Author replies to comments on “soild-1-267-2014”

Page 269, Lines 14-18, I think it is important to acknowledge here that the idea of organic matter composition/chemistry controlling decomposition (esp. in the long-term) is currently challenged in the literature (ex. see Schmidt et al. 2011, Nature).

Ok, we changed (see P. 3, L. 51-53 of the revised Word file) to: “The current common belief is that environmental and biological factors predominate on the molecular structure in controlling SOM stability (Kleber, 2010; Kleber et al., 2011; Schmidt et al., 2011); however, in some environments, vegetation cover is a good proxy for soil C dynamics, since it controls the input and quality of litter (De Deyn et al., 2008).”

Page 269, line 17: Please replace humification with 'decomposition', 'transformation' or 'processing' (see comment above).

“Humification processes” were replaced by “SOM transformation” (see P. 3, L. 55 of the revised Word file).

Page 271, lines 1-4: here and later in discussion you have to address how measuring changes in rate of decomposition (and chemistry of decomposing litter) over 1 yr will give reliable information of SOM quality and dynamics? This is important in order to establish a logical link between the short-term dynamics and long-term dynamics when it comes to amount, composition, and decomposability of the litter that grows and decomposes on the surface, and the portion of the litter that actually enters the soil organic matter pool (even in the near-surface soil layers).

More prudentially, now we inserted the terms “initial” or “early stages of” before litter decomposition rate. Furthermore, hereafter we clearly specified that our short-term study only provides indications (and nothing more) on the long-term fate of SOM.

Page 273, line 4: please include information on the type of filter was used for filtration of dissolved OM.

The used filter was a 0.45 µm membrane filter (Millipore). This is now included in the text, here and some lines above (see P. 7, L. 163 and 175 of the revised Word file).

Page 275, lines 15-18: was there any soil material that couldn't be cleaned (i.e. was too strongly attached to the remaining litter)? If so, how was it handled?

No, everything was quite easy to remove from the bags by using a soft brush. Such information is now included in the text (see P. 9, L. 228 of the revised Word file).

276, line 6: Please replace 'intrinsic recalcitrance' (see comment above and Kleber 2010, Environmental Chemistry in addition to Schmidt et al 2011), possibly you can say 'short-term resistance to decomposition'?

We understand this criticism, which is in line with the first one. In that case we acknowledged that the persistence of soil organic matter is partly an ecosystem property and in fact we quoted the paper by Schmidt et al. 2011 published in Nature, as the Reviewer suggested, and two valuable others. However, here we just wanted to say “intrinsic recalcitrance”, which means the ability to resist degradation due to its chemical nature. Recent findings and new concepts cannot lead to deny that a simple sugar is less resistant to degradation than lignin and that high C/N ratios are symptomatic of more difficult degradation. Anyway, the sentence in question, which is not a result, was deleted. Additionally, please note that after your comment, at P. 15, L. 413 we changed “…intrinsic resistance of Sphagnum” to “…short-term resistance of Sphagnum to degradation”.

276, line 6-9: this statement is not supported by any data or conclusive evidence and should be taken out.

Ok, we removed this sentence.

Table 1: please mark which of the differences in means are statistically significant. Does the statement in line 15-17, page 276 suggest that all the values are insignificant or just the C:N ratio?

Correct comment: this part was in fact unclear. Significant differences between the plant species with respect to C, N and C/N ratios have been included in Table 1 and the reorganized text to account for significant differences only (see P. 10 in the Results of the revised Word file).

Please revise the statement in Page 278, lines 26-28, what does 'side-effect' refer to?

The term we used is “site” effect, not “side” effect. It is largely used in the literature and we think it does not require further explanation in the text.

Pages 280 (last line) -281 line 1, alkyl/O-alkyl ratio as per Baldock and co-workers and others is an index of SOM transformation/processing, not 'recalcitrance' per se (even though in some cases these two can be related). Please either provide a correlation coefficient or at least prior reference to support this statement.

Actually, the Reviewer is right in saying it. We adjusted this sentence accordingly, based also on the enlightening papers by Kleber and co-authors now cited in the manuscript.

Page 281, lines 5-8: this statement can also be valid only if supported by a qualifying statement as to the short-term (1yr) nature of this study and findings there in. It is possible that after a year (possibly after a year of processing by meso- and macroorganisms), it could be rendered more decomposable.

To be honest, everything is possible after the period we investigated, i.e. after 1yr. However, we do not like to make any hypothesis beyond the monitored time lapse. Also, we do not feel the necessity to further specify, here, what requested by the Reviewer. In fact, in the statement in question we were talking about the chemical capacity of the material in the litterbags to resist decomposition (of course, up to the time when we removed it from the ground for analysis).

