (Figure 1). One
28 line of evidence suggests that they are important contributors to stable SOM. It bases
29 primarily on data from research on fluxes and behaviour of dissolved organic matter (DOM)
30 in soil, hence, we will denote it as the `dissolved phase line of evidence`. A contrasting line of
31 evidence suggests a quick degradation of aromatic compounds in soil derives primarily from
32 analyses of the composition of solid SOM (`solid phase line of evidence`). Herein, we sum up

1 and confront the arguments of the two views, discuss potential reasons for the controversies
2 (including limitations in analytical methods and process understanding) as well as their
3 implications for our basic understanding ~~on~~of SOM formation.

4

5 **2 Dissolved phase line of evidence**

6 The view that plant-derived aromatics are a major source of stable SOM is based on the
7 following ~~two~~-main arguments:

8 (1) DOM produced during litter decomposition and leached into mineral soil is a main
9 source of stable SOM adsorbed on mineral surfaces.

10 (2) Aromatic DOM components produced during litter decomposition are resistant to
11 mineralization and preferentially sorb to mineral surfaces. Hence, they are
12 preferentially stabilized in mineral soil.

13 ~~(2)~~(3) Root decomposition in mineral soil could be another important source of
14 aromatic DOM components that contribute to stable SOM.

15

16 **2.1 Argument 1: DOM as source of stable SOM**

17 Leaching of DOM is a major pathway for organic matter translocation from forest floor into
18 the topsoil horizons. Estimates for acidic forest soils with permanent forest floor suggest that
19 25-89% of the SOM stored in mineral soils derives from DOM (Neff and Asner, 2001;
20 Michalzik et al., 2003; Kalbitz and Kaiser, 2008), based on the typical observation of
21 decreasing DOC fluxes with depth of the mineral soil (a large compilation of data from
22 studies on forest and grassland soils is presented by Neff and Asner, 2001). Two processes
23 can explain the decrease: mineralization and sorption.

24 Sorption of DOM to mineral surfaces likely is a major process forming stable SOM in many
25 soils. Evidence for its importance comes from findings that the turnover and storage of SOM
26 in mineral soil horizons is related to the contents of reactive secondary minerals (e.g., Fe
27 hydrous oxides, short-range ordered Al hydroxides). Such relationships have been found
28 across a wide range of soil types (comprehensive data sets are have been presented in Torn et
29 al., 1997, Eusterhues et al., 2005, Kögel-Knabner et al., 2008, Kramer et al., 2012, and

1 Kleber et al., 2015). Also, higher radiocarbon age of SOM in heavy (i.e., mineral) fractions
2 compared to light density (i.e. organic) fractions indicates that sorption stabilizes organic
3 compounds (~~e.g., see data compilations of~~by Marschner et al., 2008, [Kögel-Knabner et al.,](#)
4 [2008, and](#) Kleber et al., 2015). Density fractionation procedures indicate that the total soil C
5 associated with minerals in any given location can vary from 30% to 90% (~~see data compiled~~
6 in Kleber et al., 2015). The relevance of sorptive stabilization depends on soil properties. Low
7 soil pH enhances the formation of reactive secondary minerals and favors the formation of
8 strong bonds between organic matter and the mineral surface (Kleber et al., 2015). Most
9 studies cited herein (for both lines of evidence) examined acidic soils under temperate forests,
10 in which sorptive stabilization clearly should play an important role [for the long-term storage](#)
11 [of organic matter in soil.](#)

12

13 **2.2 Argument 2: Preferential stabilization of aromatic DOM components**

14 Lignin, a macromolecule composed of phenyl propane units, is a major plant cell wall
15 component (Kögel-Knabner, 2002). Typically, lignin concentrations negatively correlate with
16 litter decomposition rates. They are the predominant control on litter decomposition within
17 biomes worldwide (Cornwell et al., 2008), indicating that the lignin macromolecule is among
18 the most persistent litter constituents. Nevertheless, results of recent studies suggest
19 significant chemical alteration and losses of lignin already within the first months [and years](#) of
20 litter decomposition (~~e.g.,~~[Kalbitz et al., 2006;](#) Preston et al., 2009; Klotzbücher et al., 2011;
21 [Duboc et al., 2014](#)). 'Degradation' of lignin has to be considered a step-wise process: (i) the
22 first step is the depolymerization of the macromolecule, releasing (mainly aromatic) water-
23 soluble depolymerisation products of varying molecular weight; (ii) these products can then
24 be further transformed, and low-molecular weight compounds are eventually taken up by
25 microorganisms to produce biomass or CO₂. Hence, losses of lignin-derived C during litter
26 decomposition can occur due to leaching of water-soluble products of an incomplete
27 degradation or as CO₂. Laboratory incubation tests on water-extractable organic matter from
28 various ~~forest floor~~[plant and soil](#) materials suggest that aromatic components are more
29 resistant to mineralization than non-aromatic components ([Pinney et al., 2000;](#) Kalbitz et al.,
30 2003 a,b; [Marschner and Kalbitz, 2003;](#) [Don and Kalbitz, 2005;](#) [McDowell et al., 2006;](#)
31 [Hagedorn and Machwitz, 2007;](#) [Fellman et al., 2008;](#) Hansson et al., 2010; [Kothawala et al.,](#)
32 [2012;](#) [Toosi et al., 2012](#)). This suggests that leaching is an important factor in loss of lignin-

