

Carbon nanomaterials in clean and contaminated soils: environmental implications and applications

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Abstract

The exceptional sorptive ability of carbon nanomaterials (CNMs) for hydrophobic organic contaminants (HOCs) is driven by their characteristically large reactive surface areas and highly hydrophobic nature. Given these properties, it is possible for CNMs to impact on the persistence, mobility and bioavailability of contaminants within soils, either favourably through sorption and sequestration, hence reducing their bioavailability, or unfavourably through increasing contaminant dispersal. This review considers the complex and dynamic nature of both soil and CNM physicochemical properties to determine their fate and behaviour, together with their interaction with contaminants and the soil micro-flora. It is argued that assessment of CNMs within soil should be conducted on a case-by-case basis, and further work to assess the long-term stability of sorbed contaminants and the toxicity of CNMs is required before their sorptive abilities can be applied to remedy environmental issues.

Key words: Carbon nanoparticles; bioavailability; remediation; hydrophobic contaminants; soil

1. Introduction

With the continued up scaling of carbon nanomaterial (CNM) production (Nowack and Bucheli, 2007) as well as the diverse array of consumer (Sharma and Ahuja, 2008), medical (Peretz and Regev, 2012) and industrial applications in which they are increasingly becoming incorporated, widespread environmental release of these physically and chemically unique macromolecules has become inevitable (Köhler et al., 2008). Once released, soils are likely to be a primary repository (Mueller and Nowack, 2008; Gottschalk et al., 2009), with the quantities anticipated to increase on an annual basis (Gottschalk et al., 2009). In spite of this, studies focused on CNMs within soils are scarce, and many areas of uncertainty remain. Understanding the interactions between CNMs, soils and components therein is therefore an urgent and essential aspect of any risk assessment process.

In their pristine form, CNMs are broadly characterised by their large reactive surface areas, highly hydrophobic characteristics and high degree of bio-geochemical recalcitrance. They are known to be toxic to various soil microbiota (Riding et al., 2012a; Riding et al., 2012b), and possess a high affinity for the sorption of a range of hydrophobic organic compounds (HOCs), such as polycyclic aromatic hydrocarbons (PAHs), and polychlorinated biphenyls (PCBs) (Pan and Xing, 2010). As both PAHs and PCBs are important classes of hydrophobic, toxic organic compounds, which are both abundant and persistent in soils (Stokes et al., 2005), the potential for CNMs to modify the availability and mobility of HOCs, either favourably through sorption and sequestration, or unfavourably through increasing contaminant dispersal, is currently unknown. Presently, there is only limited and occasionally contradictory information regarding the implications of contaminants while sorbed to CNMs, as well as the fate and behaviour of CNMs in uncontaminated soils. Exploring these issues in light of the emerging nature of CNMs as xenobiotic soil components is therefore essential.

This review seeks to answer three key questions. (i) What factors influence the behaviour and fate of CNMs within the soil environment? (ii) To what extent can CNMs influence the sorption,

desorption and mobility of contaminants in soils? (iii) What are the impacts of CNMs on soil microorganisms and the biodegradation of contaminants in soils?

2. Carbon nanomaterial diversity and detection

Within the environment, some CNMs can occur naturally or have close naturally occurring relatives due to various environmental events (Heymann et al., 1994; Chijiwa et al., 1999; Velasco-Santos et al., 2003; Esquivel and Murr, 2004). However, concentrations occurring naturally are likely to be relatively small (0.1 to 0.2 parts per million) (Heymann et al., 1994; Chijiwa et al., 1999). Therefore, when referring to CNMs, this review explicitly focuses on those that are anthropogenic in origin.

The properties of CNMs vary dramatically between the different methods of production, functionalization status and cleaning/purification methods employed (Nowack and Bucheli, 2007). Hence, determining their environmental behaviour is all the more challenging, and generalisation of the characteristics of CNMs is not possible, with each type requiring careful characterisation (Nowack and Bucheli, 2007). Of the many different forms of CNMs available, this review focuses specifically on carbon nanotubes (CNTs) and C₆₀ fullerene, which are two of the most widely utilised and investigated classes of CNMs (Mueller and Nowack, 2008; Gottschalk et al., 2009; 2010; Petersen and Henry, 2012).

To date, CNTs are arguably the most promising of all nanomaterials produced (Giles, 2006). In their pristine form, CNTs are extremely hydrophobic and consist of graphene sheets rolled into nanoscale diameter cylinders, the ends of which may contain spherical fullerene cappings (Mauter and Elimelech, 2008). One single-rolled graphite sheet is called a single-walled carbon nanotube (Fig. 1A) (SWCNT), while several SWCNTs nested together in a concentric fashion comprise a multi-walled carbon nanotube (MWCNT) (Fig. 1B) (Pan and Xing, 2008). They consist of sp² carbon atom arrangements in a fused benzene ring configuration, which results in exceptional physicochemical properties and consequentially their incorporation into a vast array of composite

materials (Liu et al., 1999; Snow et al., 2005; Mauter and Elimelech, 2008; Almecija et al., 2009).
An excellent and more detailed discussion about the unique physicochemical properties of CNTs is
provided by Mauter and Elimelech (2008).

Fullerenes are spherically arranged carbon atoms resembling a geodesic dome. The size of the
fullerene dome can vary depending on the number and spherical configuration of carbon atoms.
C₆₀ Fullerene (Buckminster fullerene or Bucky Ball) (Fig. 2) has arguably the best defined
physicochemical parameters, produced in the largest quantities and has been the focus of most
scientific engagement (Eleanor and Frank, 2000; Petersen and Henry, 2012). C₆₀ is comprised of a
spherical configuration of 5- and 6-carbon rings, consisting of 60 carbon atoms in total. It
commonly exists as nano-C₆₀ (nC₆₀) particles (regarded as the most environmentally relevant
form), which are crystalline structures containing 100-1000 C₆₀ molecules (Colvin, 2003; Sayes et
al., 2004). Presently, fullerenes have proposed applications in biology (Lucafò et al., 2012) and
electronic/optical devices as thin films combined with polymers (Richards et al., 2012).

Unlike organic chemicals with well-defined structures, the diversity of particle sizes, lengths,
diameters, charges, surface areas, coatings, molecular weight, impurities and aggregation states,
which are often specific to their particular environmental compartment and not necessarily
constant, limits their detection and characterisation in soils through chromatographic techniques
(Petersen et al., 2011). In addition, as the life cycles of CNM containing products are likely to vary
greatly, the routes by which these materials enter the soil environment is also likely to be highly
variable (Pan and Xing, 2012). Further complicating their detection is the emerging nature of
manufactured CNMs as soil xenobiotic components, and hence their presently low
concentrations, together with their interaction with naturally occurring nanomaterials and other
environmental components, which leads to particles with sizes and compositions that significantly
differ from their native forms (Nowack and Bucheli, 2007; Darlington et al., 2009; Chen et al.,
2011). As such, careful consideration of multiple environmental variables is required to determine
their impact on CNM fate and behaviour.

98 3. CNM behaviour and fate within the soil environment

99 Once released into the soil, the fate and behaviour of CNMs is governed by their interactions
100 with various components within the environment. Derjaguin Landau Verwey Overbeek (DLVO)
101 interactions, such as electrostatic interactions and van der Waals (vdW) forces, and non-DLVO
102 interactions, such as hydrogen bonding and steric hindrance, ultimately determine the mobility,
103 aggregation and adhesion of CNMs within soils. These forces may operate in concert to various
104 extents, with the predominating force controlled by factors such as the properties and quantity of
105 soil organic matter (SOM), characteristics of inorganic matter, together with properties of CNMs
106 themselves. Each of these factors are heavily influenced by variables that are not necessarily
107 constant over time, such as pH and ionic strength.

108 3.1 The impact of soil organic matter

109 **SOM plays** a substantial role in both the fate and behaviour of CNMs through alterations in
110 the dominance of the various DLVO and non-DLVO interactions. SOM (which consists primarily of
111 decomposed plant and animal remains (Lee et al., 1981)) is an all-encompassing term describing
112 organic matter (OM) dispersed ubiquitously throughout the soil environment, and is composed of
113 a heterogeneous mixture of lipids, carbohydrates, carboxylic acids, humic substances, hydrophilic
114 acids, proteins, carbohydrates, hydrocarbons and amino acids. However, the mechanism by which
115 SOM maintains CNMs stability in suspension is still under investigation and debate (Dinesh et al.,
116 2012). Specifically, the aim of this section is therefore to provide an overview of recent
117 investigations in which the manner of solid SOM, dissolved organic matter (DOM), humic acid
118 (HA) and tannic acid (TA) influence the behaviour of CNMs.

119 Adsorption of molecular DOM onto CNMs occurs through either aromatic ring sorption or
120 binding of aliphatic chains via π - π or CH- π interactions, leaving the hydrophilic moieties exposed
121 (Lin and Xing, 2008). Consequentially, the surfaces of CNMs significantly change from a

hydrophobic, aromatic-like structure, to that of the organic, hydrophilic functional groups in DOM (Zhang et al., 2011a), with potentially large implications for other DLVO and non-DLVO interactions. In studying nC_{60} , Kwon (2012), found the type of DOM determined suspension stability, with those containing long, hydrophobic carbon backbones readily adsorbing via vdW interactions onto nC_{60} surfaces, promoting their stability. However, Zhang et al. (2011a) found peat (diagenetically young SOM) in dissolved form (DOM) prevented MWCNT aggregation through both steric hindrance and electrostatic repulsion in sodium concentrations >4 mM or in solutions of $pH \geq 4$ (Zhang et al., 2011a). Increasing ionic strength resulted in greater adsorption of DOM onto MWCNTs (Hyung et al., 2006; Hyung and Kim, 2008; Zhang et al., 2011a) due to reduced electrostatic repulsion between the DOM and the particle surface. As a result, cations impairing electrostatic interactions at high ionic strengths in the presence of DOM may slightly shift the relative importance of suspension mechanisms from electrostatic repulsion, in favour of steric hindrance (Zhang et al., 2011a); broadly similar results have been identified with nC_{60} (Qu et al., 2012). Ionic strength therefore primarily affects the balance between electrostatic repulsion and steric hindrance mechanisms of particle suspension in the presence of DOM.

Typically, frequently occurring cations within the environment (K^+ , Na^+ , Ca^{2+} and Mg^{2+}) induce aggregation and deposition in systems devoid of SOM through reductions in electrostatic repulsion between particles and hence CNM stability (Zhang et al., 2012b). The influence of cations on the behaviour of CNMs is well illustrated by the extent to which physical straining (filtering out) of nC_{60} occurs in saturated porous media. Zhang et al. (2012a) found that columns of pure quartz resulted in very limited nanoparticle deposition even at low flow velocities, whereas a heterogeneous sandy soil with low OM content and small, irregular and rough grains of sand, significantly inhibited nC_{60} transport. With the addition of $CaCl_2$, greater deposition of nC_{60} was observed in both sand and soil; however, significantly more straining occurred in the soil due to the greater number of complexation sites for Ca^{2+} clay and OM fractions relative to sand (Zhang et al., 2012a). For both the sand and soil columns, Ca^{2+} had a much larger effect on the transport of

nC_{60} than Na^+ at the same ionic strengths (Zhang et al., 2012a). This most likely occurred due to efficient neutralisation of surface charges on both nC_{60} and sand and soil particles by Ca^{2+} relative to Na^+ , which reduced the electrostatic repulsion (Kuznar and Elimelech, 2004; Zhang et al., 2012a).

Solid peat, however, may behave differently to molecular DOM under environmentally relevant ionic conditions, as particle phase distributions may be altered due to direct sorption, as well as the possibility of DOM or cation release from the soil particles themselves (Zhang et al., 2011a). In the absence of sodium ions, Zhang et al. (2011a) found no adsorption of MWCNTs to solid peat, indicating a limited affinity of DOM-MWCNT composites towards the solid phase relative to the aqueous phase, possibly due to electrostatic repulsion and hydrophilicity of DOM coated nanotubes. With the addition of Na^+ ions, the relative affinity between DOM-MWCNTs and peat was increased due to reductions in the charge potential and subsequent increase in interactions occurring through hydrophobic interactions and/or vdW attraction (Zhang et al., 2011a). Zhang et al. (2011a) concluded that increasing ionic strength, such as in environments containing seawater or hard freshwater, increased precipitation of MWCNTs from the aqueous phase will occur, impeding their transport within the environment.

Consideration of the ζ -potential (diffuse surface charge) of CNMs in relation to the soil matrix is important when considering the dynamics of smaller particles dispersal, transport and deposition, for which charge forces are likely to be highly influential relative to larger particles or agglomerates (Darlington et al., 2009). Its measure, in part, provides information on the likely mobility, rates of interaction and aggregation status due to electrostatic forces generated by charged surfaces (Hu et al., 2005; Jafar and Hamzeh, 2013). Pristine CNTs typically express limited surface charge (Mohanty et al., 2007); however, Wang et al. (2008a) found that the ζ -potential of CNTs with a HA coating was highly negative, which resulted in electrostatic repulsion between the particles and hence stability when partitioned into the aqueous phase. They concluded that mobility and environmental transport within typically negatively charged porous media, such as

certain types of soils was highly likely, with particles remaining stable over a wide range of ionic strengths (Wang et al., 2008a). A reduction in pH to the point at which the CNTs had no charge was identified as an effective means of causing CNTs to precipitate, through the destabilization of the HA coatings (Wang et al., 2008a).

The properties of the humic substances dictate the extent to which DLVO and non-DLVO interactions dictate particle behaviour. TA (Chibowski et al., 1998) and HA sorbed to CNTs enhances stabilisation in water through reducing vdW forces between particles and increasing steric repulsion (Terashima and Nagao, 2007; Ren et al., 2011). However, Qu et al. (2012) identified that high molecular weight (HMW) HAs were more effective in promoting suspension stability due to stronger steric repulsion than that of low molecular weight (LMW) HAs. Similarly, HAs containing large quantities surfactive domains, such as those which are strongly hydrophilic and lipophilic, promote the dispersal of CNTs in solutions, while those containing carbohydrates and predominantly hydrophilic domains resulted in limited dispersal (Chappell et al., 2009).

The composition of SOM in relation to ionic strength and pH dictates the behaviour of CNMs within soils. Presently, however, insufficient data regarding the relative impact of different SOM fractions and combinations on DLVO and non-DLVO forces in soils is lacking, reducing an ability to estimate how CNMs may behave based on analysis of soil OM content. Furthermore, in addition to the organic fraction of soils and coating of CNMs, the role of the inorganic fraction in determining particle behaviour must also be considered.