Page 283, line 1, please consider revising 'parent vegetation'

Ok, "…quality of the parent vegetation" was changed to "…litter quality" (see P. 16, L. 487 of the revised Word file).

Page 283, line 2: please revise 'refractory' (see comment above)

The term "early stage of" decomposition was actually included in this conclusion, to clarify that it just refers to a short-term fate. Moreover, we replaced the term "refractory" with the more comprehensive and less compromising "stable", which refers to the capacity to resist degradation not only due to the intrinsic molecular recalcitrance.

Page 283, line 7: please revise 'higher concentration of rainfall' … do you mean higher amount or distribution of rainfall?

Such a long and a bit confused sentence was changed to "…assessing the consequences of climate change on SOM stocks and dynamics".

Page 283, line 9: please note that your study can only suggest this conclusion for short-term. You haven't presented any model fitting (or other approaches) to demonstrate what can be expected in the long-term.

Right. The "short-term" perspective of our study was acknowledged.

Other general comments:

Please address how use of litterbags could have affected your results. This is important because several studies have shown there is a significant effect of litterbags in affecting rate of organic substrate decomposition and chemistry of decomposing litter, in addition the effect of litterbags is also modulated by soil hydrologic conditions (ex. see Berhe 2013, Journal of Soils and Sediments).

We honestly admitted, at P. 16, L 464-467, what types of bias from using litterbags could have affected our results. The enlightening paper by Berhe was of course quoted. Nevertheless, we may assume that such a bagging effect in our experiment is uniform through the samples, also because the bags are very similar.

Please include error bars in Figures 3 and 4.

We included error bars in Figure 3, but preferred to not do the same with Figure 4, because otherwise it would become unreadable. On the other hand, there are papers where quite similar patterns are devoid of error bars (e.g., Rezig et al., 2014. International Journal of Recycling of Organic Waste in Agriculture, 3: 1-9).

I think it is important to fit the litter decomposition data with one or two pool decomposition models to derive important indices (ex. long-term rate of decomposition) that would make findings of this study even more significant and easier to interpret. Please see works of Mark Harmon and co-workers (1992, 1999, 2009) as a reference for this.
We read the suggested papers by Harmon and co-workers and some others dealing with long-term patterns of mass loss during litter decomposition. Nevertheless, we feel that one year of investigation is not enough to reveal dynamics in the later phases. As a consequence, we prefer to avoid using decomposition models and making inferences concerning the long-term fate of litter. Actually, what we were interested to in this paper was to check the site- and litter-quality effects in the early stage of decomposition.

RE: Author responses to reviewer comments

The authors are strongly encouraged to address EACH comment (separated by start of the new line, or page and line numbers proceeding comments) raised by the reviewers individually. Please add your response to each comment in bold, italic, indented (or other format) text for the sake of clarity.

Done.

We'd like to thank the Editor and Reviewers for their help in improving the quality of our manuscript with their useful comments.

The Authors

Litter decomposition rate and soil organic matter quality in a patchwork heathland of Southern Norway

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Abstract

Norwegian heathland soils, although scant and shallow, are major reservoirs of carbon (C). We aimed at assessing whether vegetation cover and, indirectly, its driving factor soil drainage are good proxies for soil organic matter (SOM) composition and dynamics in a typical heathland area of Southern Norway consisting in a patchwork of three different types of vegetation, dominated by Calluna (*Calluna vulgaris* (L) Hull), Molinia (*Molinia caerulea* (L) Moench), or Sphagnum (*Sphagnum capillifolium* (Ehrh.) Hedw.). Such vegetation covers were clearly associated to microtopographic differences, which in turn dictated differences in soil moisture regime, Calluna growing in the driest sites, Sphagnum in the wettest, and Molinia in sites with intermediate moisture.

Litter decomposition was followed over a period of 1 year, by placing litterbags filled with biomass from each dominant species in each type of vegetation cover. The composition of the plant material and SOM were investigated by chemical methods and solid-state ^{13}C nuclear magnetic resonance (NMR) spectroscopy.

Litter decomposition was faster for Molinia and Calluna, irrespective of the vegetation cover of the site where they were placed. Sphagnum litter decomposed very slowly, especially under Calluna, where the soil environment is by far more oxidising than under itself. In terms of SOM quality, Calluna covered areas showed the greatest differences from the others, in particular a much higher contribution from lipids and aliphatic biopolymers, apparently related to biomass composition.