1 derived matter during litter decomposition. Consistent with this conceptual model, the
2 typically high UV absorptivity of DOM leached from forest floors (i.e., higher than values
3 found for DOM leached from Oi and Oe horizons) is indicative of a large contribution of
4 resistant aromatic components (e.g., Kalbitz et al., 2007; Fröberg et al., 2007; Kalbitz and
5 Kaiser, 2008).

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6 Another factor for the export of aromatic DOM from forest floors is leaching of tannins.
7 Tannins are water-soluble polyphenols of a molecular weight ranging from 500 to 3000
8 Daltons. Tannins rapidly leach from fresh litter; most studies suggest losses of ~80% within
9 the first year of litter decomposition (Kraus et al., 2003).

10 It has been commonly found that the contribution of components likely derived from lignin
11 and tannins to DOM decreases with depth of the mineral soil (summarized in Table 1), i.e.,
12 the decrease in fluxes of these compounds with depth is more pronounced than the decrease
13 of bulk DOM. One explanation might be intensive biodegradation of aromatics in mineral
14 soil. However, this would contradict results of the DOM biodegradation studies previously
15 discussed. Hence, a more likely explanation is sorption to mineral surfaces. Laboratory
16 sorption experiments support this view; a typical observation is that lignin-derived aromatic
17 DOM components are preferentially sorbed by minerals and soils (Davis and Gloor, 1981;
18 Jardine et al., 1989; McKnight et al., 1992; Dai et al., 1996; Kaiser and Zech, 1997, 2000;
19 Chorover and Amistadi, 2001; Guo and Chorover, 2003; Kalbitz et al., 2005; Kawahigashi et
20 al., 2006; Hunt et al., 2007; Mikutta et al., 2007; Jagadamma et al., 2012; Sodano et al.,
21 2016; e.g., Kaiser et al., 1996; Chorover and Amistadi, 2001; Hunt et al., 2007), and for some
22 soils it has been shown they displace previously bound organic components from mineral
23 surfaces (e.g., Kaiser et al., 1996). The degree of preferential sorption may depend on the
24 composition of the soil mineral assemblage. Chorover and Amistadi (2001) observed that
25 high molecular weight aromatic components preferentially sorbed onto goethite, while for
26 montmorillonite no preference for aromatic moieties was observed. A likely reason for the
27 preferential sorption is the large content of carboxyl groups linked to the aromatic rings,
28 which bind to metals at mineral surfaces via ligand exchange reactions.

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30 **2.3. Argument 3: Roots as source of stabilized aromatic SOM?**

31 We have so far focused on DOM leached from aboveground litter. However, roots might also
32 be a crucial source of stable SOM. The contribution of root and aboveground litter as major

1 source of SOM has been debated in numerous studies, but the available information allows no
2 definite conclusions yet (~~see discussion in recent comprehensive discussions on the topic can~~
3 ~~be found in~~ -Lajtha et al., 2014, and Hatton et al., 2015). Presumably, the relative importance
4 of the two types of organic matter input for SOM storage in topsoils differs between
5 ecosystems (Crow et al., 2009) and the importance of root-derived matter increases with soil
6 profile depth (Rumpel et al., 2015).

7 This raises issue of whether results from aboveground litter decomposition would also apply
8 to root litter decomposition. Data by Crow et al. (2009) suggest that lignin concentrations of
9 roots are in the range of those of leaf and needle litter. Root-derived DOM shows higher
10 concentrations of aromatic compounds than DOM from foliar litter (Hansson et al., 2010;
11 Uselman et al., 2012). Hansson et al. (2010) showed that DOM production during root
12 decomposition occurs in patterns that are similar to those of needle decomposition.
13 Particularly during later decomposition stages, root-derived DOM is enriched in aromatics
14 resistant to mineralization. Hence, available information suggests that root decomposition is
15 just another important source of soluble aromatics in mineral soils. However, to the best of
16 our knowledge, studies to quantify the contribution of root-derived aromatics to DOM fluxes
17 in the field have yet to be conducted. Overall, the available information is limited to data from
18 laboratory experiments and refers to acid temperate forest soils, so that it is not yet possible to
19 draw general conclusions.