3.2 The impact of soil inorganic matter

In addition to the organic fraction, CNM stability in saturated soil/water suspensions is strongly influenced by the impact of the inorganic fraction, and is largely neglected within the present literature. Broadly, all soils can be divided into two groups; permanent-charge (P-C) and variable-charge (V-C) (Sollins et al., 1988). In P-C soils, the substitution of ions with lower valence for ions with higher valence results in the alteration of crystal lattice structures within layer-

silicate clays (illite, smectite, chlorite and kaolin), and a permanent charge deficit, which persists irrespective of variations in the composition of soil solutions and pH (Sollins et al., 1988). In V-C soils, protonation and de-protonation of surface hydroxyl groups results in the positive charge and hence anion exchange capacity (AEC); whereas deprotonation results in cation exchange capacity (CEC) (Sollins et al., 1988). The structure of V-C soils is also modified in response to increasing pH, resulting in increased repulsion and more limited aggregation (Sollins et al., 1988). Both P-C and V-C surfaces are present in all soil types; however, only one charge system typically dominates, dictated largely by soil mineralogy (Sollins et al., 1988). While V-C soils occur more frequently in tropical regions due to the typical mineralogical composition which forms under humid, warm conditions, they do not occur ubiquitously, and many areas with predominantly P-C characteristics occur (Sanchez, 1976; Sollins et al., 1988). Hence, while V-C soils represent a small fraction of global soil types, interactions between nanoparticles and soils are likely to be much more dynamic relative to those with a P-C. Despite this, almost all investigations have predominantly focused on P-C soils, restricting the applicability of CNM fate and behaviour investigations.

The behaviour of CNMs in V-C soils has been assessed by Zhang et al. (2012b), who investigated the stability of MWCNTs suspended in water with soil minerals kaolinite, smectite and shale over a range of sodium concentrations. Without additional Na^+ , no significant difference in the stability of MWCNTs between each of the soil minerals was observed; however, with increasing ionic strength, the removal of MWCNTs from the aqueous phase followed the order smectite > kaolinite > shale (weakest to largest MWCNT association). As ionic strength increased, the MWCNT removal tendency for smectite and kaolinite was inversely correlated to the mineral surface potential. However, the electrostatic potential of shale is higher than either kaolinite or smectite, yet shale demonstrated the strongest sorption of MWCNTs (Zhang et al., 2012b). This was attributed to the large, hydrophobic, organic content of shale, which is able to strongly sorb MWCNTs. Hence, under these soil conditions, the transport of CNMs in soils is directly correlated

with mineral hydrophobicity, but inversely correlated with surface potential (Zhang et al., 2012b). It is therefore imperative that studies of nanomaterial fate and behaviour in soils include detailed information regarding soil mineralogy.

3.3 The impact of CNM preparation methods, functional groups and UV exposure

In addition to the influence of soil type and properties, the properties of CNMs themselves vary greatly depending on an array of parameters as commercial applications of CNMs will likely employ surface functional groups and a variety of different preparation techniques, nanoparticle properties and behaviour within the environment will become increasingly complex (Turco et al., 2011). For example, the physicochemical properties of pristine nC_{60} , such as surface charge and particle size, heavily depends on the method of synthesis and preparation, with a corresponding impact upon the stability of the nanomaterial (Chen and Elimelech, 2008; 2009; Isaacson et al., 2011). As discussed previously, deposition of CNMs in their pristine state is highly susceptible to variations in ionic strength; however, the exact extent of sensitivity is known to vary significantly according to the method of preparation. For example, deposition of nC_{60} occurs in NaCl at concentrations of 18 mM when prepared by sonication (100 W probe, 30 min) (Qu et al., 2012) and 30 and 32 mM NaCl when prepared by solvent exchange methods (Chen and Elimelech, 2008; Smith et al., 2008). However, most investigations have been conducted in simple solutions of electrolytes using pristine nC_{60} , and in complex environmental systems (Qu et al., 2012). As a result, the effects of cation valence on the stability of CNMs may substantially differ to those in a laboratory setting.

Typically, agglomeration of CNMs in the presence of divalent (Ca^{2+}) cations occurs to a greater extent than with monovalent (Na^{+}) cations. However, when exposed to solar irradiation, nC_{60} can undergo surface oxidation and decomposition (Hou and Jafvert, 2008; 2009), with large implications for environmental behaviour and fate (Qu et al., 2012). Following exposure to UV-A (the largest component of UV in sunlight), the oxygen containing functional groups formed on nC_{60} hindered aggregation in NaCl solutions due to their elevated hydrophilicity and negative

surface charge (Hou and Jafvert, 2009). Conversely, neutralisation of the negative surface charge on oxidised nC_{60} due to interactions with Ca^{2+} ions when suspended in $CaCl_2$ can result in particle agglomeration (Li and Liang, 2007). This potentially occurs due to the charge screening ability of Ca^{2+} relative to Na^+ , which reduces the stability of colloids (Li and Liang, 2007). Qu et al. (2012) expanded upon this work through studying the effects of UV-A exposure for either 20 h or 7 d on the rate of deposition and the attachment efficiency of nC_{60} on to silica bead surfaces. The stability of nC_{60} increased proportionally with increasing UV-A exposure time against aggregation in solutions containing NaCl, which was attributed to the increase in surface oxidation and hydrophilicity (Qu et al., 2012). Further, while the attachment efficiency of nC_{60} exposed to UV-A for 7 d was at a maximum in NaCl concentrations of 250 mM, attachment decreased following an increase in NaCl concentration to 300 mM. This indicates that stability of nC_{60} containing oxygen surface functional groups was attributed to the hydration force and not DLVO forces, which was more significant with the 7 d UV exposed nC_{60} than either the pristine or the 20 h UV due to the greater hydrophilicity (Qu et al., 2012).

In other investigations, surface immobilisation of macromolecules, such as HAs at environmentally relevant concentrations, has increased the solubility of C_{60} due to the effect of steric hindrance caused by the sorbed SOM and a reduction in the hydrophobicity of the nanoparticle surface, preventing re-aggregation and reducing attachment efficiency (Li et al., 2009; Qu et al., 2012). However, Qu et al. (2012) found 7 d UV exposed nC_{60} had negligible surface sorption of either HWM or LMW HAs due to the negative surface charge and elevated surface hydrophilicity. Hence, DOM is likely to be less significant in determining the suspension stability of irradiated nC_{60} (Qu et al., 2012). A similar relationship may occur due to the formation of oxygen containing hydroxyl- and carboxyl-groups on MWCNTs due to surface oxidation, which can promote colloidal stability and hydrophilicity of CNTs in addition to inducing alterations to surface charge (Shieh et al., 2007; Smith et al., 2009). This is illustrated by Hu et al. (2005), in which carboxylic acid groups as a result of nitric acid treatment of SWCNTs had high ζ -potentials (-28

277 mV) over a pH range of 2-10, indicating their moderate stability in water in contrast to pristine
278 CNTs (Hu et al., 2005). Reduced deposition of pristine nC_{60} occurred on silica glass beads coated in
279 HMW HA than LMW HA due to steric hindrance (Qu et al., 2012). The effect was more
280 pronounced at lower ionic strengths due to electrostatic repulsion between charge groups
281 resulting in a more stretched-out conformation of HA molecules (Qu et al., 2012). However, both
282 HMW and LMW HA coated beads facilitated the deposition of 7 d UV exposed nC_{60} , with reduced
283 sensitivity to changes in ionic strength as a result of reduced steric hindrance (due to the compact
284 conformation of HAs at high ionic strengths (~ 60 mM)), lower surface potential and increased
285 hydrogen bonding between the oxygen containing groups of the functionalised nC_{60} and nitrogen-
286 and oxygen-groups on the HA (Qu et al., 2012).

287 Comparatively determining the relative importance of CNM functionalization and ionic
288 strength on CNTs and nC_{60} behaviour in soils is difficult due to the myriad of different
289 experimental configurations. To overcome this, Jaisi and Elimelech (2009) used carboxyl-
290 functionalised SWCNTs and nC_{60} (radius of 51 nm) in natural soil columns containing 29% clay and
291 pore sizes of 22 μ m, to determine the impact of ionic strength on particle transport and
292 deposition. As ionic strength increased (0.03 – 100 mM), the rate of SWCNT deposition within the
293 soil column also increased, with the observed effect more apparent with divalent cations (Ca^{2+})
294 than monovalent cations (K^+). Interestingly, while nC_{60} was highly sensitive to variations in ionic
295 strength, far lower deposition rates were observed. It was proposed that the structure and shape
296 of SWCNTs, in particular their large aspect ratio and bulky agglomerated states, in addition to soil
297 particle heterogeneity increased the straining effect and retention by the soil matrix (Jaisi and
298 Elimelech, 2009). Nanomaterial structural conformation is therefore a further consideration in the
299 relative extent to which CNMs will be dispersed and transported within the soil, with nC_{60} more
300 likely to experience leaching than SWCNTs under a variety of solution chemistries (Jaisi and
301 Elimelech, 2009) within negatively charged soil media.

Functionalization status is therefore a fundamental consideration to the behaviour of CNMs, resulting in distinct characteristics, which significantly modify behaviour in relation to their unfunctionalised counterparts. However, key questions as to the behaviour of CNMs within the environment remain unaddressed; for example, how does the repeated exposure of CNMs to weathering cycles within the soil influence their fate and behaviour?

4. CNM-contaminant sorption, desorption and mobility in soils

The ability of natural colloids to assist in the transport of organic contaminants has been well documented and reviewed (de Jonge et al., 2004; Sen and Khilar, 2006; Li et al., 2013). Typically, hydrophobic compounds such as PCBs and PAHs have limited environmental mobility due to strong sorption to SOM. Kan and Tomson (1990), however, demonstrated that high concentrations of colloidal materials such as DOM may enhance the transport of hydrophobic compounds such as phenanthrene and naphthalene by a factor of a thousand or greater, with possible implications for the spread of contamination and groundwater quality (de Jonge et al., 2004). Although CNMs may be tailored to suit specific requirements, their behaviour is not necessarily different to colloids naturally occurring in the environment (Colvin, 2003; Lead and Wilkinson, 2006). To determine the relevance of natural nanoparticle facilitated transport of contaminants in porous media such as soils, Kretzschmar et al. (1999) identified four key factors that will be used as a framework for this section:

- 1) Sufficiently high concentration of nanoparticles
- 2) Mobility of the nanoparticles carrying sorbed HOCs
- 3) Sorbate toxicity even when present in trace quantities
- 4) The ratio of sorption to desorption relative to the timescale of particle mobility

The sorption affinity of CNMs for common environmental contaminants such as PAHs, known to pose significant risks to both the environment and human health due to their toxic properties

(Menzie et al., 1992; Shaw and Connell, 1994; Cebulska-Wasilewska et al., 2007), has been reported as over three orders of magnitude greater than that of natural soil/sediments (Yang et al., 2006b). The potential for these emerging materials to become widespread in the soil environment, particularly those with a strongly hydrophobic nature and large reactive surface area, such CNMs, raises questions and concerns about the environmental consequences of their release (Pan and Xing, 2010).

4.1 CNM contaminant sorption and desorption in soils

Understanding the adsorption and desorption of HOCs to CNMs in soils is critical to the environmental risk assessment processes, as well as determining their potential applications as environmental adsorbents (Yang et al., 2006a). As the fundamentals of CNM-HOC sorption have been extensively reviewed, the reader is referred to a review by Ren et al. (2011) for a comprehensive overview. This section addresses the manner in which soils may alter the HOC sorption/desorption properties of CNMs, focusing specifically on two conflicting effects; (i) CNM dispersal by DOM (increasing the surface area and hence the number of adsorption sites (Hyung et al., 2006; Lin and Xing, 2008)); versus (ii) the formation of CNM-DOM coatings (blocking and/or competing for adsorption sites reducing the number available for organic contaminants (Chen et al., 2008; Wang and Keller, 2009; Cui et al., 2011; Wang et al., 2011; Zhang et al., 2011c)). The relative importance of these two phenomena are poorly understood in relation to their sorption and desorption of organic contaminants (Zhang et al., 2011c; Pan and Xing, 2012), and are highly dependent on the nanoparticle properties, and the nature of SOM and the sorbate (Wang et al., 2009; Zhang et al., 2011c; Lerman et al., 2013).

In assessing the impact of OM on CNM sorption in the environment, further complications arise as contaminants are able to sorb to both the CNM and CNM-OM coating (Wang et al., 2008b). Hyung and Kim (2008) identified SOM adsorption to nanotubes was highly variable depending on the type of SOM, occurring proportional to its aromatic carbon content. This has implications for determining the ability of CNMs to sorb organic compounds, yet most

investigations fail to consider the role of different OM fractions in CNT-pollutant interactions (Lerman et al., 2013).

Wang et al. (2008b) assessed the extent to which HAs and peptone altered the sorption of phenanthrene, naphthalene or 1-naphthol on-to MWCNTs (outer diameter of 40 nm), by fitting sorption data with Freundlich and Polanyi models. Their results showed that each type of DOM resulted in nonlinear sorption isotherms to the MWCNTs, following the order peptone > HAs. Although the inherent sorptive ability of HA for each of the contaminants was more limited than that of pristine MWCNTs, HA coatings did not result in large changes to the sorption of any of the contaminants, which is inconsistent with models indicating that “fouling” of sorption sites by DOM will reduce contaminant sorption capacity (Carter and Weber, 1994; Wang et al., 2008b). The authors proposed that either uneven coating of the MWCNT by HA occurred, or that the anticipated sorption reduction as a result of polar moieties from the HA coating, was offset due to the increase in O-containing moieties resulting in particle repulsion and dispersal, exposing new sites available for HOC sorption. Contrastingly, peptone, due to the large quantity sorbed relative to each of the other DOM fractions, resulted in the largest reduction in available sorption sites (Wang et al., 2008b). Similar interactions as to the relative ability of different OM coatings to alter CNM sorption have been identified by Cui et al. (2011) and Gai et al. (2011). Although direct comparison of the studies is not possible due to the different particles used, Cui et al. (2011) found HAs, TAs and peptone pre-interacted with SWCNTs resulting in the formation of polar functional groups on the nanotube surface, reducing the area available for phenanthrene sorption in the order of peptone > TAs > HAs. Similarly, Gai et al. (2011) identified a reduction in C₆₀ agglomerate sizes due to the dispersal effects of HA, increasing atrazine sorption due to dispersal and re-arrangement, rather than interactions between the atrazine and HA (Gai et al., 2011). Hence, it may be argued that the impact of DOM on CNM-HOC sorption is dependent on the type of OM present and possibly also CNM type.