Our findings showed that in the studied environment litter decomposition rate and SOM composition are actually dependent on vegetation cover and/or soil drainage. On this basis, monitoring changes in the patchwork of vegetation types in boreal heathlands could be a reliable cost-effective way to account for climate change induced modifications to SOM and its potential to last.

1. Introduction

Heathland vegetation covers approximately 60% of Norway’s land area. Norwegian heathland soils, although scant and shallow, are so rich in organic matter that they represent a stock of carbon (C) at least one order of magnitude larger than the aboveground vegetation they sustain (Rosberg et al., 1981). To predict the ecological effects of climate and land use changes, it is essential to understand the nature and environmental dependencies of soil organic matter (SOM) in these widespread systems. In fact, any change influencing their SOM stocks and dynamics may have major consequences for both C balance and the water quality of lakes and rivers (Stuanes et al., 2008).

Following changes in SOM stocks is not a simple task, and several approaches have been proposed for this purpose (*e.g.,* Johnson and Curtis, 2001; Trumbore, 2009; Chiti et al., 2011). The current common belief is that environmental and biological factors predominate on the molecular structure in controlling SOM stability (Kleber, 2010; Kleber et al., 2011; Schmidt et al., 2011); however, in some environments, vegetation cover is a good proxy for soil C dynamics, since it controls the input and quality of litter (De Deyn et al., 2008). In turn, vegetation depends, among other factors, on soil drainage, which also influences litter decay and SOM transformation (Wickland et al., 2010), so representing another possible proxy for SOM storage.

Although present-day vegetation may be different from the one the underlying SOM originated from (Chambers et al., 1999; Hjelle et al., 2010), many studies have demonstrated that the most active part of SOM is the youngest (*e.g.,* Leavitt et al., 1996; Trumbore, 2000; Chiti et al., 2009). Trumbore (2000) found that the average age of the carbon dioxide (CO_2) released by decomposition processes in boreal forest soils is 30 years, and 50-60% of total soil respiration arises from SOM with mean residence time less than 1 year. The dominant

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contribution of recently synthesized organic matter to soil respiration was also assessed by Certini et al. (2003) for forests in temperate regions. Theoretically, the moister and colder the pedoclimate, the better preserved the dead biomass in soil (Hobbie et al., 2000; Hicks Pries et al., 2013). Hence, the wet boreal heathlands are environments where the investigation of a possible relationship between vegetation covers and SOM dynamics is particularly meaningful. Here, due to the intense leaching, lost dissolved organic C (DOC) may be much older than the respired C (Karltun et al., 2005), rendering any possible relationship between present day vegetation and bulk SOM quality less clear. Nonetheless, in the uppermost soil, where SOM is younger and less degraded than below, such relationship is expected to be strong enough.

In Southern Norway, heathland areas are in most cases characterised by the alternate occurrence – essentially dictated by the soil drainage, in turn controlled by topography, particle size distribution, and soil depth to bedrock – of three vegetation types, which are dominated by the heather Calluna (*Calluna vulgaris* (L) Hull), the moor grass Molinia (*Molinia caerulea* (L) Moench), and the peat moss Sphagnum (*Sphagnum capillifolium* (Ehrh.) Hedw.). Such different vegetation types are cause and effect of the properties and behaviour of the underlying soil. This is undoubtedly true for the soil profile morphology and the sequence of horizons, generally ranging from the O-E-Bhs soil sequum of Calluna-sustaining podzols to multiple H horizons consisting histosols where Sphagnum grows (Strand et al., 2008).

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In this study we report an *in situ* investigation of the relationships between vegetation cover, initial litter decay rate and soil organic matter composition for a typical montane heathland area in Southern Norway where the alternation between Calluna, Molinia, and Sphagnum occurs on decametric scale. The objective of the study was to assess whether in this environment the current vegetation cover is a good proxy for SOM quality and dynamics. To this end, litter decomposition was followed over a period of 1 year, by placing litterbags filled with biomass from each dominant species under each type of vegetation cover, so as to simulate the effects of possible climate change induced shift of vegetation on early stages of litter decomposition. Furthermore, the composition of the aboveground biomass and the bulk SOM were investigated by chemical methods and solid-state ^{13}C nuclear magnetic resonance (NMR) spectroscopy.