20

21 **3 Solid phase line of evidence**

22 Many of the recent conceptual papers on SOM formation are built on the assumption that
23 lignin-derived aromatics disappear quickly in soil, while SOM in mineral soils is dominated
24 by non-aromatic and microbial-derived compounds (Grandy and Neff, 2008; Schmidt et al.,
25 2011; Dungait et al., 2012; Miltner et al., 2012; Cotrufo et al., 2013; Castellano et al., 2015).
26 Empirical support is provided by studies characterizing the chemical structure of solid SOM
27 using a variety of analytical methods. ~~In the following, we sum up the most widely cited~~
28 ~~work.~~

29 Numerous studies on a wide variety of soil types used the cupric oxide (CuO) method to
30 analyse the distribution of lignin-derived phenols in profiles. ~~Most of them~~ A typical
31 observation is that the ~~reported decreasing~~ phenol contribution to SOM decreases (i) from
32 forest floor to A horizons ~~and, (ii)~~ with depth of the mineral soil, (iii) with decreasing soil

1 ~~particle size~~ (reviewed in Thevenot et al., 2010) ~~and Measured phenols also decreases~~ with
2 increasing density of soil fractions, hence, are ~~relatively small component in~~ ~~contributing little~~
3 ~~to~~ heavy (i.e., mineral-associated) and old soil fractions (e.g., ~~Leifeld and Kögel-Knabner,~~
4 ~~2005; Grünewald et al., 2006; Sollins et al., 2009; Kögel-Knabner et al., 2008; Cerli et al.,~~
5 2012). Similar results are reported by studies using pyrolysis-gas chromatography/mass
6 spectrometry (see e.g., ~~data and references provided by Grandy and Neff, 2008 and by~~
7 ~~Gleixner et al. 2002; Nierop et al., 2005; Buurman et al., 2007; Grandy and Neff, 2008;~~
8 ~~Tonneijck et al., 2010; Rumpel et al., 2012~~) and tetramethylammonium hydroxide (TMAH)
9 thermochemolysis (Nierop and Filley, 2007; Mason et al., 2012).

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10 Analysis of heavy and clay-sized soil fractions using cross polarization and magic angle
11 spinning (CPMAS) ¹³C nuclear magnetic resonance spectroscopy (¹³C-NMR) typically finds
12 high peak intensities of alkyl and O/N alkyl C (mostly assigned to polysaccharides and
13 proteins) and low peak intensities of aryl C (mostly assigned to lignin and tannins) (see
14 ~~reviews data compilations by Mahieu et al., 1999, and Kögel-Knabner et al., 2008 and Miltner~~
15 ~~et al., 2012~~). For instance, in a comprehensive study on Ah horizons from 8 European forest
16 sites, O/N alkyl C contributed up to 41-49% of total peak intensity in the <2- μ m fraction, and
17 the peak intensities were on average 10% higher than the those reported for bulk soil; the
18 intensities of aryl C in the <2- μ m fraction contributed 13-15% of total peak intensities, and
19 they were on average 24% lower than values found for bulk soil (Schöning et al., 2005). Also
20 ~~studies using near-edge X-ray absorption fine structure (NEXAFS) spectra supported the~~
21 ~~conclusions drawn from ¹³C-NMR analysis of a significant contribution of microbial-derived~~
22 ~~compounds to SOM at mineral surfaces (Lehmann et al., 2007; Kleber et al., 2011).~~

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23 -Consistent with ~~these~~ findings, the heavy soil fraction is characterized by low C/N values
24 close to those of microbial tissues (Kögel-Knabner et al., 2008).

25 ~~Analysis of microbial-derived polysaccharides by acid hydrolysis suggest an enrichment of~~
26 ~~these compounds in fine and heavy soil fractions (Kiem and Kögel-Knabner, 2003; Rumpel et~~
27 ~~al., 2010). Studies using near edge X ray absorption fine structure (NEXAFS) spectra~~
28 ~~supported the conclusions drawn from ¹³C-NMR analysis of a significant contribution of~~
29 ~~microbial derived compounds to SOM at mineral surfaces (Lehmann et al., 2007; Kleber et~~
30 ~~al., 2011).~~

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31 Lignin turnover (i.e., transformation into CO₂ or non-lignin products) in temperate arable,
32 grassland, and tropical forest soils has been estimated using a combination of isotopic labeling

1 and compound-specific isotope analysis of lignin-derived aromatics applying the CuO
2 method. Most of the studies using this approach suggest that the turnover of lignin-derived
3 aromatics is faster than the turnover of bulk SOM (Dignac et al., 2005; Heim and Schmidt,
4 2007; Heim et al., 2010). A ~~modelling~~ modeling study based on the data by Dignac et al.
5 (2005) suggested that about 90% of the lignin is mineralized as CO₂ or transformed into
6 compounds devoid of lignin-type signatures within one year (Rasse et al., 2006). However, a
7 study by Hofmann et al. (2009) suggests that after 18 years, approximately two-thirds of the
8 initial lignin phenols remained in an arable soil. The authors concluded that lignin was
9 preferentially preserved in the soil. ~~It should be noted all of these land use successional~~
10 ~~studies only determined turnover times in the top 10-30cm of the soil horizon and may not~~
11 ~~accurately represent lignin dynamics in the deeper soil.~~