378 Within a soil environment, Li (2012), identified the sorption behaviour of naphthalene,
379 phenanthrene and fluorine in a sandy loam soil, silt loam soil and Ottawa sand was unaffected
380 following amendment of MWCNTs at concentrations of 2 mg g⁻¹. For each contaminant
381 investigated, sorption in both the MWCNT amended and unamended samples followed the same
382 order; silt loam > sandy loam > sand, indicating the sorptive ability was driven by the organic
383 carbon content (Li, 2012). Additionally, no difference between sorption isotherms of MWCNT
384 amended and unamended samples was apparent, indicating that MWCNTs held no influence over
385 the samples inherent sorption capacity (Li, 2012). Similarly, the order in which PAHs sorbed was
386 unaffected by the MWCNTs, occurring as anticipated according to the PAHs K_{ow} values (Li, 2012).
387 After 24 h hydroxypropyl- β -cyclodextrin (HPCD) desorption, no statistically significant differences
388 in the percentage desorbed were detected between nanotube amended and unamended
389 samples. Hence, the sorptive properties of MWCNTs in the environment may be similar to hard
390 carbon, and did not influence the sorption/desorption behaviour of PAHs (Li, 2012). When this is
391 related to Factor 4 proposed by Kretzschmar et al. (1999), in which the sorption to desorption
392 ratio over timescale of particle transport is considered, MWCNTs at 2 mg kg⁻¹ concentration may
393 not be considered significant in determining the behaviour of some PAHs in soils over the
394 duration of their experiment. It may be, however, that Factor 1 was not met, and the lack of
395 MWCNT-influence on PAH behaviour was merely a result of an insufficient quantity added to the
396 soils. Hence, while this study indicates the sorption of multiple contaminants in the different soil
397 types considered will not present an environmental concern in terms of pollutant mobilisation at
398 2 mg kg⁻¹, it was not possible to consider the desorption of contaminants if transport were to
399 occur.

400 An excellent study by Towell et al. (2011), assessed the extent to which HPCD extraction of
401 HOCs with different physicochemical properties desorbed from soils amended with CNMs at
402 concentrations between 0.05% and 0.5% (substantially larger than that employed by Li (2012)). At
403 concentrations $\geq 0.05\%$, Towell et al. (2011) identified significantly less ¹⁴C-B[a]P extracted from

CNM amended soils than ^{14}C -phenanthrene due to the high hydrophobicity and $\log K_{ow}$ value reducing the ability of ^{14}C -B[a]P to partition into the aqueous phase. This was exacerbated by the relatively HMW of ^{14}C -B[a]P, which has been proven as a critical factor determining the bonding energy between SWCNTs and PAHs (Debnath et al., 2008; Towell et al., 2011). In relation to Factor 4, the nature of the sorbate may therefore influence the extent of desorption, and therefore the duration for which contaminants will remain sorbed. It may be proposed that CNM sorbed HMW HOCs represent a greater risk of increased distance of transportation within the environment than LMW HOCs.

While sorption of HOCs to CNMs in soils can occur, the extent of sorption and desorption is dependent on the type of OM and concentration of CNMs. With a view to the manner in which the properties outlined above potentially facilitate transportation of contaminants sorbed to CNMs in soils, studies in which mobility has been directly investigated will also be discussed.

4.2 CNM-HOC mobility

Once sorbed to freely suspended CNMs within the soil matrix, the mobility of HOCs is potentially increased; however, very few studies have focused on determining the impact of CNMs on contaminant movement in soils. An overview of the basic principal of CNM facilitated HOC transport is presented in Figure 1. Using column leach tests, Li (2012) examined the behaviour of phenanthrene, fluorine, naphthalene and pyrene in a saturated sandy loam soil amended with MWCNTs, functionalised MWCNTs (f-MWCNTs) and functionalised SWCNTs (f-SWCNTs) at a concentration of 5 mg kg^{-1} . Significant retention of PAHs within the soil column was observed, due to the strong sorption of contaminants by CNTs and their limited mobility within the soil column (Li, 2012). In control soils and those amended with MWCNTs and f-MWCNTs, retention of PAHs occurred in the order naphthalene < fluorine < phenanthrene < pyrene, with hydrophobic interactions between the CNTs and PAHs cited as the predominant cause of the observed pattern (Li, 2012). Contrastingly, retention of contaminants within soils amended with f-SWCNTs occurred in the order of naphthalene > fluorine > phenanthrene > pyrene, the sorption of

which could not be accounted for by hydrophobic forces alone (Li, 2012). The trend was negatively correlated to molecular size, indicating that larger sorbate molecules may have less space for sorption due to the additional hydrophilic functional group (Yang et al., 2006b; Li, 2012). It was concluded that leaching behaviours were determined by physical characteristics of both CNTs and contaminants (Yang et al., 2006b; Li, 2012).

To determine the extent to which CNMs facilitated the movement of contaminants relative to various types of DOM, Zhang et al. (2011b) used saturated, sandy soil columns contaminated with either PCBs or phenanthrene to comparatively assess the mobilising ability of nC_{60} at 1.55 – 12.8 $mg\ l^{-1}$ relative to DOM at 10–11 $mg\ l^{-1}$. The results showed that even the lowest concentration of nC_{60} significantly enhanced the dispersal of both PCB, and phenanthrene; whereas, columns containing only various types of DOM had no effect on contaminant transport (Zhang et al., 2011b). The enhanced contaminant mobilisation ability of nC_{60} relative to naturally occurring DOM was attributed to its unique porous structure and surface enthalpies of interaction, which generate a large sorption affinity together with an irreversibly or slowly desorbable fraction of adsorbed phenanthrene/PCBs (Hofmann and von der Kammer, 2009; Zhang et al., 2011b; Wang et al., 2012a). CNMs may therefore be much more efficient at enhancing the mobility of contaminants than natural colloidal materials.

Different processes of nC_{60} formation have also been identified as contributing to large differences in their ability to alter the fate and transport of contaminants (Wang et al., 2012b). Wang et al. (2012b) assessed nC_{60} samples prepared using either the standard solvent exchange method, eight different types of SOM or surfactant modifications, or by the phase-transferring of nC_{60} from a solution of toluene to either SOM or a surfactant (Wang et al., 2012b). Their results indicated that while the mobility of nC_{60} was similar between each of the preparation methods, the contaminant mobilising capability significantly differed. Relative to the unmodified nC_{60} , transport of PCBs through a saturated column of sandy soil increased by 42.2 – 227% with surfactant modified nC_{60} , and by 233–370% with SOM-modified samples (Wang et al., 2012b). The

results were attributed to both increased adsorption affinities together with enhanced resistance to desorption due to alterations to nC_{60} aggregation properties as a result of the SOM surfactant (Wang et al., 2012b). During the process of aggregate formation, it is possible that a fraction of SOM or another surfactant was intercalated within the C_{60} aggregates, significantly influencing the porosity and geometry of the resulting nC_{60} aggregates, contributing to the enhanced desorption resistance of PCBs (Wang et al., 2012b). With differing types of SOM and surfactants, variations in the quantity and geometry of pores will occur, with the possibility that nC_{60} could be tailored to specific physicochemical properties for use in *in situ* site remediation (Wang et al., 2012b). Hence, the adsorption, desorption and transport of contaminants by nC_{60} will vary greatly depending on the condition of its formation (Wang et al., 2012a).

Using a different approach, Hofmann and von der Kammer (2009) computer modelled the extent to which CNMs could result in the movement of HOCs in soils under various scenario-based conditions, to determine when relevant CNM transport of sorbed HOCs might occur. Worst-case scenarios were adopted, assuming fully mobile CNMs within the porous medium, over a range of realistic yet high CNM concentrations ($100 \text{ mg L}^{-1} - 1 \text{ g}^{-1}$) occurring in aggregate sizes of 10 – 100 mm. It was also assumed that CNMs were pre-equilibrated with the HOC at source and that diffusion was the rate-limiting step for desorption (Hofmann and von der Kammer, 2009). From this, it was possible to estimate the fraction of contaminants bound to CNMs at different distances from the source over different time periods (days to years) using the Streamtube Model for Advective and Reactive Transport (SMART) (Finkel et al., 1998), combined with the application of retarded pore diffusion approximations (Bold et al., 2003) and combinations of two first-order rate expressions (Cornelissen et al., 1997). The results showed that for aggregates of 100 mm, 2, 7, 40, 75, 82% of bound contaminants were transported > 1 m at nanoparticle concentrations of 0.1, 1, 10, 100, 1000 mg l^{-1} , respectively. Conversely, modelled transportation of contaminants sorbed to aggregates of 10 mm in size were reduced to 0.1, 0.5, 3.6, 8% for the same respective

concentrations. Breakthrough of the 1 m modelled column did not occur in any of the considered scenarios and all contaminants remained bound to the nanoparticle.

Parameterisation of the ratio of desorption to sorption and particle transportation is achieved by the Damköhler number (Da) (Equation 1) (Jennings and Kirkner, 1984), and was employed to simulate the HOC desorption from CNMs.

Equation 1

$$Da_{NP} = \lambda s$$

Where Da_{NP} = Damköhler number for the NP, λ = rate constants (first order) for the reaction in s^{-1} , s = average residence time within the system, which may also be expressed as average flow velocity (Hofmann and von der Kammer, 2009). A ratio of 0 identifies equilibrium between particle-contaminant interactions, whereas 1 indicates a decoupled transport (i.e. the HOC in solution will be transported independent of contaminants sorbed to the CNM) and < 0.01 indicates fully decoupled transport. In these instances, HOCs sorbed to CNMs will not desorb over the transportation time frame, rather relocate within the media (Hofmann and von der Kammer, 2009). At Da numbers > 100 , an equilibrium exists between the immobile porous media and mobile CNM (Kretzschmar et al., 1999; Bold et al., 2003), resulting in limited nanoparticle relocation of the contaminant (Hofmann and von der Kammer, 2009).

Hofmann and von der Kammer (2009) calculated Damköhler numbers for CNM aggregates of different sizes and partitioning coefficients according to the rate constant data shown in Figure 2, and based on different flow velocities of 1 m in 50 d (fast flow) – 1 m in 10 y (slow movement). It was inferred that the CNM-contaminant transport mechanisms are strongly dependent on the size of CNM agglomerates together with the distribution coefficients ($\log K_d$) (Hofmann and von der Kammer, 2009). For example, Figure 2 shows contaminants sorbed to 1 mm aggregates at a flow velocity of 1 m 50 d⁻¹ will not experience contaminant desorption until the $\log K_d$ of HOC-CNMs is 8 m³ kg⁻¹, reaching equilibrium at $\log K_d$ 1 m³ kg⁻¹ (Hofmann and von der Kammer, 2009).

506 However, at a flow velocity of 1 m y^{-1} , decoupled transport will predominate for 1 mm aggregates
507 at a $\log K_d$ of $9 \text{ m}^3 \text{ kg}^{-1}$ reaching equilibrium at $\log K_d$ of $2 \text{ m}^3 \text{ kg}^{-1}$ (Hofmann and von der Kammer,
508 2009). It was concluded that under equilibrium sorption/desorption conditions, CNM mobility
509 resulted in negligible transport of sorbed contaminants (Hofmann and von der Kammer, 2009).
510 However, the mobility and concentration of CNMs becomes increasingly important in instances
511 with slow to very slow desorption (Hofmann and von der Kammer, 2009). While there are many
512 assumptions and simplifications associated with every modelling technique, the model identifies
513 scenarios in which transport and desorption of sorbed contaminant could potentially occur,
514 possibly providing useful guidelines for risk-assessment if applied on a case by case basis.
515 However, further work aimed specifically at validating the model against traditional column leach
516 tests in both V-C and P-C soils and additional desorption kinetics are urgent prerequisites.

517 From the above discussion, it can be concluded that each of the four factors identified by
518 Kretzschmar et al. (1999) for significant transport of contaminants by CNMs have been met.
519 However, more work examining the subsurface transport of CNMs through well-defined soils of
520 various types (such as clays, peats and silts) and CNMs with a variety of functional groups, sizes
521 and sorbed compounds in both saturated and unsaturated conditions are required (Jaisi and
522 Elimelech, 2009; Petersen et al., 2011). Of studies that are available, variation in experimental
523 conditions between the investigations renders comparisons of the efficiency of contaminant
524 mobility between CNM types tentative until standardised comparative testing is conducted.
525 Additionally, the molecular weights and sizes of CNMs may not be constant during their transport
526 within the soil environment, due to their physical, chemical or biological interaction with soil
527 components, which will likely influence their aggregation status, shape, surface charge (Pan and
528 Xing, 2012), and possibly also their ability to sorb and mobilise contaminants over long timescales.
529 Furthermore, definitive data of the desorption kinetics of HOCs from CNMs in soils are essential to
530 understanding their ability to transport contaminants (Ibaraki and Sudicky, 1995; Choi and Yavuz
531 Corapcioglu, 1997; Corapcioglu et al., 1999; Bold et al., 2003; Hofmann and von der Kammer,

2009), with slow desorption identified as a critical requirement (Roy and Dzombak, 1998). The lack of experimentally derived desorption kinetic data from a range of soil types and conditions makes determining the extent to which HOC sorption is strong enough and desorption slow enough to allow CNMs to transport sorbed HOCs and the implications challenging (Qu et al., 2012).

5. CNM – microorganism interactions

As soils represent one of the ultimate sinks for nanomaterials (Nowack and Bucheli, 2007), terrestrial microorganisms which interact directly with soils may be significantly affected (Navarro et al., 2008). However, understanding the impact of CNMs on the soil microbial community is a subject still in its infancy (Dinesh et al., 2012). If CNMs within soils are bioaccessible to microflora, the possibility of disruption to bio-geochemical processes within soils may be increased (Neal, 2008). This section discusses recent literature related to toxicity of CNMs in soils, modification of CNM fate and behaviour by microbiota, and the possible implications for the biodegradation of contaminants.

5.1 CNM toxicity in soils

The toxicity of CNMs is dependent upon the bioaccessibility of nanoparticles to bacteria, and retention of some the nanoparticles reactivity (Neal, 2008). Currently, little literature is available related to the toxicity of CNMs within soils (Dinesh et al., 2012). Hence, the discussion presented here provides a theoretical estimation of the specific microbial communities that may be more vulnerable to soil-borne CNMs, followed by an overview of recent CNM-amended soil toxicity findings published within the literature.