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2. **Materials and Methods**

2.1. Study site

The study area, Storgama (59°02’47”N, 8°39’37”E), is located in the Telemark county, southern Norway, at an elevation of 560-m above sea level. The mean annual precipitation in Storgama for the period 1961-1990 was 994 mm, the mean annual air temperature for the same period was 5.0 °C. Approximately 30 % of the area is barren granite bedrock and boulders, and soil often occurs as pockets in depressions in the bedrock surface (Fig. 1a). The average soil depth generally varies between 10 and 35 cm but greater thicknesses, up to 100 cm, do occur. According to the U.S. Soil Taxonomy (Soil Survey Staff, 2010) and moving from drier to wetter locations, soils are Lithic Haplorthods, Lithic Udipsamments, Lithic Endoaquents, and Lithic Haplosaprists. Although there are some scattered or vaguely grouped Scots pines (*Pinus sylvestris* L) and Downy birch trees (*Betula pubescens* Ehrh), the vegetation is largely dominated by heather (*Calluna vulgaris* (L) Hull) at well drained sites, peat moss (*Sphagnum capillifolium* (Ehrh.) Hedw.) at poorly drained sites, and moor grass (*Molinia caerulea* (L) Moench) at intermediately drained sites (Figs. 1a and 1b). These dominant vegetation types are interspersed in the area, forming a patchwork dictated by topography, which in turn is a driving factor of water supply. At the Calluna sites *Calluna vulgaris* was virtually 100 % of the vegetation cover. At the Molinia sites some Calluna,

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Erica (*Erica tetralix* L), and Nartecium (*Narthecium ossifragum* (L) Huds) were associated with *Molinia caerulea* but, on a visual basis, amounting to no more than 5_% of the total cover. At the Sphagnum sites, *Sphagnum capillifolium* covered the entire surface except for a few scattered individuals of Molinia, Erica and Calluna. Hereafter, we will refer to such vegetation assemblages simply as Calluna, Molinia and Sphagnum, respectively. Further pictures and information on vegetation and soils at Storgama are reported in Strand et al. (2008).

2.2. Vegetation sampling and analysis

Three sampling sites per dominant vegetation were chosen within an area of approximately one hectare. At each location, three soil pits were dug down to bedrock, which was 35 to 50 cm deep. All the vegetation above the pit had been previously sampled and divided according to species. In the case of Calluna, the woody stems and branches were separated from the leaves and flowers. Capitula and the five upper centimetres were used to represent the whole Sphagnum material. Visible roots were picked out from the soil samples and separated according to species when possible. The aboveground biomass and the roots were analysed for C and N by dry combustion on oven-dried (60 °C to constant weight) and finely ground samples using a LECO® CHN1000 Analyser. The aboveground biomass also underwent NMR investigation.

2.3. Soil sampling and analysis

We focused our attention on the uppermost soil layer, where we expected the closest relationship between SOM quality and current vegetation. Two undisturbed soil samples, to be used for soil solution extraction, were taken by completely inserting 7.0 cm high and 4.6 cm in inner diameter, rigid cylinders at about five cm depth in each soil profile. The filled cylinders were carefully extracted from the soil and placed in a cooling box after sealing the ends with plastic lids. The samples were stored at 4 °C, for a maximum of one week, until they were processed further. Two standard disturbed soil samples were taken near the holes left by the cylinders and used for C, N, and pH determination, and NMR analysis. As for the plant material, soil C and N concentrations were measured by dry combustion on oven-dried and ground samples, while soil pH was determined potentiometrically in a 1:2.5 V/V distilled water suspension.

The soil-containing cylinders were inserted in two-compartment buckets and centrifuged at 4620 *g* for 20 min, following the method described by Giesler et al. (1996). The obtained solution was filtered through a 0.45 µm membrane filter (Millipore). An aliquot of the filtrate was analysed for total C (Shimadzu TOC-V element analyser) and, after oxidation by peroxodisulphate (NS4743 1975), for total N (FiaSTAR, Tecator Spectrophotometer system). Another aliquot of the filtrate was used to measure hydrophobicity, by determining the ratio between the absorbances of the solution at 285 and 254 nm using an UV-VIS spectrophotometer (UV-1201 Shimadzu). These two absorbances are, in fact, correlated to hydrophobic C (π-π* electron transitions occur at ~285 nm for a number of aromatic substances, as described in Chin et al. 1994) and total C (Brandstetter et al., 1996), respectively.

After centrifugation the soil was immediately passed through a 2 mm-mesh sieve. Two grams of the moist sieved soil was treated as in the second step of the procedure proposed by Ghani et al. (2003) to obtain hot-water extract (80 °C for 16 h). After centrifugation for 20 min at 2,000 *g* and filtration through 0.45 µm membrane filters (Millipore), the extract was analysed for total C (HWC), total N (HWN), and carbohydrate C (Carb-C). HWC and HWN were determined by the same method as DOC and total dissolved nitrogen (TDN), while the analysis of Carb-C was done according to the "direct determination" method proposed by

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Safarík and Santrucková (1992). In brief, 1 mL of the extract was combined in a polyethylene tube with 1 mL 5% phenol solution and 5 mL concentrated sulphuric acid and immediately shaken on a vortex mixer. The absorbance of the mixture was read after 1 h at 485 nm on a UV-VIS spectrophotometer (UV-1201 Shimadzu). A calibration curve was built with the following standards: 0.00, 0.05, 0.10, 0.25, 0.40 g L^{-1} of α-D glucose (R^2=0.9907).