13 4 Reasons for the controversies

15 4.1. Analytical limitations

16 The controversies in current literature might (partly) be due to difficulties in the analyses of
17 aromatic ~~matter-OM compounds~~ in soils. Studies on DOM typically use bulk methods for
18 inferring aromatic content, including UV absorbance and fluorescence spectroscopy.
19 Limitations of this research include lack of identification of the source of aromatic
20 compounds, and poor quantification of the fluxes. Also data on contribution of aromatic
21 components to solid SOM are semi-quantitative or qualitative.

22 Commonly applied methods such as CuO oxidation, pyrolysis or TMAH thermochemolysis
23 focus on few defined lignin-derived monomers to estimate the overall contribution of lignin.
24 These estimates, however, can largely differ depending on the method applied (Klotzbücher et
25 al., 2011). As outlined by Amelung et al. (2008), compound-specific isotope analysis of
26 lignin-derived compounds with the CuO method presumably overestimates the turnover rates
27 of lignin as only part of the lignin-derived aromatics can be extracted from soil (incomplete
28 extraction might also be a problem in all analyses of biomarkers, for which turnover times
29 typically are estimated to be faster than turnover rates of bulk SOM). Firstly, CuO oxidation
30 (as well as conventional pyrolysis or TMAH thermochemolysis) does not completely
31 depolymerize lignin (Johansson et al., 1986; Goñi and Hedges, 1992; Filley et al., 2000).

1 Secondly, lignin-derived aromatics bound to mineral surfaces are only partly assessed by the
2 CuO method (Hernes et al., 2013). Thirdly, lignin-derived aromatics might be altered in a way
3 that they escape the `analytical window` and cannot be ascribed to a lignin source anymore.
4 For instance, the CuO method yields a number of aromatic monomers of unknown origin
5 besides the lignin-derived monomers (Cerli et al., 2008). These compounds are typically not
6 quantified, and thus, not considered in estimates of the SOM composition. Hence, monomer
7 yield is a commonly used but uncertain measure of lignin concentration in soil. ~~Our~~
8 ~~knowledge about how much lignin is `hidden` (Hernes et al., 2013) in soil is still insufficient.~~
9 Solid-state CPMAS ¹³C-NMR has been widely used in the last decades to study the
10 composition of SOM. Whether ~~The uncertainties related to quantification of SOM~~
11 ~~compositions with solid state CPMAS ¹³C NMR have the results are quantitative has been~~
12 subject of an intensive ~~been intensively discussed in the literature~~ debate (see Knicker, 2011).
13 Mineral soil samples are commonly pretreated with hydrofluoric acid (HF) in order to remove
14 paramagnetic minerals that disturb the analysis. The treatment can result in significant losses
15 of SOM, and one might lose important information on SOM adsorbed onto minerals (e.g.,
16 SOM losses of 10-30% in topsoil samples and up to 90% in subsoil; Eusterhues et al., 2003).
17 Eusterhues et al. (2007) attempted to assess the chemical composition of HF-soluble SOM by
18 comparing CPMAS ¹³C-NMR spectra of untreated and HF-treated soil samples. The data
19 suggest that the composition of HF-soluble SOM considerably varies ~~differs between soil type~~
20 and soil horizons. The effect of HF treatment, thus, produces unpredictable changes in
21 composition and questions the meaning of spectra obtained on HF-treated samples. This may
22 imply that there is no general selective loss of a specific C type. Also, The approach used in
23 this study comes with the uncertainty that in the untreated soil part of the SOM probably was
24 `invisible`; i.e., in the untreated samples, SOM attached to mineral surfaces might have been
25 invisible due to the proximity to paramagnetic material (Kinchesh et al. 1995), ~~and while in~~
26 the HF-treated soil, the same SOM might have been invisible as it was become removed
27 during the treatment, thus, is was no more detectable. Further uncertainties of CPMAS ¹³C-
28 NMR arise from signal overlapping and a general low sensitivity for aromatic C in soils (e.g.,
29 Skjemstad et al., 1996; Mao et al., 2000; Simpson and Simpson, 2012).
30 ~~The common approach used to quantify the relative contribution of different C types to SOM~~
31 ~~is to integrate and compare peak areas of different spectra regions without considering any~~
32 ~~non-proportional signal responses (e.g., Kögel-Knabner 2002). It has been shown that this~~

1 ~~approach~~ ~~It has been shown that the technique~~ underestimates lignin vs. cellulose in ligno-
2 cellulose isolated from wheat ~~-(Gauthier et al., 2002)~~. Also, peaks of methoxyl C often tend to
3 be larger than those of aromatic C. Both, methoxyl C and aromatic C derive from lignin, but
4 lignin has much less methoxyl C than aromatic C. Spectra of wood samples (e.g., Bonanomi
5 et al. 2014), needle litter (Preston and Trofymow, 2015) and grass litter (McKee et al., 2016)
6 all show that patterns of methoxyl signals being larger than those of aromatic and phenolic C
7 combined.