Soil conditions will ultimately dictate the extent to which CNMs are able to interact with terrestrial microflora. Based on the discussion earlier relating to the fate and behaviour of CNMs in soils, in addition to information regarding cell properties (Mehmannavaz et al., 2001), it may be possible to tentatively speculate as to the bioavailability or bioaccessibility of CNMs to different microbial populations. When assessing nanotoxicity, consideration must be given to both the likelihood of a nanoparticle coming into contact with microbial cells together with the initial concentration added to soils, to provide an accurate means of estimating the particle availability (Dinesh et al., 2012). It can be considered that a strong interplay exists between the dispersal status of nanoparticles and their bioaccessibility to specific soil microbial populations (Turco et al., 2011). As bacteria frequently adhere to surfaces in the soil environment, attached cells within biofilms constitute a large proportion of the bacterial community in the subsurface environment (Neal, 2008). Neal (2008) therefore proposed that the study of nanotoxicity towards biofilm communities is a more appropriate measure of toxicity in environmental systems than planktonic cells. However, it is conceivable that given appropriate DLVO and non-DLVO forces between CNMs, microorganisms and the soil matrix, CNMs could also become available to planktonic cells. One example of which may be that CNM-SOM coatings could result in easier access to the cell surface relative to uncoated particles due to the similarities in solubility between the cell membrane and surfactant; however, the coating itself may attenuate the toxicity due to a lack of physical contact between the CNM and a microbial cell (Lubick, 2008). Further work into the conditions under which CNMs will be available to different microbial communities in soils is needed.

The extent to which soils with different properties determine the toxicity of some CNMs was directly investigated by Chung et al. (2011). The impact of MWCNTs at 50, 500 and 5000 μg^{-1} soil on the activity of soil microorganisms in a sandy loam (pH 6.98, OC content 17.69 g kg^{-1} , CEC 13.51 ± 0.78) and loamy sand (pH 5.21, OC content 8.33 g kg^{-1} , CEC 9.05 ± 0.10) was considered. Based on an analysis of the activity of enzymes involved with cycling carbon (1,4- β -glucosidase,

cellobiohydrolase, xylosidase), nitrogen (1,4- β -acetylglucosaminidase) and phosphatase together with lower microbial biomass-nitrogen and carbon in soil, their results indicated that MWCNTs exhibited antimicrobial properties within both soil types (Chung et al., 2011). As these findings are consistent with culture studies outside of the soil environment, in which reduced microbial activity was a result of membrane damage, physical piercing and oxidative stress (Kang et al., 2007; Simon-Deckers et al., 2009), the authors assumed similar antimicrobial mechanisms of action may be responsible. However, there is a strong argument against basing assumptions of nanotoxicity mechanisms occurring in soils on those known to occur in culture studies, due to typically large variations in test conditions.

Other investigations of nanotoxicity within soil using nC_{60} have found more limited toxicity effects. For example, Johansen et al. (2008) found microbial respiration and biomass to be unaffected by pristine C_{60} agglomerates (50 nm – 50 μ m-size) applied at concentrations of 0, 5, 25 and 50 mg kg⁻¹ to dry, clay loam textured soil containing 2.5% OM and with a pH of 6.7. However, polymerase chain reaction-denaturing gradient gel electrophoresis (PCR-DGGE) measurements of the diversity and number of bacteria over a 14 d period showed that a three to four fold reduction in rapidly growing bacteria occurred immediately following the addition of C_{60} (Johansen et al., 2008). The authors proposed the results may have been observed as a direct consequence of reactive oxygen species (ROS) formed by the C_{60} , which disrupted DNA and lipids within membranes (Johansen et al., 2008). However, confirmation of ROS damage could not be acquired due to the complexity of the soil environment (Dinesh et al., 2012), and a recent publication by Chae et al. (2012) casts some doubt on the extent to which ROS are generated in the presence of SOM. It may therefore be considered more likely that the observed alterations to the diversity and number of bacteria may be an indirect result of a reduction in nutrient bioavailability due to adsorption by C_{60} (Johansen et al., 2008).

In a similar investigation, Tong et al. (2007) assessed the role aggregation status plays in determining nanotoxicity within soils. The impact of either nC_{60} at 1 μ g C_{60} g⁻¹ soil, or 1000 μ g C_{60} g⁻¹

¹ soil in granular form on the function and structure of soil microbial community was assessed (Tong et al., 2007). The silty clay loam soil (pH of 6.9, OM content 4%) was incubated with each of the nanoparticle treatments for 180 d (Tong et al., 2007). Both C₆₀ and nC₆₀ resulted in limited alteration to either the function or structure of microbial processes or communities (Tong et al., 2007). These findings are similar to those of Johansen et al. (2008), and consistent with other investigations in which the bioavailability and antibacterial activity of nC₆₀ reportedly diminished following sorption to soil, with the overall sorption capacity dictated by the soil OM content (Li et al., 2008).

Despite differences in experimental setups between the studies by Johansen et al. (2008) and Tong et al. (2007), from the data presented, it is not possible to rule out the bioaccessibility and toxicity of C₆₀ to a proportion of microbiota within soils. Although alterations to microbial respiration as a result of fullerene addition to soil were not observed in either study, functional substitution of specific impaired microorganisms may have occurred, masking any apparent variation (Ekelund et al., 2003; Johansen et al., 2008). The studies presented here provide credible insight into the possible toxicity of CNMs within the environment; however, insufficient data comparatively analysing the impact of all CNMs on microbial populations within a range of well-defined soil types is a major obstacle in determining their potential environmental impact. Experimental work aimed at addressing the real world implications for particle toxicity to different microbial communities, systematically testing the factors determining the behaviour and fate of CNMs in soils highlighted previously, is required before firm conclusions can be drawn regarding the impact of CNMs on soil microbial activity and structure. Specifically, the implications of abiotic alterations and methods of CNM preparation on the bioaccessibility and toxicity to soil microbiota have received little investigation.

5.2 *Biological modification of CNMs in soils*

The influence of microbial populations on the physical and chemical state of nanoparticles must also be considered (Aruguete and Hochella, 2010). Degradation of C₆₀ through

photochemical processes have been identified by numerous investigations (Hou and Jafvert, 2008; Lee et al., 2009; Li et al., 2009), which may be an important step in both its breakdown and the activation of precursors for subsequent biological interactions (Turco et al., 2011). A reduction in the size of nC_{60} aggregates and alterations to surface chemistry and solubility have been observed following exposure to light (Turco et al., 2011). This suggests that release of C_{60} into soils could possibly result in the formation of photochemical products and changes to C_{60} molecular structure (Turco et al., 2011). While C_{60} photochemical reactions at the soil surface have not been studied, its oxidation and transformation to fullerenol has been observed in water and in the presence of oxygen (Turco et al., 2011). Following the abiotic photochemical modification of C_{60} through sunlight into fullerols (i.e. $C_{60}-OH$), white-rot fungi was able to attack and subsequently incorporate a small amount of fullerol carbon into fungal lipids after 32 weeks of decay (Schreiner et al., 2009). By contrast, unmodified C_{60} was recalcitrant to such attack (Schreiner et al., 2009); hence, following minor surface alterations, biological interactions with C_{60} were substantially altered, changing the fate of the particle. Similarly, the potential for horseradish peroxidase to biodegrade CNTs is strongly related to the presence of carboxyl groups on the nanotubes surface, which permitted enzyme mediated oxidation relative to pristine CNTs (Allen et al., 2008; Allen et al., 2009). Furthermore, Fenton's reagents oxidised carboxyl-functionalised SWCNTs (SWCNT-COOH) through the formation of hydroxyl radicals (Allen et al., 2008; Allen et al., 2009). It has therefore been suggested that both white and brown rot mediated fungal activity could modify surface functionalised CNTs in a similar manner to fullerols (Turco et al., 2011).

Ultimately, Turco et al. (2011) suggested that the fate of C_{60} in soil is potentially controlled by the rate of abiotic alterations to the formation of more reactive precursors, as opposed to a simple dose response, and the toxicity of UV-modified CNMs in soils has not yet been investigated. If degradation of CNMs by fungi were to occur on a large scale in the natural environment, their potential environmental risk would be significantly reduced; however, it is

unlikely to occur in sufficiently large quantities to efficiently reduce the burden of CNM presence in soils.

5.3 *The bioavailability and bioaccessibility of CNM associated contaminants*

Sorption of contaminants is a fundamental mechanism in the regulation of organic compound bioavailability (Lou et al., 2011). Given their strong sorptive capability, the addition of CNMs to soil may result in the sequestration of organic contaminants, reducing their extractability and bioaccessibility, operating in a similar manner to hard or black carbon (Chen et al., 2007). However, the extent to which the processes identified in Section 4.1 impact upon the bioaccessibility of contaminants and biodegradation have not received much research within soils.

The conditions under which CNMs enter the soil are also critical to determining their impact upon contaminant bioaccessibility. Zhou et al. (2013) incubated ^{14}C -2,4-dichlorophenol (^{14}C -2, 4-DCP) in a soil containing either 0, 2, 20 or 2000 mg kg⁻¹ SWCNTs or MWCNTs to determine the impact of carbon nanomaterials on the mineralisation, degradation and distribution of ^{14}C -2-4-DCP in the soil. The impact of the order in which the nanomaterials were added to the soil was also assessed, with nanomaterials added either after spiking, simulating disturbance of CNTs on pre-existing contamination in soils, or as a mixture with ^{14}C -2, 4-DCP, simulating HOC degradation when carried or accumulated (concentrated) by CNMs within the environment from other sources. CNTs added at concentrations ≤ 20 mg kg⁻¹ after spiking resulted in no significant effects on the time course of mineralisation, indicating that the activity of microorganisms was not significantly influenced, nor did the desorption from CNTs reduce ^{14}C -2, 4-DCP bioavailability in soil (Zhou et al., 2013). However, following the addition of CNTs at 2000 mg kg⁻¹ added after spiking, mineralisation of ^{14}C -2, 4-DCP was significantly ($P < 0.05$) inhibited, which was attributed to a reduction in the aqueous phase concentration of ^{14}C -2, 4-DCP in soil solution by 1/5 and 1/12 for SWCNTs and MWCNTs, respectively (Zhou et al., 2013). CNT interactions with contaminants

683 within the soil environment due to aggregation therefore reduced available sorption sites, as well
684 as aggregate interaction with soil components such as humic substances, DOM, peptone and TA,
685 which potentially coat CNTs modifying surface polarity, reducing surface area and hence reducing
686 HOC sorption capacity as discussed in Section 4.1 (Wang et al., 2008b; Cui et al., 2011; Zhou et al.,
687 2013). As the adsorption of ^{14}C -2, 4-DCP to CNTs was reversible, the bioaccessibility of 2, 4-DCP
688 was not reduced; however, indigenous microorganisms were not able to mineralise desorbed ^{14}C -
689 2, 4-DCP at the same rate of desorption due to the effects of CNTs on microbial activity (Zhou et
690 al., 2013). CNTs are therefore potentially able to increase the persistence of organic pollutants
691 within soil through reducing biodegradation, with greater effects observed for pre-adsorbed
692 contaminants (Zhou et al., 2013).

693 Similar results were obtained by Cui et al. (2011), who assessed the bioavailability of
694 phenanthrene to microorganisms in sediments amended with either SWCNTs, biochar or
695 charcoal, with mineralisation inhibited to the greatest extent by SWCNTs. It was proposed that
696 the larger surface area and pore volume of SWCNTs relative to the other sorbents was responsible
697 for the observed results. However, following the coating of SWCNTs with either HAs, TAs or
698 peptone, a reduction in phenanthrene sorption occurred due to reduced pore volumes and
699 surface area, ultimately also reducing the extent to which sorption to SWCNTs reduced
700 mineralisation (Cui et al., 2011).

701 In addition to the impact of soil types on the impact of CNMs on organic contaminant
702 sorption, properties of the organic chemicals within soils are also influential in dictating their
703 interaction with different types of CNMs. Towell et al. (2011) assessed the impact of fullerene
704 soot (FS), SWCNTs and MWCNTs at 0, 0.05, 0.1 and 0.5% concentrations, on the HPCD
705 extractability (proven as an indicator of PAH bioaccessibility to soil micro flora (Reid et al., 2000;
706 Doick et al., 2005; Stokes et al., 2005; Rhodes et al., 2008b) and mineralisation of ^{14}C -
707 phenanthrene, and HPCD extractability of ^{14}C -benzo[a]pyrene (^{14}C -B[a]P) in soils over an 80 d
708 period. At concentrations $\geq 0.05\%$ CNMs, ^{14}C -phenanthrene mineralisation was significantly

inhibite



suggesting enhanced PAH sorption reduced the aqueous substrate available for microbial mineralisation (Towell et al., 2011). Differences were also apparent between CNM types, with SWCNTs generally resulting in greater mineralisation inhibition in relation to MWCNTs and FS (Towell et al., 2011). However, at a concentration of 0.5% CNMs, ^{14}C -phenanthrene was mineralised to a greater extent with SWCNT amendments than FS. This disparity was attributed to variation in rates of phenanthrene desorption from the solid to aqueous phase, as desorption hysteresis occurs more commonly with fullerenes than CNTs due to differences in aggregate structure and availability of sorption sites (Cheng et al., 2005; Yang and Xing, 2007; Towell et al., 2011). The HPCD extractability of ^{14}C -phenanthrene was significantly reduced as a result of CNM amendment in a concentration dependant manner due to increased numbers of sorption sites resulting in enhanced phenanthrene sorption (Towell et al., 2011). However, while the HPCD extractability of ^{14}C -B[a]P reduced with increasing concentrations of SWCNTs and MWCNTs, no significant concentration dependant differences were observed with FS (Towell et al., 2011). The ability of CNMs to sorb and hence modify the bioaccessibility of HOCs is therefore dependent on the differences in physicochemical properties of the PAH in relation to the properties of the CNM.