2.5. Nuclear magnetic resonance spectroscopy

The chemical structure of the aboveground vegetation (one composite sample per dominant species, after removal of stems and coarse branches in the case of Calluna) and SOM (one composite sample per soil pit; hence, three samples per vegetation type) was investigated by solid-state ^{13}C nuclear magnetic resonance (NMR) spectroscopy using the cross polarization with magic angle spinning (CP MAS) technique. Prior to analysis, soil samples underwent 2 % HF treatment according to Skjemstad et al. (1994) in order to remove possible paramagnetic oxides, which cause broadened resonances and signal loss. NMR spectra were obtained by a Bruker AMX 300-WB spectrometer equipped with a 4 mm CP MAS probe. The operating frequencies were 300.13 and 75.47 MHz for ^{1}H and ^{13}C, respectively; the $\pi/2$ pulse was 3.4 μs on the ^{1}H channel. A contact time of 2 ms and a relaxation delay of 4 s were used. The MAS speed was set to 8 kHz and the number of scans recorded ranged between 4800 and 40000, depending on the sample. The chemical shifts were referenced to tetramethylsilane (TMS) using adamantane as external standard. Seven chemical-shift regions of the NMR-spectra, corresponding to the main C forms, were integrated and expressed as percent contribution to total area subtended by the spectrum between 0 and 220 ppm. The seven regions account for alkyl C (0-45 ppm, mainly comprising lipids, waxes, resins, suberin), methoxyl and N-alkyl C (45-60 ppm, comprising the methoxy group of guaiacyl and the two methoxy groups of syringyl lignin moieties at ~56 ppm), O-alkyl C (60-90 ppm,

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carbohydrates, mainly cellulose and hemicellulose, with contributions from carbohydrate carbons bonded to one oxygen), di-O-alkyl C (90-110 ppm, mainly from polysaccharides, with contributions from anomeric carbons of carbohydrates, *i.e.*, bonded to two oxygens), H- and C-substituted aromatic C (110-140 ppm), O-substituted aromatic C (140-162 ppm, mainly from lignin structures, tannins, polyphenols), and carboxyl C (162-190 ppm, esters, acids and amides); no carbonyl intensity in the 190-220 ppm region, ascribable to aldehydes and ketones, was detected.

2.6. Litter decomposition

Litter decomposition was determined *in situ* by the litterbag technique. Recently formed aboveground biomass of Calluna, Molinia, and Sphagnum (approximately, the top 5 cm), were collected at the end of the growing season in late September. This material was oven dried (35 °C to constant weight) and used for filling 10 x 12 cm nylon mesh bags (0.5-1 mm mesh), with 3.0 g Calluna, 2.0 g Molinia, or 1.0 g Sphagnum. In November, 32 litterbags of each vegetation type were installed on the surface of each sampling site, except Calluna under Sphagnum, since a substitution of Calluna by Sphagnum was judged to be highly improbable. Eight to ten litterbags per type of content were sampled from each site after 6, 9 and 12 months of decomposition. The removed litterbags were cleaned of plant remnants and other minor foreign material by gently using a soft brush, oven dried (35 °C to constant weight) and weighed for determining mass loss. Their content was thus ground and analysed for carbon and nitrogen as described for the vegetation and soil samples.

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2.7. Statistics

All statistical analyses were performed using the software program SAS (SAS Institute, Inc., 1990, Cary, NC). After checking the dataset for normality and variance heterogeneity, effects of vegetation and decomposition site on soil pH and SOM were tested by one-way analysis of variance (General Linear Model, GLM). Two separate one-way ANOVAs were performed on litterbags data: one assessing differences in decomposition rate between the three litter types disregarding the dominant vegetation effect, the other assessing differences between the vegetation types disregarding the litter effect. Pairwise comparisons were done by the Tukey's Simultaneous test.

3. Results

The experimental data set indicated marked differences in composition among the dominant plant species. The C concentration in the aboveground biomass increased in the order Sphagnum<Molinia<Calluna, whereas the C/N ratio increased in the order Molinia<Sphagnum<Calluna, with most interspecific differences being significant (Table 1). Belowground, Calluna, and Molinia also showed different composition, hence reflecting the aboveground biomass (Table 1).