8 -By applying Bloch decay, another type of ^{13}C -NMR technique, one can overcome the
9 problem of the reduced sensitivity for aromatic ^{13}C . The technique has been applied in studies
10 on pyrogenic organic matter, for which CPMAS ^{13}C -NMR should be even less sensitive than
11 for lignin as it is more condensed (Golchin et al., 1997; Simpson and Hatcher, 2004; Knicker
12 et al., 2005). Bloch decay ~~is~~, however, ~~also comes with problems, such as general low signal~~
13 ~~intensity. It is~~ not routinely applied in SOM research as it is an extremely time-consuming
14 experiment and the required instrument time is frequently not available (Simpson and
15 Hatcher, 2004).

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~~Pyrogenic organic matter is an important source of aromatic compounds in many soils.~~
17 ~~Despite extensive research efforts, rates and pathways of pyrogenic organic matter~~
18 ~~decomposition are still not well established (Schmidt et al., 2011; Kuzyakov et al., 2014).~~
19 ~~Analyses of benzenecarboxylic acids as molecular markers suggest that aromatic compounds~~
20 ~~derived from pyrogenic organic matter are translocated within soil profiles and bind to~~
21 ~~mineral surfaces (Haumaier, 2010). The quantitative contribution of pyrogenic organic matter~~
22 ~~to DOM in soil is, however, still poorly studied (Smebye et al., 2016). Bulk analyses of~~
23 ~~aromatic matter used in most research on DOM fluxes cannot distinguish if the compounds~~
24 ~~derive from plant litter or from pyrogenic organic matter. This limits our understanding of the~~
25 ~~processes controlling turnover of aromatics. If a significant part of aromatic DOM in mineral~~
26 ~~soil derives from pyrogenic organic matter, the `loss` of plant litter derived aromatics in~~
27 ~~mineral soil would be even more pronounced. Problems distinguishing sources of aromatic~~
28 ~~compounds can also occur in analysis of solid-phase SOM with CPMAS ^{13}C -NMR analysis~~
29 ~~(Simpson and Hatcher, 2004).~~

31 4.2. Limits in process understanding

1 ~~Another source of uncertainty in quantifying the role of aromatics in SOM stabilization is that~~
2 ~~the timescales of the different processes affecting aromatic compounds vary considerably.~~
3 ~~Solubilization, leaching, and sorption of lignin-derived compounds might occur in a few days.~~
4 ~~In contrast, the composition of SOM as well as the mineral assemblage in soils is the result of~~
5 ~~years to centuries of biogeochemical processing. Possibly, one cannot simply extrapolate~~
6 ~~from patterns observed in 'short term' DOM dynamics to explain 'long term' SOM formation.~~
7 ~~In their comprehensive review on organo-mineral interactions, Kleber et al. (2015) question~~
8 ~~the view that strong bonds between organic matter and mineral surfaces really guarantee slow~~
9 ~~turnover rates and hence long turnover times. Many factors that potentially determine the~~
10 ~~long-term fate of sorbed organic matter are still not well understood. They include exchange~~
11 ~~reactions between sorbed organic matter and new organic matter inputs or the impact of~~
12 ~~mineral weathering activity of roots on stability of sorbed organic matter. As proposed in~~
13 The contradictions outlined herein might also suggest gaps in the understanding of SOM
14 turnover processes. Here we argue that, in particular, our knowledge about the turnover of
15 SOM at mineral surfaces is insufficient. This is due to the yet uncertain quantitative
16 composition of SOM. ; In addition, but also assumptions of current-prevailing conceptual
17 ideas and paradigms have been questioned in recent years.
18 It has been frequently observed that the ¹⁴C age of DOM increases with profile depth. Kaiser
19 and Kalbitz (2012) proposed that this can be explained by a), the commonly observed
20 increasing ¹⁴C age of DOM with profile depth might be explained by a) temporal sorptive
21 immobilization, followed by microbial processing and re-release of altered compounds into
22 soil solution. That would mean the assumption that aromatic compounds are stable after being
23 sorbed onto mineral surfaces is could be erroneous. Hence, the microbial processing of sorbed
24 compounds might be the 'missing argument' that proves the view of a fast disappearance of
25 aromatic compounds in mineral soil. However, these processes are have hardly been studied
26 yet, and empirical evidence for their importance is missing.
27 Also, root activity might have a-significant effects on stability and composition of SOM
28 sorbed onto mineral surfaces. The recent study of Keiluweit et al. (2015) showed that root
29 exudation of oxalic acid promotes the-release of SOM-sorbed onto-mineral
30 surfacescompounds into soil solution through dissolution of mineral phases. Taken together,
31 the view that DOM leached from litter decomposition sorbs onto mineral surfaces and then
32 contributes to stable SOM might be too simple. Processing of sorbed material may exert a