When considering the fraction of contaminants sorbed to CNMs within these investigations, and the resulting reduced bioavailability, two schools of thought may be adopted; (i) over time the non-degradable, bound fraction may innocuously degrade (Gevao et al., 2000a), or (ii) the bound fraction is potentially re-mobilised over long time scales with potential environmental implications (Gevao et al., 2000b). This draws on the discussion by Semple et al. (2013), in which the significance of distinguishing between bioavailability and bioaccessibility is significant, particularly when dealing with environmental 'super sorbents' such as CNMs with reference to remediation of contaminated land and risk assessment. Semple et al. (2004) defined bioavailability as 'that which is freely available to cross an organism's cellular membrane from the medium the organism inhabits at a given time', and is considered as a rate of substrate delivery to cells. While bioaccessibility encompasses this fraction, it additionally extends to those which are

potentially available over time, but are currently chemically or physically removed from the microorganism (Semple et al., 2004). In other words, it provides a definition of the total extent of substrate that will be available to cells. Arguably, bioaccessibility is of relatively greater importance when considering the fate and behaviour of CNM sorbed contaminants, due to the larger temporal range and lack of implied immediacy. However, under some environmental conditions, microbial colonisation of CNM agglomerates can occur, with potential implications for the bioaccessibility of the bound contaminant fraction.

5.4 Microbial sorption and biofilm formation

While the toxicity of CNMs in soil is dependent on their bioaccessibility in addition to retention of reactivity, if agglomerates of CNMs are present with a reduced cytotoxic nature, it is conceivable that interstitial gaps in the agglomerate with mesopore dimensions will result in their increased suitability for the sorption of microorganisms (Agnihotri et al., 2005; Upadhyayula and Gadhamshetty, 2010). When this is related to the previous discussion of CNM contaminant sorption and the implications for biodegradation, it is possible to re-consider the lack of bioaccessibility of CNM sorbed contaminants reported in some studies, and consider their potential to increase contaminant bioaccessibility in certain situations. Properties of particular importance to when considering CNMs for such applications include: (i) structures with high porosities readily colonisable by microorganisms; (ii) potential ability to encourage biofilm formation through offering a buffering capacity and (iii) the ability to adsorb high concentrations of contaminants from bulk solution yet regulate the microbial biodegradation through desorption (Abu-Salah et al., 1996).

Biofilms are groups of well-organised, adjoining cells encapsulated within a matrix of insoluble, extracellular polymeric substances (EPS) (Morikawa, 2006). EPS encapsulation supports cell substance and growth through the trapping, binding and dissemination of external nutrients by charged polysaccharide groups (Cheng et al., 2007), and offers greater protection against external stresses within the environment relative to those residing in a planktonic state (Pang et

al., 2005). Materials that allow a high degree of bacterial colonisation and possibly biofilm formation are potentially suited to facilitating biodegradation (Upadhyayula and Gadhamshetty, 2010), which is typically most effective when microorganisms are in biofilm state relative to planktonic, due to greater bioavailability, protection and adaptability to toxic conditions and hence more rapid pollutant degradation (Singh and Cameotra, 2004; Singh et al., 2006). Furthermore, bacterial colonisation may stabilise nanoparticle aggregates, as polysaccharides such as those generated by bacteria, have been observed to significantly increase the aggregation of C₆₀ fullerene, reducing particle mobility within the environment (Espinasse et al., 2007).

Upadhyayula and Gadhamshetty (2010) conducted hypothetical calculations to determine the quantity of cells that an agglomerate of CNTs could potentially sorb. The dimensions of a typical bacterium such as *Shewanella oneidensis* (*S. oneidensis*) are 2 µm in height with a radius of 0.5 µm, resulting in a surface area of 7.85 x10⁻¹² m². Assuming that 10% of the surface area of 0.1g CNTs added to media was available for bacterial sorption, the CNTs would be able to sorb 3.18 E+13 *S. oneidensis* cells (Upadhyayula and Gadhamshetty, 2010). Furthermore, Upadhyayula et al. (2009) confirmed that the adsorptive capacity of nanotubes for the bacterial strain *Bacillus subtilis* to be 37 times greater than the capacity of activated carbon; however, this may vary depending upon pore volumes and surface area, which are key determinants of immobilisation capacity (Upadhyayula and Gadhamshetty, 2010). Given these parameters, it is conceivable that biofilms could develop on CNM aggregates given sufficient pore volumes and diminished CNM reactivity.

When the potential for biofilm development on CNMs is considered in relation to their HOC sorptive ability and aggregation within soils, it has been suggested that CNMs may be useful for enhancing biodegradation of organic pollutants that cannot be easily concentrated. With CNM aggregates behaving as an organic chemical collector and accumulator, biofilm development on CNMs potentially increases the bioavailability/bioaccessibility of the contaminant (Yang et al., 2006b). Given adequate reversibility of organic compound adsorption and limited desorption hysteresis, sorption of bacterial cells to the surface of CNM aggregates may shorten the diffusion

distance, facilitating the utilisation of the sorbed organic compound by the bacteria. This is well illustrated by Yan et al. (2004), who studied the removal efficiency of microcystins (MCs) toxins from solution by *Ralstonia solanacearum* bacteria (Gram-negative cells which are able to readily coalesce on fibrous material) immobilised as a biofilm on a nontoxic form of CNTs. Their results showed that the removal efficiencies of MCs were 20% greater by CNT biological composites than either CNTs or bacteria alone (Yan et al., 2004). The findings were explained through absorption of large amounts of MCs and *R. solanacearum* by CNTs, even when the concentration of MCs was highly diluted in water, resulting in a concerted biodegradation reaction (Yan et al., 2004). In a similar investigation, Kanepalli and Donna (2006) used CNT-bacteria nanocomposites to assess the bioremediation of highly persistent trichloroethylene (TCE) in ground water. The study revealed that TCE instantly sorbed to bacteria-nanocomposites, which was later released to bacteria that were immobilised on the surface and metabolised.

Xia et al. (2013) studied the bioavailability and desorption (Tenax TA) of ^{14}C phenanthrene aged over 60 d with four different MWCNTs with varying surface areas in aqueous solution. MWCNTs significantly ($P < 0.05$) reduced the mineralisation of phenanthrene in accordance with their properties, with particles possessing larger specific surface areas together with large meso- and micro-pore volumes resulting in the lowest mineralisation efficiencies. Bacteria were also observed to colonise the surface of MWCNT aggregates, proportional to the quantity of phenanthrene desorbed through Tenax TA extractions (Xia et al., 2013). Although slight changes to the physical appearance of the bacteria were observed when sorbed to MWCNT aggregates, potentially indicating a toxicity effect, the ability of the cells to metabolise phenanthrene sorbed to low surface area particles may not have been significantly reduced (Xia et al., 2013). However, the lack of a control sample in which the metabolism of cells under conditions devoid of CNMs was assessed, limited the ability of the paper to determine the overall impact of MWCNT aggregates on phenanthrene mineralisation.

Very little information is available on how CNMs act within soil matrices, especially in relation to their adsorption to organic fractions, organic pollutants and their subsequent toxicity (Dinesh et al., 2012). With an angelus sorbents such as black carbon (BC), elevated mineralisation of a phenanthrene substrate has been observed as a direct result of BC addition to soil, which was tentatively attributed to microbial sorption and utilisation of phenanthrene from the sorbed phase (Rhodes et al., 2008a; Rhodes et al., 2012). Only one study has identified an increase in contaminant mineralisation in soils following the addition of CNMs. Xia et al. (2010) studied phenanthrene biodegradation and desorption characteristics (using XAD-2) in 21-40 day aged MWCNT-amended soils relative to soils amended with wood char and black carbon. Following each ageing interval, *Agrobacterium* (the degrading inoculum) was added to the soil, and the contaminant degradation efficiency measured. After 28 and 40 days ageing, the degradation efficiency in MWCNT-amended soils was 54.2% and 24.6%, respectively; wood char amended 73.5% and 25.1%, respectively and black carbon amended 83.8% and 38.3%, respectively. Thus a reduction in bioavailability of contaminants sorbed to each of the sorbents with increasing soil contact time is observed (Xia et al., 2010), together with the relatively low bioavailability of contaminants sorbed to MWCNTs relative to other environmental sorbents. Desorption studies identified similar residual concentrations of phenanthrene; however, during rapid stages of degradation, desorption rates were found to under-predict the rate of degradation (Xia et al., 2010). This potentially suggests that for each of the sorbents, phenanthrene was available to bacteria either through the promotion of desorption or direct access (Xia et al., 2010).

Given the discussion above, it is possible to consider an additional factor to those proposed by Kretzschmar et al. (1999) in Section 4, to determine the significance of contaminant facilitated transport by CNMs. If the CNM sorbed contaminant is available to the cells through utilisation from the sorbed phase, the importance of desorption of sorbed compounds from CNMs during transport is reduced. It is therefore proposed that incorporation of a fifth factor, 'the bioavailability and bioaccessibility of CNM sorbed contaminants to microorganisms from the solid

phase', may be appropriate, as inferring bioaccessibility through desorption investigations may lead to incorrect assumptions. However, substantially more work is required to identify the exact mechanism involved in these findings, and the specific conditions under which contaminant and microbial sorption to CNMs could potentially result in toxicity from the CNM itself, from the sorbed contaminant or both (Nowack and Bucheli, 2007). It is also possible that under some environmental conditions, rapid desorption or excessive bioavailability of sorbed contaminants may shock load sorbed bacteria and prove toxic (Upadhyayula and Gadhamshetty, 2010). Biodegradation of contaminants sorbed to CNMs therefore still requires substantial investigation into specific combinations of pollutants and microorganisms (Upadhyayula and Gadhamshetty, 2010), to determine whether the bioaccessibility of sorbed contaminants is either increased or decreased, and if the addition of CNMs will increase the mobility of contaminants in the environment. The general paucity of knowledge regarding the duration for which contaminants will remain sorbed to CNMs requires addressing to determine the long-term stability of contaminants sorbed to different nanoparticle types. Furthermore, the extent to which CNMs influence the transformation residues of HOCs in soils such as bound residues formed during organic pollution degradation in soil is unknown (Barriuso et al., 2008; Shan et al., 2011; Zhou et al., 2013).

6. Summary and conclusions

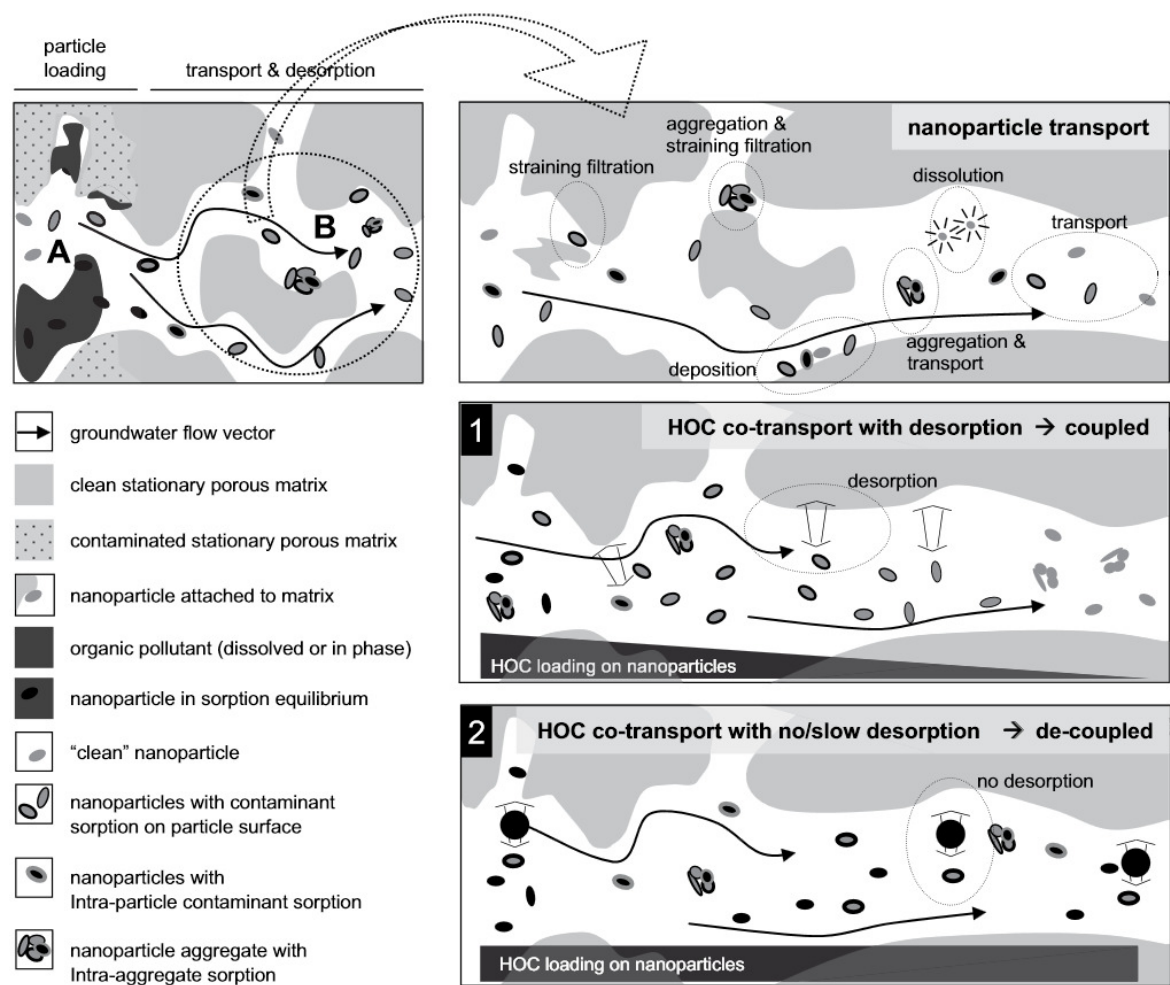
The complex and dynamic nature of both soil environments and CNM physicochemical properties generates enormous uncertainty in attempting to predict their behaviour and impact on contaminant sorption, sequestration and transport as well as microbial interactions. This review argues that the fate and behaviour of CNMs in soils is influenced by multiple parameters such as the type and quantity of SOM, the dominant charge characteristics of the matrix as dictated by the soil inorganic fraction, together with properties of the CNM, each of which is heavily influenced by pH and ionic strength. However, presently only limited research into the

864 manner in which these parameters interact and collaboratively influence the fate and behaviour
865 CNMs is available and significantly more research is required.

866 The extent to which CNMs are able to modify the behaviour of contaminants in soils and
867 facilitate their transport is dependent on the CNM concentration, the properties of SOM,
868 molecular weight of the HOC and interaction of the CNM with the HOC before the addition to
869 soils. When present in sufficient concentrations, CNMs have the ability to facilitate the transport
870 of co-existing contaminants such as PAHs to a greater extent than naturally occurring colloids
871 such as DOM, the extent of which is dependent on the physicochemical properties of the
872 contaminant, CNM functionalization status, aggregation size and method of preparation. Further
873 work derived from experimental research is needed to address the lack of data relating to the
874 transport of CNMs through soils of different properties. Additionally, CNM-HOC desorption
875 kinetics within soils require defining, as this presently limits the understanding of how significant
876 CNM facilitated transport is likely to be.