Concerning the soil, the measured pH values, all much below neutrality (Table 2), ensured that all C there present was in organic forms. The N content of Sphagnum was significantly higher than that of Calluna, although there were no differences in terms of C/N ratio. The latter, clustering around 20, was in all cases much smaller than the C/N ratio of the respective dominating plant species.

There was a large variability in soil DOC and TDN concentrations, and vegetation types did not show any significant difference with respect to these two variables (Table 2). On the contrary, the hydrophobicity index was significantly different in soils under the three types of vegetation, being highest for Calluna and lowest for Molinia. This difference indicates that a greater proportion of DOC under Calluna was hydrophobic. For the rest, the only other significant differences were in terms of HWN and HW-C/N ratio between Calluna, on the

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one hand, and Molinia and Sphagnum, on the other (Table 2).

The ^{13}C CPMAS NMR spectra of the aboveground biomass and soil are shown in Fig. 2, and the relative contributions of the different chemical shift regions are reported in Table 3. The NMR spectra of the aboveground vegetation suggested more similar compositions for Molinia and Sphagnum with respect to Calluna. The spectrum of the Calluna biomass was dominated by signals between 60 and 104 ppm, characteristic of polysaccharides; the relatively high intensity in the alkyl C region (0-50 ppm) was due to lipids and aliphatic biopolymers. The spectrum also revealed the presence of lignin and tannins, as indicated by the lignin methoxyl carbon signal at 56 ppm, and the distinct aromatic peaks at 145 and 155 ppm, typical of condensed tannins. The sharp peak at 172 ppm is normally assigned to the carboxyl C of hemicellulose esters, but may also have contributions from amides (Forte et al., 2006). The spectra of Molinia and Sphagnum aboveground biomasses showed the same dominant polysaccharide features of Calluna in the 50-110 ppm range, but a significantly lower intensity of signal in the alkyl and aromatic C regions, which means lower contribution of lipids and lignin/tannins, respectively. In the case of Molinia, the slightly narrower signals in the 60-100 ppm region and the relatively smaller peak shoulder at about 103 ppm compared with both Calluna and Sphagnum, suggested the occurrence of less hemicellulose and some crystalline cellulose, respectively. Sphagnum did not show the typical lignin signals, in agreement with the common lignin-free composition of bryophytes (Kļaviņa et al., 2012). The only aromatic signals in the sphagnum spectrum were due to unsubstituted or C-substituted aryl C at 130 and 117 ppm, while the signal at 158 ppm was ascribable to phenolic structures. In the case of Calluna, the two sharp tannin peaks at 145 and 155 ppm observed in the aboveground biomass spectrum were totally absent in the SOM spectrum. In the case of Molinia, differently from the other two vegetation types, the relative contribution of aromatic C significantly increased in soil compared to that observed in the aboveground

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biomass. In the case of Sphagnum, no major changes occurred in the aromatic region, except for the absence in the soil spectrum of the signal at 158 ppm detected for the aboveground vegetation (Fig. 2). The alkyl C/O-alkyl C ratio increased for all vegetation types on passing from the intact biomass to its decomposition products in soil (Table 3), with large differences in absolute values between Calluna, on the one side, and Molinia and Sphagnum, on the other side.

The *in situ* decomposition study using litterbags showed that the litter mass remaining after one year of decomposition varied between 62 and 66% in the case of Molinia and Calluna and 83 and 94% for Sphagnum (Fig. 3). The discrepancy between the couple Calluna-Molinia and Sphagnum was lower, although significant, in the intermediate stages of the experiment. After six months, Calluna showed significantly lower mass loss than Molinia under itself, while at the end of the experiment Calluna resulted to be better preserved than Molinia only under Molinia (Fig. 3).

In terms of relative C content of the residual litter, Calluna did not change throughout the 12 months of the experiment, while Molinia and Sphagnum experienced a marked decrease compared to the original value (Fig. 4). Relative concentrations of N in the litter changed more than the C ones. Except for Sphagnum under itself or under Molinia, all litters increased their N content from November to May; later, all of them increased until August, with the exception of Sphagnum under Calluna and Molina under itself; finally, in the period from August to November, N concentration continued to increase in Calluna, whereas it decreased in Molinia and showed an irregular trend in Sphagnum (Fig. 4). These C and N trends implied progressive, although slight, decrease in C/N ratio for Calluna and Sphagnum, and a sharper decrease for the same ratio for Molinia until August, after which it increased (Fig. 4). Contrary to Calluna and Sphagnum, Molinia degraded maintaining significantly higher values of C/N ratio under Sphagnum than under the other types of vegetation (Molinia and

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Calluna). At the end of the experiment, in November, the C/N ratio in Molinia under Sphagnum was even higher than the original value.