1 ~~significant effect on SOM dynamics. This possibly prevents the long-term storage of plant~~
2 ~~derived aromatics in soil and explains the controversies outlined herein. Besides oxalic acid,~~
3 ~~many other organic compounds are released from living roots (the so-called~~
4 ~~‘rhizodeposition’). Estimates suggest that rhizo-deposition constitutes a loss term of 17% (on~~
5 ~~average) of the net C assimilated by plants (Nguyen, 2003). It comprises many other low-~~
6 ~~molecular weight, compounds, including compounds (organic acids, simple sugars, amino~~
7 ~~sugars, phenolics,) as well as high-molecular compounds (exoenzymes, root cells) are~~
8 ~~released from living roots~~ (Wichern et al., 2008). Most of the compounds are degraded
9 quickly, but a smaller portion ~~might seems to~~ contribute to stable SOM (Nguyen, 2003;
10 Pausch et al., 2013). The rhizosphere is considered a ‘hot-spot’ in soil, where microbial
11 processes are ~~accelerated as they are~~ not C-limited (Kuzyakov and Blagodatskaya, 2015).
12 ~~Hence, root activity possibly may accelerate the turnover of sorbed plant-derived aromatics,~~
13 ~~and at the same time foster the production and stabilization of microbial-derived compounds.~~
14 ~~Studies addressing these assumptions are not yet available. Possibly, root activity not only~~
15 ~~fosters production, but also stabilization of microbial-derived compounds in soil. To date, root~~
16 ~~activity effects on DOM dynamics and stabilization of plant-derived aromatics at mineral~~
17 ~~surfaces are poorly studied. This is partly because traditional analyses are focused on mixed~~
18 ~~soil samples, while the rhizosphere may only comprise a small volume of bulk soil, and fluxes~~
19 ~~of DOM are averaged for a soil surface (Kuzyakov and Blagodatskaya, 2015).~~

20 Recent investigation at the submicrometer scale using Nano Secondary Ion Mass
21 Spectrometry (NanoSIMS) or NEXAFS in combination with scanning transmission X-ray
22 microscopy suggest that SOM associated with clay-sized minerals exists in small patches of
23 varying chemical composition (Lehmann et al., 2008; Remusat et al., 2012; Vogel et al.,
24 2014). Distinct patches of predominantly aromatic C can be differentiated from patches
25 dominated by aliphatic C (Lehmann et al., 2008). Knowledge about processes controlling the
26 sub-micro-meter-scale distribution of SOM on mineral surfaces is still limited. Some of the
27 patches are cell wall structures of microorganisms, which may contribute to stable SOM as
28 they are composed of insoluble polymers and possibly ~~sorb-attach~~ to the mineral surface
29 (Miltner et al., 2012). Hence, stable sorbed organic matter might not only be derived from
30 low-molecular weight compounds. On the other hand, it needs to be considered that
31 microbial-derived compounds are continuously synthesized at the mineral surface. The
32 microorganisms might use some of the older C (¹⁴C age) for synthesis of relatively labile
33 compounds. The age of the C atoms is, thus, decoupled from the stability of the organic

1 matter, and microbial-derived compounds may `mimic` a similar or even higher stability than
2 the plant-derived compounds (Gleixner, 2013). Hence, concentrations or ¹⁴C age of microbial-
3 derived compounds at mineral surfaces do not per se allow for conclusions on their
4 contribution to stable SOM.

5 Pyrogenic organic matter is an important source of aromatic compounds in many soils.
6 Despite extensive research efforts, rates, and pathways of pyrogenic organic matter
7 decomposition are still not well established (Schmidt et al., 2011; Kuzyakov et al., 2014).
8 Analyses of benzenecarboxylic acids as molecular markers suggest that aromatic compounds
9 derived from pyrogenic organic matter are ~~translocated~~transported within soil profiles and
10 bind to mineral surfaces (Haumaier, 2010). The quantitative contribution of pyrogenic organic
11 matter to DOM in soil is, however, still poorly studied (Smebye et al., 2016). Bulk analyses of
12 aromatic matter used in most research on DOM fluxes (i.e., UV absorption) cannot
13 distinguish if the compounds derive from plant litter or from pyrogenic organic matter. This
14 limits ~~our~~the understanding of the processes controlling turnover of aromatics. If a significant
15 part of aromatic DOM in mineral soil derives from pyrogenic organic matter, the `loss` of
16 plant litter-derived aromatics in mineral soil would be even more pronounced.