877 Finally, CNMs are undoubtedly efficient sorbents for a range of HOCs. However, while a
878 reduction in bioaccessibility of contaminants in soils has been demonstrated (Towell et al., 2011),
879 possibly indicating to uses as agents to land reduce bioaccessibility of contaminants, information
880 regarding sorption stability together with their potential to increase contaminant mobilisation
881 and other secondary effects are as yet too poorly developed to fully anticipate the possible
882 environmental impact. To determine the behaviour of CNMs within soils, it is concluded that no
883 one set of environmental or CNM characteristics can be viewed in isolation. Hence, given the
884 diverse array of variables, it is argued that risk-assessment of CNMs within the soil environment
885 should be conducted on a case-by-case basis. Detailed analysis of other environmental
886 compartments in which CNMs can potentially accumulate such as sediments, should also be
887 considered.

888



890 **Figure 1.** 'Worst case scenario' processes by which CNMs may facilitate the transport of HOCs.

891 Top left panel: (A) HOC equilibrates with CNM, and is (B) transported. Top right panel shows the

892 processes by which CNMs may be transported. The centre right panel (1) show the transport and

893 rapid desorption of HOCs from CNMs. Equilibrium is achieved between the liquid phase, CNM and

894 matrix. (2) shows slow desorption kinetics, with no desorption from the CNM (Hofmann and von

895 der Kammer, 2009). Re-printed with permission from Elsevier, © 2014.

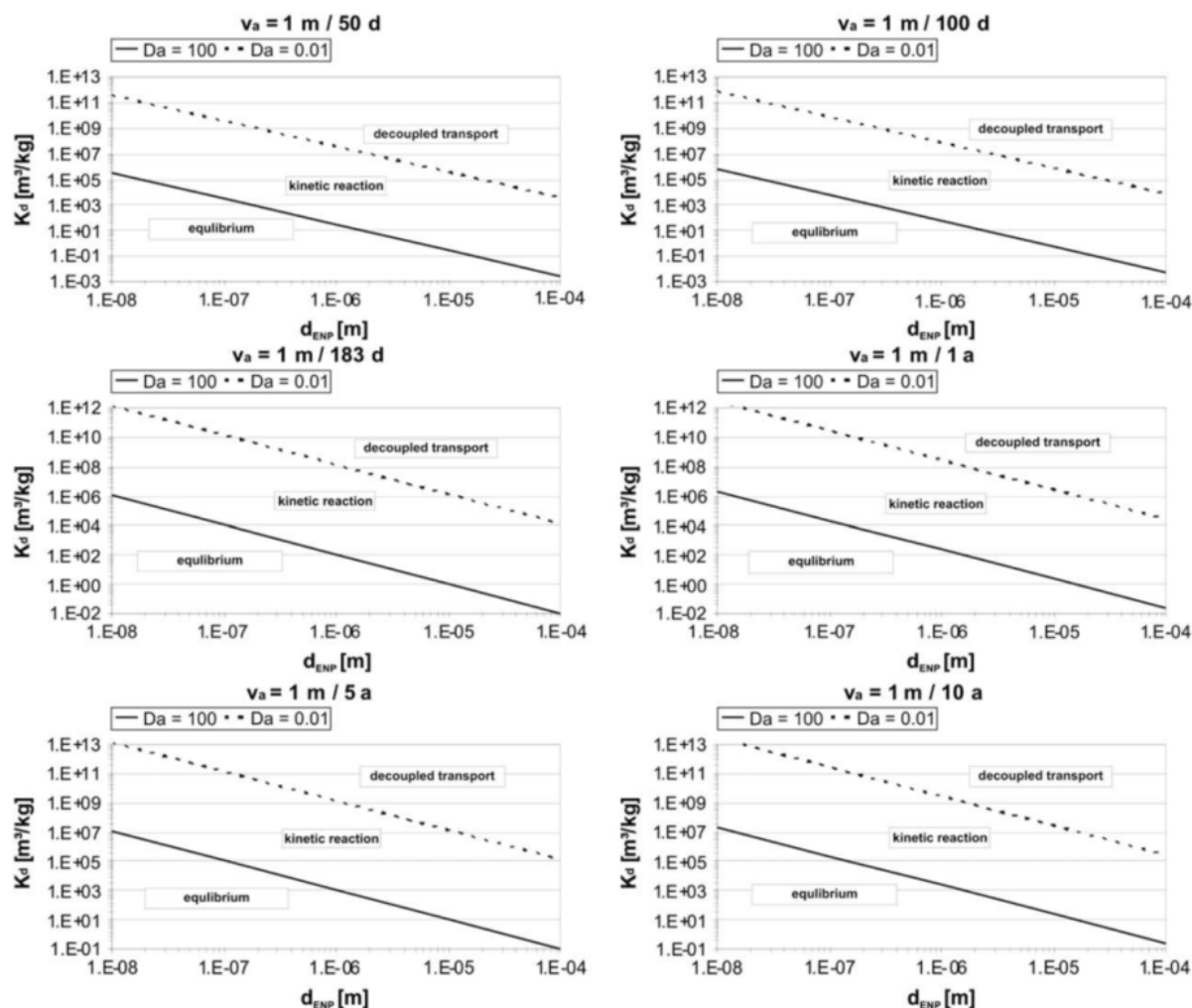


Figure 2. Simulation of diffusion limited desorption using of pore water velocities (v_a) between 1 m/50 d to 1 m/10m/y. The solid line represents the Damköhler number of 100 (representing equilibrium transport above which the HOC will equilibrate between the CNM and soil matrix), the dashed line indicates a Damköhler number of 0.01 (decoupled transport below which HOC desorption will not occur within the timeframe of transport). If Damköhler numbers are < 100 or > 0.01 , kinetics of sorption should be considered in transport models (Hofmann and von der Kammer, 2009). Re-printed with permission from Elsevier, © 2014.

- 905 Abu-Salah, K., Shelef, G., Levanon, D., Armon, R. & Dosoretz, C. G. 1996. Microbial degradation of
906 aromatic and polyaromatic toxic compounds adsorbed on powdered activated carbon.
907 *Journal of Biotechnology*, 51, 265-272.
- 908 Agnihotri, S., Mota, J. P. B., Rostam-Abadi, M. & Rood, M. J. 2005. Structural characterization of
909 single-walled carbon nanotube bundles by experiment and molecular simulation.
910 *Langmuir*, 21, 896-904.
- 911 Allen, B. L., Kichambare, P. D., Gou, P., Vlasova, I. I., Kapralov, A. A., Konduru, N., Kagan, V. E. &
912 Star, A. 2008. Biodegradation of Single-Walled Carbon Nanotubes through Enzymatic
913 Catalysis. *Nano Letters*, 8, 3899-3903.
- 914 Allen, B. L., Kotchey, G. P., Chen, Y., Yanamala, N. V. K., Klein-Seetharaman, J., Kagan, V. E. & Star,
915 A. 2009. Mechanistic Investigations of Horseradish Peroxidase-Catalyzed Degradation of
916 Single-Walled Carbon Nanotubes. *Journal of the American Chemical Society*, 131, 17194-
917 17205.
- 918 Almecija, D., Blond, D., Sader, J. E., Coleman, J. N. & Boland, J. J. 2009. Mechanical properties of
919 individual electrospun polymer-nanotube composite nanofibers. *Carbon*, 47, 2253-2258.
- 920 Aruguete, D. M. & Hochella, M. F. 2010. Bacteria–nanoparticle interactions and their
921 environmental implications. *Environmental Chemistry*, 7, 3-9.
- 922 Barriuso, E., Benoit, P. & Dubus, I. G. 2008. Formation of pesticide nonextractable (bound)
923 residues in soil: Magnitude, controlling factors and reversibility. *Environmental Science &*
924 *Technology*, 42, 1845-1854.
- 925 Bold, S., Kraft, S., Grathwohl, P. & Liedl, R. 2003. Sorption/desorption kinetics of contaminants on
926 mobile particles: Modeling and experimental evidence. *Water Resources Research*, 39,
927 1329.
- 928 Carter, M. C. & Weber, W. J. 1994. Modeling Adsorption of TCE by Activated Carbon Preloaded by
929 Background Organic Matter. *Environmental Science & Technology*, 28, 614-623.
- 930 Cebulska-Wasilewska, A., Pawlyk, I., Panek, A., Wiecheć, A., Kalina, I., Popov, T., Georgieva, T. &
931 Farmer, P. B. 2007. Exposure to environmental polycyclic aromatic hydrocarbons:
932 Influences on cellular susceptibility to DNA damage (sampling Košice and Sofia). *Mutation*
933 *Research*, 620, 145-154.
- 934 Chae, S.-R., Xiao, Y., Lin, S., Noeiaghahi, T., Kim, J.-O. & Wiesner, M. R. 2012. Effects of humic acid
935 and electrolytes on photocatalytic reactivity and transport of carbon nanoparticle
936 aggregates in water. *Water Research*, 46, 4053-4062.
- 937 Chappell, M. A., George, A. J., Dontsova, K. M., Porter, B. E., Price, C. L., Zhou, P., Morikawa, E.,
938 Kennedy, A. J. & Steevens, J. A. 2009. Surfactive stabilization of multi-walled carbon
939 nanotube dispersions with dissolved humic substances. *Environmental Pollution*, 157,
940 1081-1087.
- 941 Chen, J., Chen, W. & Zhu, D. 2008. Adsorption of Nonionic Aromatic Compounds to Single-Walled
942 Carbon Nanotubes: Effects of Aqueous Solution Chemistry. *Environmental Science &*
943 *Technology*, 42, 7225-7230.
- 944 Chen, K. L. & Elimelech, M. 2008. Interaction of Fullerene (C60) Nanoparticles with Humic Acid
945 and Alginate Coated Silica Surfaces: Measurements, Mechanisms, and Environmental
946 Implications. *Environmental Science & Technology*, 42, 7607-7614.
- 947 Chen, K. L. & Elimelech, M. 2009. Relating Colloidal Stability of Fullerene (C60) Nanoparticles to
948 Nanoparticle Charge and Electrokinetic Properties. *Environmental Science & Technology*,
949 43, 7270-7276.
- 950 Chen, W., Duan, L. & Zhu, D. 2007. Adsorption of Polar and Nonpolar Organic Chemicals to Carbon
951 Nanotubes. *Environmental Science & Technology*, 41, 8295-8300.
- 952 Chen, Z., Yadghar, A. M., Zhao, L. & Mi, Z. 2011. A review of environmental effects and
953 management of nanomaterials. *Toxicological & Environmental Chemistry*, 93, 1227-1250.

954 Cheng, G., Zhang, Z., Chen, S., Bryers, J. D. & Jiang, S. 2007. Inhibition of bacterial adhesion and
 955 biofilm formation on zwitterionic surfaces. *Biomaterials*, 28, 4192-4199.

956 Cheng, X., Kan, A. T. & Tomson, M. B. 2005. Uptake and Sequestration of Naphthalene and 1,2-
 957 Dichlorobenzene by C60. *Journal of Nanoparticle Research*, 7, 555-567.

958 Chibowski, E., Espinosa-Jiménez, M., Ontiveros-Ortega, A. & Giménez-Martin, E. 1998. Surface
 959 Free Energy, Adsorption and Zeta Potential in Leacril/Tannic Acid System. *Langmuir*, 14,
 960 5237-5244.

961 Chijiwa, T., Arai, T., Sugai, T., Shinohara, H., Kumazawa, M., Takano, M., Kawakami, S. & Iti 1999.
 962 Fullerenes found in the Permo-Triassic mass extinction period. *Geophysical Research
 963 Letters*, 26, 767-770.

964 Choi, H. & Yavuz Corapcioglu, M. 1997. Transport of a non-volatile contaminant in unsaturated
 965 porous media in the presence of colloids. *Journal of Contaminant Hydrology*, 25, 299-324.

966 Chung, H., Son, Y., Yoon, T. K., Kim, S. & Kim, W. 2011. The effect of multi-walled carbon
 967 nanotubes on soil microbial activity. *Ecotoxicology and Environmental Safety*, 74, 569-575.

968 Colvin, V. L. 2003. The potential environmental impact of engineered nanomaterials. *Nature
 969 Biotechnology*, 21, 1166-1170.

970 Corapcioglu, M. Y., Jiang, S. & Kim, S.-H. 1999. Comparison of Kinetic and Hybrid-Equilibrium
 971 Models Simulating Colloid-Facilitated Contaminant Transport in Porous Media. *Transport
 972 in Porous Media*, 36, 373-390.

973 Cornelissen, G., Van Noort, P. C. M. & Govers, H. a. J. 1997. Desorption kinetics of chlorobenzenes,
 974 polycyclic aromatic hydrocarbons, and polychlorinated biphenyls: Sediment extraction
 975 with Tenax® and effects of contact time and solute hydrophobicity. *Environmental
 976 Toxicology and Chemistry*, 16, 1351-1357.

977 Cui, X. Y., Jia, F., Chen, Y. X. & Gan, J. 2011. Influence of single-walled carbon nanotubes on
 978 microbial availability of phenanthrene in sediment. *Ecotoxicology*, 20, 1277-1285.

979 Darlington, T. K., Neigh, A. M., Spencer, M. T., Guyen, O. T. N. & Oldenburg, S. J. 2009.
 980 Nanoparticle characteristics affecting environmental fate and transport through soil.
 981 *Environmental Toxicology and Chemistry*, 28, 1191-1199.

982 De Jonge, L. W., Kjaergaard, C. & Moldrup, P. 2004. Colloids and Colloid-Facilitated Transport of
 983 Contaminants in Soils: An Introduction. *Vadose Zone Journal*, 3, 321-325.

984 Debnath, S., Cheng, Q., Hedderman, T. G. & Byrne, H. J. 2008. An experimental study of the
 985 interaction between single walled carbon nanotubes and polycyclic aromatic
 986 hydrocarbons. *Physica Status Solidi (B)*, 245, 1961-1963.

987 Dinesh, R., Anandaraj, M., Srinivasan, V. & Hamza, S. 2012. Engineered nanoparticles in the soil
 988 and their potential implications to microbial activity. *Geoderma*, 173-174, 19-27.