4. Discussion

In the heathland environment of Storgama, the composition of SOM appeared to partly reflect that of the parent vegetation. Hence, for example, the abundance in alkyl C in the Calluna biomass relative to the other two vegetation types was transferred to the SOM. Nevertheless, SOM accumulated over a long period of time; as a consequence, it could be the result of multiple changes in vegetation cover in the area and, thus, be partly unrelated to the current vegetation cover. Actually, there was no direct or indirect evidence in this regard, but inputs of wind-blown or water-transported material cannot be excluded at any site.

Sphagnum showed a composition potentially more prone to decay than Calluna and Molinia. Nevertheless, there were no significant differences in the SOM content of the topsoil of the three vegetation covers. Evidently, the prevailing anoxic conditions limited decomposition at the Sphagnum sites. This is in accordance with several studies that used the type of vegetation cover as a proxy for carbon dynamics, based on the consideration that vegetation chiefly reflects the soil moisture regime (Bridgham et al., 2008; Couwenberg et al., 2011; Delarue et al., 2011), which is in turn a driving factor of litter decomposition (Hobbie et al., 2000; Laiho 2006). Large variability in DOC concentrations and no significant effect of vegetation was observed (Table 2). It must be noted, however, that our study shows the conditions only at one sampling occasion, *i.e.* at the end of the growing season, when DOC concentrations are affected by a considerable contribution from senescing plant material. The measured DOC concentrations were generally in agreement with those recorded in autumn using zero tension lysimeters in soils at Storgama and other Norwegian heathland areas (Strand et al., 2002; Vestgarden et al., 2010), although DOC concentrations in centrifuged

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and freely drained soil solutions are not directly comparable (Giesler et al., 1996). Similarly to DOC, TDN showed a large variability and no apparent correlation with vegetation. The relatively small amount of water extracted by centrifugation limited the number of possible analyses, preventing N speciation. TDN therefore included both organic N and inorganic N, the latter amounting to 25-50% of TDN in soil water from southern Norway (Austnes et al., 2008; Kaste et al., 2008).

The hydrophobicity index of soil water differed significantly among vegetation types. Apparently, Calluna released DOC with the highest proportion of hydrophobic organic compounds, perhaps mostly arising from tannins and decomposition of lignin (Dilling and Kaiser, 2002), which are indeed important components of the Calluna litter (Fig. 2).

Hot water C approximately amounted to 4.5% of SOC in all samples, irrespective of vegetation. This percentage is in the range reported by von Lützow et al. (2007). Significantly lower amounts of HWN were extracted from the Calluna soils compared to the Molinia and Sphagnum ones, which also implied significantly higher HWC / HWN-ratio for Calluna (Table 2). We did not partition HWN, however Curtin et al. (2006) demonstrated that it is mainly organic and, in suborder, NH_4-N generated by hydrolysis of heat-labile organic N. The quality of the hot water extract rather well discriminated Calluna from Molinia and Sphagnum. Some authors have proposed hot water extraction of SOM as a method to measure the labile SOM pool (Chodak et al., 2003; Ghani et al., 2003; Curtin et al., 2006); however, other authors consider this method not selective enough for this purpose (Landgraf et al., 2006; von Lützow et al., 2007). In our case, approximately half the C extracted by hot water belonged to carbohydrates.

The NMR spectra showed clear structural differences in aboveground plant material (Fig. 2 and Table 3). In particular, Calluna was richer in alkyl C and poorer in O-alkyl C than Molinia and Sphagnum, as well reflected in the alkyl C / O-alkyl C ratio. The richness in

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alkyl C has been correlated to slow decomposition rates in heathland ecosystems (van Vuuren and van der Eerden, 1992; van Vuuren and Berendse, 1993). However, in our litterbags experiment there were little and variable differences between the mass losses of Calluna and Molinia, and both of them were much higher than the one in Sphagnum wherever the latter was placed (Fig. 3). A possible explanation for such short-term resistance of Sphagnum to degradation could be that this type of vegetation is particularly rich in sphagnan pectin-like polysaccharides, which, unlike the other types of polysaccharides, induce processes that prevent organic matter decay (Hájek et al., 2011; Ballance et al., 2012). Moreover, it must be considered that Sphagnum might have experienced a “non-additive” pattern of mass loss, *i.e.* a decomposition behaviour sometimes observed in litter mixes that deviates from the response predicted for the individual species because of the influence of the other species present in the mix (Gartner and Cardon, 2004). In this case, the necromasses of Molinia and, in particular, Calluna could have partly inhibited the decomposition of the Sphagnum in the litterbags.