17

18 **5 Implications and future research strategies**

19 The contradictions outlined herein limit our basic understanding on SOM formation, and our
20 ability to model and manage effects of global change on SOM stocks.

21 For instance, elevated atmospheric CO₂ levels can induce increasing concentrations of
22 aromatic components in plant litter (Cotrufo et al., 1994; Tuchman et al., 2002), and this
23 raises the question whether this causes enhanced or decreasing storage of SOM in mineral
24 soils. If aromatic matter is quickly degraded, and mineral-associated SOM primarily derives
25 from microbial sources (as suggested by the solid SOM line of evidence), a `microbial filter`
26 would control the built-up of stable SOM, which may then be determined by the microbial
27 substrate use efficiency (i.e., the amount of organic C used by the microbial community to
28 build biomass vs. the amount that is mineralized). As such, Cotrufo et al. (2013) hypothesized
29 that input of labile substrates fosters the build-up of stable SOM. Available data on effects of
30 litter quality and SOM formation are, however, inconsistent (Castellano et al., 2015), and we
31 believe that understanding ~~on~~ these effects is in part limited by uncertainties about the
32 incorporation of aromatics into stable SOM.

1 The issue is also related to the question of links between chemical structure of organic matter
2 and its persistence. It is oftentimes assumed that structural properties of plant-derived matter
3 do not determine stable SOM formation. This argument is based on data suggesting that
4 specific compound classes (lignin, cellulose, alkanes, proteins etc.) turn over faster than bulk
5 SOM (Schmidt et al., 2011). However, conclusions of DOM research imply that structure
6 plays a role for the behavior of organic compounds in soil, and eventually their contribution to
7 stable SOM: soluble aromatics may resist oxidation by microbes as they yield less energy
8 than other structures; furthermore, they bind to mineral surfaces due to carboxyl groups
9 attached to the rings.

10 How could we resolve the controversies? Based on our literature analysis we propose the
11 following research strategies:

- 12 • ~~Our knowledge on aromatics in soils is limited by the analytical constraints in~~
13 ~~analysis of aromatics in soil. Quantification of total amounts and sources of aromatics~~
14 ~~in soil are still problematic. Even if the problems cannot be fully solved with the~~
15 ~~currently available techniques, there might be strategies to obtain improved estimates.~~
16 ~~The work of Hernes et al. (2013) provides a first hint about how much lignin might be~~
17 ~~not accessible to CuO oxidation analyses. The authors evaluated the extraction~~
18 ~~efficiency for lignin-derived aromatics bound to different minerals. The size of the~~
19 ~~non-extractable fraction depended strongly on the mineral. Almost all of the~~
20 ~~aromatics bound to ferrihydrite were extractable, but for kaolinite the non-extractable~~
21 ~~fraction made >40%. But how about extractability in soil under field conditions?~~
22 ~~Possibly, a combination of tracking of C isotopes, DOM flux/composition~~
23 ~~assessment, and analysis of solid-phase SOM composition could provide better~~
24 ~~estimates on the hidden aromatics in soil.~~
- 25 • ~~More studies addressing links between microbial processes, composition/fluxes of~~
26 ~~DOM, and composition of solid phase SOM are needed. The study by Kramer et al.~~
27 ~~(2012) is a first attempt to relate the fluxes of aromatics to solid phase SOM~~
28 ~~properties. The causes of the commonly observed decreasing fluxes of aromatic DOM~~
29 ~~fluxes with depth of the mineral soil need to be re-examined. Are they really mainly~~
30 ~~the result of sorption to mineral surfaces (as proposed herein in eChapter 2), or do~~
31 ~~other processes such as the binding of tannins to proteins or mineralization also play a~~
32 ~~decisive role? In particular, Moreover, the presumed microbial processing of sorbed~~

1 material, causing desorption and subsequent mineralization or further transport in the
2 soil profile is poorly studied. Knowledge gaps also exist concerning the question
3 whether root activity affects de-/sorption processes. Eventually, these processes might
4 cause loss of aromatic compounds. ~~A related question is how root activity affects de-~~
5 ~~/sorption processes?~~

- 6 • Computer simulations could help to unravel the complex interrelationships between
7 DOM fluxes and solid-phase SOM composition. Recently developed models integrate
8 sorption, DOM transport, and microbial processes (Ahrens et al., 2015). In order to
9 address the problems discussed herein, effects of molecular structure on behavior of
10 the compounds in soil (e.g., differences in mineralization rate and affinity for sorption
11 between aromatics and non-aromatics) could be implemented in the models, in order
12 to develop novel hypotheses on turnover of plant-derived aromatics.