989 Doick, K. J., Dew, N. M. & Semple, K. T. 2005. Linking Catabolism to Cyclodextrin Extractability:
 990 Determination of the Microbial Availability of PAHs in Soil. *Environmental Science &
 991 Technology*, 39, 8858-8864.

992 Ekelund, F., Olsson, S. & Johansen, A. 2003. Changes in the succession and diversity of protozoan
 993 and microbial populations in soil spiked with a range of copper concentrations. *Soil
 994 Biology & Biochemistry*, 35, 1507-1516.

995 Eleanor, E. B. C. & Frank, R. 2000. Fullerene reactions. *Reports on Progress in Physics*, 63, 1061.

996 Espinasse, B., Hotze, E. M. & Wiesner, M. R. 2007. Transport and Retention of Colloidal
 997 Aggregates of C60 in Porous Media: Effects of Organic Macromolecules, Ionic
 998 Composition, and Preparation Method. *Environmental Science & Technology*, 41, 7396-
 999 7402.

1000 Esquivel, E. V. & Murr, L. E. 2004. A TEM analysis of nanoparticulates in a Polar ice core. *Materials
 1001 Characterization*, 52, 15-25.

1002 Finkel, M., Liedl, R. & Teutsch, G. 1998. Modelling surfactant-enhanced remediation of polycyclic
 1003 aromatic hydrocarbons. *Environmental Modelling & Software*, 14, 203-211.

1004 Gai, K., Shi, B., Yan, X. & Wang, D. 2011. Effect of Dispersion on Adsorption of Atrazine by
 1005 Aqueous Suspensions of Fullerenes. *Environmental Science & Technology*, 45, 5959-5965.

1006 Gevao, B., Mordaunt, C., Semple, K. T., Pearce, T. G. & Jones, K. C. 2000a. Bioavailability of
 1007 Nonextractable (Bound) Pesticide Residues to Earthworms. *Environmental Science &*
 1008 *Technology*, 35, 501-507.

1009 Gevao, B., Semple, K. T. & Jones, K. C. 2000b. Bound pesticide residues in soils: a review.
 1010 *Environmental Pollution*, 108, 3-14.

1011 Giles, J. 2006. Top five in physics. *Nature*, 441, 265.

1012 Gottschalk, F., Sonderer, T., Scholz, R. W. & Nowack, B. 2009. Modeled Environmental
 1013 Concentrations of Engineered Nanomaterials (TiO₂, ZnO, Ag, CNT, Fullerenes) for
 1014 Different Regions. *Environmental Science & Technology*, 43, 9216-9222.

1015 Gottschalk, F., Sonderer, T., Scholz, R. W. & Nowack, B. 2010. Possibilities and limitations of
 1016 modeling environmental exposure to engineered nanomaterials by probabilistic material
 1017 flow analysis. *Environmental Toxicology and Chemistry*, 29, 1036-1048.

1018 Heymann, D., Chibante, L. P., Brooks, R. R., Wolbach, W. S. & Smalley, R. E. 1994. Fullerenes in the
 1019 Cretaceous-Tertiary Boundary-Layer. *Science*, 265, 645-647.

1020 Hofmann, T. & Von Der Kammer, F. 2009. Estimating the relevance of engineered carbonaceous
 1021 nanoparticle facilitated transport of hydrophobic organic contaminants in porous media.
 1022 *Environmental Pollution*, 157, 1117-1126.

1023 Hou, W.-C. & Jafvert, C. T. 2008. Photochemical Transformation of Aqueous C₆₀ Clusters in
 1024 Sunlight. *Environmental Science & Technology*, 43, 362-367.

1025 Hou, W.-C. & Jafvert, C. T. 2009. Photochemistry of Aqueous C₆₀ Clusters: Evidence of 102
 1026 Formation and its Role in Mediating C₆₀ Phototransformation. *Environmental Science &*
 1027 *Technology*, 43, 5257-5262.

1028 Hu, H., Yu, A., Kim, E., Zhao, B., Itkis, M. E., Bekyarova, E. & Haddon, R. C. 2005. Influence of the
 1029 Zeta Potential on the Dispersability and Purification of Single-Walled Carbon Nanotubes.
 1030 *The Journal of Physical Chemistry B*, 109, 11520-11524.

1031 Hyung, H., Fortner, J. D., Hughes, J. B. & Kim, J.-H. 2006. Natural Organic Matter Stabilizes Carbon
 1032 Nanotubes in the Aqueous Phase. *Environmental Science & Technology*, 41, 179-184.

1033 Hyung, H. & Kim, J.-H. 2008. Natural Organic Matter (NOM) Adsorption to Multi-Walled Carbon
 1034 Nanotubes: Effect of NOM Characteristics and Water Quality Parameters. *Environmental*
 1035 *Science & Technology*, 42, 4416-4421.

1036 Ibaraki, M. & Sudicky, E. A. 1995. Colloid-facilitated contaminant transport in discretely fractured
 1037 porous media: 1. Numerical formulation and sensitivity analysis. *Water Resources*
 1038 *Research*, 31, 2945-2960.

1039 Isaacson, C., Zhang, W., Powell, T., Ma, X. & Bouchard, D. 2011. Temporal Changes in Aqu/C₆₀
 1040 Physical-Chemical, Deposition, and Transport Characteristics in Aqueous Systems.
 1041 *Environmental Science & Technology*, 45, 5170-5177.

1042 Jafar, G. & Hamzeh, G. 2013. Ecotoxicity of Nanomaterials in Soil. *Annals of Biological Research*, 4,
 1043 86-92.

1044 Jaisi, D. P. & Elimelech, M. 2009. Single-Walled Carbon Nanotubes Exhibit Limited Transport in Soil
 1045 Columns. *Environmental Science & Technology*, 43, 9161-9166.

1046 Jennings, A. A. & Kirkner, D. J. 1984. Instantaneous Equilibrium Approximation Analysis. *Journal of*
 1047 *Hydraulic Engineering*, 110, 1700-1717.

1048 Johansen, A., Pedersen, A. L., Jensen, K. A., Karlson, U., Hansen, B. M., Scott-Fordsmand, J. J. &
 1049 Winding, A. 2008. Effects of C₆₀ fullerene nanoparticles on soil bacteria and protozoans.
 1050 *Environmental Toxicology and Chemistry*, 27, 1895-1903.

1051 Kan, A. T. & Tomson, M. B. 1990. Ground water transport of hydrophobic organic compounds in
 1052 the presence of dissolved organic matter. *Environmental Toxicology and Chemistry*, 9,
 1053 253-263.

1054 Kanepalli, S. & Donna, F. E. 2006. Enhancing the remediation of trichloroethene (TCE) using
 1055 double-walled carbon nanotubes (DWCNT). United States Geological Survey.

1056 Kang, S., Pinault, M., Pfefferle, L. D. & Elimelech, M. 2007. Single-Walled Carbon Nanotubes
 1057 Exhibit Strong Antimicrobial Activity. *Langmuir*, 23, 8670-8673.

- Köhler, A. R., Som, C., Helland, A. & Gottschalk, F. 2008. Studying the potential release of carbon nanotubes throughout the application life cycle. *Journal of Cleaner Production*, 16, 927-937.
- Kretzschmar, R., Borkovec, M., Grolimund, D. & Elimelech, M. 1999. Mobile Subsurface Colloids and Their Role in Contaminant Transport. In: Donald, L. S. (ed.) *Advances in Agronomy*. Academic Press.
- Kuznar, Z. A. & Elimelech, M. 2004. Adhesion Kinetics of Viable *Cryptosporidium parvum* Oocysts to Quartz Surfaces. *Environmental Science & Technology*, 38, 6839-6845.
- Kwon, J.-H. 2012. Destabilization of aqueous colloidal C60 nanoparticles in the presence of various organic matter. *CLEAN – Soil, Air, Water*, 40, 472-478.
- Lead, J. R. & Wilkinson, K. J. 2006. Aquatic colloids and nanoparticles: Current knowledge and future trends. *Environmental Chemistry*, 3, 159-171.
- Lee, J., Cho, M., Fortner, J. D., Hughes, J. B. & Kim, J.-H. 2009. Transformation of Aggregated C60 in the Aqueous Phase by UV Irradiation. *Environmental Science & Technology*, 43, 4878-4883.
- Lee, M. C., Snoeyink, V. L. & Crittenden, J. C. 1981. Activated carbon adsorption of humic substances. *American Water Works Association*, 73, 440-446.
- Lerman, I., Chen, Y. & Chefetz, B. 2013. Adsorption of Contaminants of Emerging Concern by Carbon Nanotubes: Influence of Dissolved Organic Matter. In: Xu, J., Wu, J. & He, Y. (eds.) *Functions of Natural Organic Matter in Changing Environment*. Springer Netherlands.
- Li, D., Lyon, D. Y., Li, Q. & Alvarez, P. J. J. 2008. Effect of soil sorption and aquatic natural organic matter on the antibacterial activity of a fullerene water suspension. *Environmental Toxicology and Chemistry*, 27, 1888-1894.
- Li, Q., Xie, B., Hwang, Y. S. & Xu, Y. 2009. Kinetics of C60 Fullerene Dispersion in Water Enhanced by Natural Organic Matter and Sunlight. *Environmental Science & Technology*, 43, 3574-3579.
- Li, S. 2012. *A study of environmental fate and application of commercially available carbon nanotubes*. Doctor of Philosophy Thesis, Texas Tech University.
- Li, W., Zhu, X., He, Y., Xing, B., Xu, J. & Brookes, P. C. 2013. Enhancement of water solubility and mobility of phenanthrene by natural soil nanoparticles. *Environmental Pollution*, 176, 228-233.
- Li, W. J. & Liang, W. J. 2007. Loss of characteristic absorption bands of C60 conjugation systems in the addition with aliphatic amines. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 67, 1346-1350.
- Lin, D. & Xing, B. 2008. Tannic Acid Adsorption and Its Role for Stabilizing Carbon Nanotube Suspensions. *Environmental Science & Technology*, 42, 5917-5923.
- Liu, C., Fan, Y. Y., Liu, M., Cong, H. T., Cheng, H. M. & Dresselhaus, M. S. 1999. Hydrogen Storage in Single-Walled Carbon Nanotubes at Room Temperature. *Science*, 286, 1127-1129.
- Lou, L., Luo, L., Wang, W., Xu, X., Hou, J., Xun, B. & Chen, Y. 2011. Impact of black carbon originated from fly ash and soot on the toxicity of pentachlorophenol in sediment. *Journal of Hazardous Materials*, 190, 474-479.
- Lubick, N. 2008. Risks of Nanotechnology Remain Uncertain. *Environmental Science & Technology*, 42, 1821-1824.
- Lucafò, M., Pacor, S., Fabbro, C., Da Ros, T., Zorzet, S., Prato, M. & Sava, G. 2012. Study of a potential drug delivery system based on carbon nanoparticles: effects of fullerene derivatives in MCF7 mammary carcinoma cells. *Journal of Nanoparticle Research*, 14, 1-13.
- Mauter, M. S. & Elimelech, M. 2008. Environmental Applications of Carbon-Based Nanomaterials. *Environmental Science & Technology*, 42, 5843-5859.
- Mehmannavaz, R., Prasher, S. O. & Ahmad, D. 2001. Cell surface properties of rhizobial strains isolated from soils contaminated with hydrocarbons: hydrophobicity and adhesion to sandy soil. *Process Biochemistry*, 36, 683-688.

1110 Menzie, C. A., Potocki, B. B. & Santodonato, J. 1992. Exposure to carcinogenic PAHs in the
 1111 environment. *Environmental Science & Technology*, 26, 1278-1284.

1112 Mohanty, B., Anita, K. V., Claesson, P. & Bohidar, H. B. 2007. Physical and anti-microbial
 1113 characteristics of carbon nanoparticles prepared from lamp soot. *Nanotechnology*, 18,
 1114 445102.

1115 Morikawa, M. 2006. Beneficial biofilm formation by industrial bacteria *Bacillus subtilis* and related
 1116 species. *Journal of Bioscience and Bioengineering*, 101, 1-8.

1117 Mueller, N. C. & Nowack, B. 2008. Exposure Modeling of Engineered Nanoparticles in the
 1118 Environment. *Environmental Science & Technology*, 42, 4447-4453.

1119 Navarro, E., Baun, A., Behra, R., Hartmann, N., Filser, J., Miao, A.-J., Quigg, A., Santschi, P. & Sigg,
 1120 L. 2008. Environmental behavior and ecotoxicity of engineered nanoparticles to algae,
 1121 plants, and fungi. *Ecotoxicology*, 17, 372-386.

1122 Neal, A. L. 2008. What can be inferred from bacterium–nanoparticle interactions about the
 1123 potential consequences of environmental exposure to nanoparticles? *Ecotoxicology*, 17,
 1124 362-371.

1125 Nowack, B. & Bucheli, T. D. 2007. Occurrence, behavior and effects of nanoparticles in the
 1126 environment. *Environmental Pollution*, 150, 5-22.

1127 Pan, B. & Xing, B. 2008. Adsorption Mechanisms of Organic Chemicals on Carbon Nanotubes.
 1128 *Environmental Science & Technology*, 42, 9005-9013.

1129 Pan, B. & Xing, B. 2010. Manufactured nanoparticles and their sorption of organic chemicals.
 1130 *Advances in Agronomy*, 108, 137-181.

1131 Pan, B. & Xing, B. 2012. Applications and implications of manufactured nanoparticles in soils: a
 1132 review. *European Journal of Soil Science*, 63, 437-456.

1133 Pang, C. M., Hong, P., Guo, H. & Liu, W.-T. 2005. Biofilm formation characteristics of bacterial
 1134 isolates retrieved from a reverse osmosis membrane. *Environmental Science &*
 1135 *Technology*, 39, 7541-7550.

1136 Peretz, S. & Regev, O. 2012. Carbon nanotubes as nanocarriers in medicine. *Current Opinion in*
 1137 *Colloid & Interface Science*, 17, 360-368.

1138 Petersen, E. J. & Henry, T. B. 2012. Methodological considerations for testing the ecotoxicity of
 1139 carbon nanotubes and fullerenes: Review. *Environmental Toxicology and Chemistry*, 31,
 1140 60-72.

1141 Petersen, E. J., Zhang, L., Mattison, N. T., O'carroll, D. M., Whelton, A. J., Uddin, N., Nguyen, T.,
 1142 Huang, Q., Henry, T. B., Holbrook, R. D. & Chen, K. L. 2011. Potential Release Pathways,
 1143 Environmental Fate, And Ecological Risks of Carbon Nanotubes. *Environmental Science &*
 1144 *Technology*, 45, 9837-9856.

1145 Qu, X., Alvarez, P. J. J. & Li, Q. 2012. Impact of Sunlight and Humic Acid on the Deposition Kinetics
 1146 of Aqueous Fullerene Nanoparticles (nC60). *Environmental Science & Technology*, 46,
 1147 13455-13462.

1148 Reid, B. J., Stokes, J. D., Jones, K. C. & Semple, K. T. 2000. Nonexhaustive Cyclodextrin-Based
 1149 Extraction Technique for the Evaluation of PAH Bioavailability. *Environmental Science &*
 1150 *Technology*, 34, 3174-3179.

1151 Ren, X., Chen, C., Nagatsu, M. & Wang, X. 2011. Carbon nanotubes as adsorbents in
 1152 environmental pollution management: A review. *Chemical Engineering Journal*, 170, 395-
 1153 410.

1154 Rhodes, A. H., Carlin, A. & Semple, K. T. 2008a. Impact of Black Carbon in the Extraction and
 1155 Mineralization of Phenanthrene in Soil. *Environmental Science & Technology*, 42, 740-745.

1156 Rhodes, A. H., Dew, N. M. & Semple, K. T. 2008b. Relationship between cyclodextrin extraction
 1157 and biodegradation of phenanthrene in soil. *Environmental Toxicology and Chemistry*, 27,
 1158 1488-1495.

1159 Rhodes, A. H., Riding, M. J., Mcallister, L. E., Lee, K. & Semple, K. T. 2012. Influence of Activated
 1160 Charcoal on Desorption Kinetics and Biodegradation of Phenanthrene in Soil.
 1161 *Environmental Science & Technology*, 46, 12445-12451.

Richards, J. J., Rice, A. H., Nelson, R. D., Kim, F. S., Jenekhe, S. A., Luscombe, C. K. & Pozzo, D. C. 2012. Modification of PCBM Crystallization Via Incorporation of C60 in Polymer/Fullerene Solar Cells. *Advanced Functional Materials*, 23, 514-522.

Riding, M. J., Martin, F. L., Trevisan, J., Llabjani, V., Patel, I., Jones, K. C. & Semple, K. T. 2012a. Concentration-dependent effects of carbon nanoparticles in gram-negative bacteria determined by infrared spectroscopy with multivariate analysis. *Environmental Pollution*, 163, 226-234.

Riding, M. J., Trevisan, J., Hirschmugl, C. J., Jones, K. C., Semple, K. T. & Martin, F. L. 2012b. Mechanistic insights into nanotoxicity determined by synchrotron radiation-based Fourier-transform infrared imaging and multivariate analysis. *Environment International*, 50, 56-65.

Roy, S. B. & Dzombak, D. A. 1998. Sorption nonequilibrium effects on colloid-enhanced transport of hydrophobic organic compounds in porous media. *Journal of Contaminant Hydrology*, 30, 179-200.

Sanchez, P. A. 1976. *Properties and management of soils in the tropics*, New York, USA, John Wiley and Sons.

Sayes, C. M., Fortner, J. D., Guo, W., Lyon, D., Boyd, A. M., Ausman, K. D., Tao, Y. J., Sitharaman, B., Wilson, L. J., Hughes, J. B., West, J. L. & Colvin, V. L. 2004. The Differential Cytotoxicity of Water-Soluble Fullerenes. *Nano Letters*, 4, 1881-1887.

Schreiner, K. M., Filley, T. R., Blanchette, R. A., Bowen, B. B., Bolskar, R. D., Hockaday, W. C., Masiello, C. A. & Raebiger, J. W. 2009. White-Rot Basidiomycete-Mediated Decomposition of C60 Fullerol. *Environmental Science & Technology*, 43, 3162-3168.

Semple, K. T., Doick, K. J., Jones, K. C., Burauel, P., Craven, A. & Harms, H. 2004. Peer Reviewed: Defining Bioavailability and Bioaccessibility of Contaminated Soil and Sediment is Complicated. *Environmental Science & Technology*, 38, 228A-231A.

Semple, K. T., Riding, M. J., Mcallister, L. E., Sopena-Vazquez, F. & Bending, G. D. 2013. Impact of black carbon on the bioaccessibility of organic contaminants in soil. *Journal of Hazardous Materials*, 261, 808-816.

Sen, T. K. & Khilar, K. C. 2006. Review on subsurface colloids and colloid-associated contaminant transport in saturated porous media. *Advances in Colloid and Interface Science*, 119, 71-96.

Shan, J., Jiang, B., Yu, B., Li, C., Sun, Y., Guo, H., Wu, J., Klumpp, E., Schäffer, A. & Ji, R. 2011. Isomer-Specific Degradation of Branched and Linear 4-Nonylphenol Isomers in an Oxidic Soil. *Environmental Science & Technology*, 45, 8283-8289.

Sharma, P. & Ahuja, P. 2008. Recent advances in carbon nanotube-based electronics. *Materials Research Bulletin*, 43, 2517-2526.

Shaw, G. & Connell, D. 1994. Prediction and Monitoring of the Carcinogenicity of Polycyclic Aromatic Compounds (PACs). In: Ware, G. (ed.) *Reviews of Environmental Contamination and Toxicology*. Springer New York.

Shieh, Y.-T., Liu, G.-L., Wu, H.-H. & Lee, C.-C. 2007. Effects of polarity and pH on the solubility of acid-treated carbon nanotubes in different media. *Carbon*, 45, 1880-1890.

Simon-Deckers, A. L., Loo, S., Mayne-L'hermite, M., Herlin-Boime, N., Menguy, N., Reynaud, C. C., Gouget, B. & Carrière, M. 2009. Size-, Composition- and Shape-Dependent Toxicological Impact of Metal Oxide Nanoparticles and Carbon Nanotubes toward Bacteria. *Environmental Science & Technology*, 43, 8423-8429.

Singh, P. & Cameotra, S. S. 2004. Enhancement of metal bioremediation by use of microbial surfactants. *Biochemical and Biophysical Research Communications*, 319, 291-297.

Singh, R., Paul, D. & Jain, R. K. 2006. Biofilms: implications in bioremediation. *Trends in Microbiology*, 14, 389-397.

Smith, B., Wepasnick, K., Schrote, K. E., Bertele, A. R., Ball, W. P., O'melia, C. & Fairbrother, D. H. 2008. Colloidal Properties of Aqueous Suspensions of Acid-Treated, Multi-Walled Carbon Nanotubes. *Environmental Science & Technology*, 43, 819-825.

1214 Smith, B., Wepasnick, K., Schrote, K. E., Cho, H.-H., Ball, W. P. & Fairbrother, D. H. 2009. Influence
1215 of Surface Oxides on the Colloidal Stability of Multi-Walled Carbon Nanotubes: A
1216 Structure–Property Relationship. *Langmuir*, 25, 9767-9776.

1217 Snow, E. S., Perkins, F. K., Houser, E. J., Badescu, S. C. & Reinecke, T. L. 2005. Chemical Detection
1218 with a Single-Walled Carbon Nanotube Capacitor. *Science*, 307, 1942-1945.

1219 Sollins, P., Robertson, G. P. & Uehara, G. 1988. Nutrient mobility in variable- and permanent-
1220 charge soils. *Biogeochemistry*, 6, 181-199.

1221 Stokes, J. D., Paton, G. I. & Semple, K. T. 2005. Behaviour and assessment of bioavailability of
1222 organic contaminants in soil: relevance for risk assessment and remediation. *Soil Use and*
1223 *Management*, 21, 475-486.

1224 Terashima, M. & Nagao, S. 2007. Solubilization of [60]Fullerene in water by aquatic humic
1225 substances. *Chemistry Letters*, 36, 302-303.

1226 Tong, Z., Bischoff, M., Nies, L., Applegate, B. & Turco, R. F. 2007. Impact of Fullerene (C60) on a
1227 Soil Microbial Community. *Environmental Science & Technology*, 41, 2985-2991.

1228 Towell, M. G., Browne, L. A., Paton, G. I. & Semple, K. T. 2011. Impact of carbon nanomaterials on
1229 the behaviour of 14C-phenanthrene and 14C-benzo-[a] pyrene in soil. *Environmental*
1230 *Pollution*, 159, 706-715.

1231 Turco, R. F., Bischoff, M., Tong, Z. H. & Nies, L. 2011. Environmental implications of nanomaterials:
1232 are we studying the right thing? *Current Opinion in Biotechnology*, 22, 527-532.

1233 Upadhyayula, V. K. K., Deng, S., Smith, G. B. & Mitchell, M. C. 2009. Adsorption of *Bacillus subtilis*
1234 on single-walled carbon nanotube aggregates, activated carbon and NanoCeram™. *Water*
1235 *Research*, 43, 148-156.

1236 Upadhyayula, V. K. K. & Gadhamshetty, V. 2010. Appreciating the role of carbon nanotube
1237 composites in preventing biofouling and promoting biofilms on material surfaces in
1238 environmental engineering: A review. *Biotechnology Advances*, 28, 802-816.

1239 Velasco-Santos, C., Martinez-Hernandez, A. L., Consultchi, A., Rodriguez, R. & Castaño, V. M. 2003.
1240 Naturally produced carbon nanotubes. *Chemical Physics Letters*, 373, 272-276.

1241 Wang, L., Fortner, J. D., Hou, L., Zhang, C., Kan, A. T., Tomson, M. B. & Chen, W. 2012a.
1242 Contaminant-mobilizing capability of fullerene nanoparticles (nC60): Effect of solvent-
1243 exchange process in nC60 formation. *Environmental Toxicology and Chemistry*, n/a-n/a.

1244 Wang, L., Huang, Y., Kan, A. T., Tomson, M. B. & Chen, W. 2012b. Enhanced Transport of 2,2',5,5'-
1245 Polychlorinated Biphenyl by Natural Organic Matter (NOM) and Surfactant-Modified
1246 Fullerene Nanoparticles (nC60). *Environmental Science & Technology*, 46, 5422-5429.

1247 Wang, P. & Keller, A. A. 2009. Natural and Engineered Nano and Colloidal Transport: Role of Zeta
1248 Potential in Prediction of Particle Deposition. *Langmuir*, 25, 6856-6862.

1249 Wang, P., Shi, Q., Liang, H., Steuerman, D. W., Stucky, G. D. & Keller, A. A. 2008a. Enhanced
1250 Environmental Mobility of Carbon Nanotubes in the Presence of Humic Acid and Their
1251 Removal from Aqueous Solution. *Small*, 4, 2166-2170.

1252 Wang, X., Lu, J. & Xing, B. 2008b. Sorption of Organic Contaminants by Carbon Nanotubes:
1253 Influence of Adsorbed Organic Matter. *Environmental Science & Technology*, 42, 3207-
1254 3212.

1255 Wang, X., Shu, L., Wang, Y., Xu, B., Bai, Y., Tao, S. & Xing, B. 2011. Sorption of Peat Humic Acids to
1256 Multi-Walled Carbon Nanotubes. *Environmental Science & Technology*, 45, 9276-9283.

1257 Wang, X., Tao, S. & Xing, B. 2009. Sorption and Competition of Aromatic Compounds and Humic
1258 Acid on Multiwalled Carbon Nanotubes. *Environmental Science & Technology*, 43, 6214-
1259 6219.

1260 Xia, X., Li, Y., Zhou, Z. & Feng, C. 2010. Bioavailability of adsorbed phenanthrene by black carbon
1261 and multi-walled carbon nanotubes to *Agrobacterium*. *Chemosphere*, 78, 1329-1336.

1262 Xia, X., Zhou, C., Huang, J., Wang, R. & Xia, N. 2013. Mineralization of phenanthrene sorbed on
1263 multiwalled carbon nanotubes. *Environmental Toxicology and Chemistry*, n/a-n/a.

1264 Yan, H., Pan, G., Zou, H., Li, X. & Chen, H. 2004. Effective removal of microcystins using carbon
1265 nanotubes embedded with bacteria. *Chinese Science Bulletin*, 49, 1694-1698.

1266 Yang, K., Wang, X., Zhu, L. & Xing, B. 2006a. Competitive Sorption of Pyrene, Phenanthrene, and
1267 Naphthalene on Multiwalled Carbon Nanotubes. *Environmental Science & Technology*, 40,
1268 5804-5810.

1269 Yang, K. & Xing, B. 2007. Desorption of polycyclic aromatic hydrocarbons from carbon
1270 nanomaterials in water. *Environmental Pollution*, 145, 529-537.

1271 Yang, K., Zhu, L. & Xing, B. 2006b. Adsorption of Polycyclic Aromatic Hydrocarbons by Carbon
1272 Nanomaterials. *Environmental Science & Technology*, 40, 1855-1861.

1273 Zhang, L., Hou, L., Wang, L., Kan, A. T., Chen, W. & Tomson, M. B. 2012a. Transport of Fullerene
1274 Nanoparticles (nC60) in Saturated Sand and Sandy Soil: Controlling Factors and Modeling.
1275 *Environmental Science & Technology*, 46, 7230-7238.

1276 Zhang, L., Petersen, E. J. & Huang, Q. 2011a. Phase Distribution of 14C-Labeled Multiwalled
1277 Carbon Nanotubes in Aqueous Systems Containing Model Solids: Peat. *Environmental*
1278 *Science & Technology*, 45, 1356-1362.

1279 Zhang, L., Petersen, E. J., Zhang, W., Chen, Y., Cabrera, M. & Huang, Q. 2012b. Interactions of 14C-
1280 labeled multi-walled carbon nanotubes with soil minerals in water. *Environmental*
1281 *Pollution*, 166, 75-81.

1282 Zhang, L., Wang, L., Zhang, P., Kan, A. T., Chen, W. & Tomson, M. B. 2011b. Facilitated Transport
1283 of 2,2',5,5'-Polychlorinated Biphenyl and Phenanthrene by Fullerene Nanoparticles
1284 through Sandy Soil Columns. *Environmental Science & Technology*, 45, 1341-1348.

1285 Zhang, S., Shao, T. & Karanfil, T. 2011c. The effects of dissolved natural organic matter on the
1286 adsorption of synthetic organic chemicals by activated carbons and carbon nanotubes.
1287 *Water Research*, 45, 1378-1386.

1288 Zhou, W., Shan, J., Jiang, B., Wang, L., Feng, J., Guo, H. & Ji, R. 2013. Inhibitory effects of carbon
1289 nanotubes on the degradation of 14C-2,4-dichlorophenol in soil. *Chemosphere*, 90, 527-
1290 534.

1291