~~13 • Our knowledge is limited by the constraints in analysis of aromatics in soil.~~
14 ~~Quantification of total amounts and source of aromatics in soil are still problematic.~~
15 ~~Even if the problems cannot be fully solved with the currently available techniques,~~
16 ~~there might be strategies to obtain improved estimates. The work of Hernes et al.~~
17 ~~(2013) provides a first hint about how much lignin might be not accessible to CuO~~
18 ~~oxidation analyses. The authors evaluated the extraction efficiency for lignin derived~~
19 ~~aromatics bound to different minerals. The size of the non-extractable fraction~~
20 ~~depended strongly on the mineral. Almost all of the aromatics bound to ferrihydrite~~
21 ~~were extractable, but for kaolinite the non-extractable fraction made >40%. But how~~
22 ~~about extractability in soil under field conditions? Possibly, a combination of tracking~~
23 ~~of C isotopes, DOM flux/ composition assessment, and analysis of solid phase SOM~~
24 ~~composition could provide better estimates on the hidden aromatics in soil.~~

26 Acknowledgements

27 TK acknowledges the support of his research on lignin degradation from the German
28 Research Foundation (DFG).

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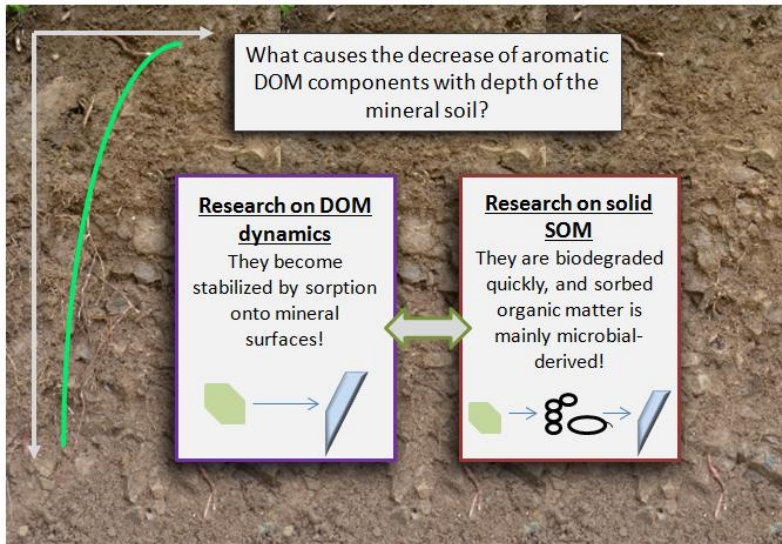
1 Table 1. Evidence from field studies suggesting that dissolved aromatics (products of lignin
 2 depolymerization or tannins) disappear quickly once entering mineral soils.

3

Reference	Study site/ soil type	Result
Qualls and Haines 1991	Oak-hickory forest in mountain region of North Carolina; soil types: Umbric Dystrochrept, Typic Hapludult, Typic Dystrochrept	Selective removal of hydrophobic acids as DOM percolates through the mineral soil.
Cronan 1985	Forests, North-Western USA; soil types: Dystrochrept, Haplorthod	Selective removal of hydrophobic acids as DOM percolates through the mineral soil.
Zech et al. 1994	Spruce forest in Bavaria, Germany; Soil types: Typic Dystrochrepts, Entic Haplorthods, Typic Haplorthods	Selective removal of lignin-derived phenols (determined with the CuO method) as DOM percolates through the mineral soil.
Gallet and Pellissier 1997	Bilberry-spruce forest in Alps, France; soil type: Humoferric Podzol	Selective removal of lignin-derived phenols (as well as of total phenols) as DOM percolates through the mineral soil.
Kaiser et al. 2004	Spruce forest in Bavaria, Germany; Soil type: Haplic Arenosol	Selective removal of lignin-derived phenols, hydrophobic compounds, and total aromatic C as DOM percolates through the mineral soil.
Dai et al. 1996	Spruce forest, Maine; Soil type: Aquic Haplorthods	Selective removal of hydrophobic acids and aromatics (¹³ C-NMR data) as DOM percolates through the mineral soil.
Lajtha et al. 2005	Coniferous forest, Oregon, USA; soil type: Typic Hapludands	Selective removal of hydrophobic acids as DOM percolates through the mineral soil.
Sanderman et al. 2008	Mediterranean climate; forest and grassland soils; soil types: Haplustols and Haplohumults	Decrease in UV absorbance (a measure for content of aromatics) as DOM percolates through the mineral soil.
Hassouna et al. 2010	Mediterranean climate; maize field; soil type: fluvic hypercalcaric cambisol	Decrease in contents of aromatic compounds (UV absorbance, fluorescence spectroscopy) in water-extractable organic matter with depth of the mineral soil.
Nakashini et al. 2012	Beech forest, Japan; soil: "brown forest soil"	Decrease in contents of hydrophobic acids in water-extractable organic matter with depth of the mineral soil.

4

- 1 |
2 Figure 1. Conflicting views on the fate of soluble aromatics once they enter the mineral soil
3 (see text for references).



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