In addition to a “vegetation effect”, the litterbags experiment showed some “site effect”, *i.e.* more rapid decomposition when litter was placed beneath the parent vegetation rather than beneath other species (Ayres et al., 2009; Perez et al., 2013; Wang et al., 2013). In fact, for Sphagnum the mass loss was significantly lower when it decayed under Calluna than under Molina or Sphagnum (Fig. 3). Calluna was better preserved under Molinia than under itself at the end of the trial, while, after six months only, Molinia litter showed significant environment-induced advantage under Sphagnum compared to under itself (Fig. 3). Unexpectedly, the well drained Calluna soils preserved Sphagnum and Molinia from decay better than the moister soils where they were growing, perhaps as an effect of a seasonal drought.

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Eliminato: led us to expect differences in decomposition rates among the three vegetation types. Overall, the NMR investigation revealed that Calluna and the related SOM were richer in alkyl C and poorer in O-alkyl C than the corresponding specimens from Molinia and Sphagnum. Actually, the apparent richness in alkyl C has been correlated to slow decomposition rates of Calluna biomass

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Eliminato: NMR spectra also revealed that the residues of all three dominant plants, once in soil, experienced a significant increase in the alkyl C contribution and a concomitant decrease in the O-alkyl C one, most probably as a result of a faster decay of carbohydrates than of other C forms and the synthesis of lipids from the biodegradation of carbohydrate and aromatic fractions (Baldock et al., 1992). Less important but noteworthy differences in the spectral features between the aboveground biomass and soil were observed in the aromatic region as well (Fig. 2 and Table 3). The alkyl C / O-alkyl C ratio, which generally increases as decomposition proceeds, was significantly higher under Calluna than under Molinia and Sphagnum. Hence, on the basis of the NMR spectra, Calluna appeared to be potentially more recalcitrant to decomposition than Molinia and Sphagnum.

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The enclosure of litter inside mesh bags may actually change its overall decomposition rate and its C and N percent concentrations compared to non-bagged substrate (Berhe, 2013). However, we must confidently assume that in our experiment such a bagging effect is uniform through the samples, also because the bags are very similar. On this basis, Molinia showed an initial C/N ratio much higher than the ones of Calluna and, especially, Sphagnum (Fig. 4), which suggested a more marked intrinsic resistance of Molinia to decay. Noteworthy is the difference in C/N ratio between the aboveground Molinia biomass analysed for basic characterisation (data of Table 1) and the Molinia used in the litterbags experiment (30 vs. circa 80). Actually, Molinia is a grass that wilts at the end of the growing season, when we sampled the material to be inserted in the bags, while the Molinia sampled for basic characterisation was still with active photosynthesis, when the C/N ratio is relatively low (Taylor et al., 2001). On the other hand, Calluna is an evergreen and no great seasonal changes in C and N concentrations occur, while Sphagnum, although it is not an evergreen, does not wilt and its C/N ratio is rather constant throughout the year. Our litterbags experiment showed that the C/N ratio is a poor predictor of decay in this environment. The anoxic conditions imposed by prolonged water saturation, commonly occurring in the Sphagnum soils and expected to have considerable influence in slackening litter decomposition, on the contrary appeared to be irrelevant in preserving organic residues during a 1-year long experiment (Fig. 3). In this regard, during a 3-year study in heathlands on *Molinia caerulea* and *Erica tetralix*, van Vuuren and Berendse (1993) did not find any site effect and litter quality appeared to be the sole driving factor. Also Scheffer et al. (2001), studying the decomposition process in fens dominated by Sphagnum species or without Sphagnum, concluded that decomposition was controlled more by intrinsic differences in litter quality than by the environment.

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The NMR investigation revealed that soils, compared to the litter they receive, showed significantly higher contribution of alkyl C and lower contribution of O-alkyl C (Fig. 2 and Table 3), most probably as a result of a faster decay of carbohydrates than of other C forms and the synthesis of lipids from the biodegradation of carbohydrate and aromatic fractions (Baldock et al., 1992). The alkyl C / O-alkyl C ratio, which generally increases as decomposition proceeds, was significantly higher in the Calluna soil than under Molinia and Sphagnum.

5. Conclusions

We found that in the varied heathland of Storgama there were many significant differences in terms of SOM composition between the Calluna dominated areas and the interspersed Sphagnum-covered areas. Most differences were clearly due to the litter quality. A “vegetation effect” on the early stage of litter decomposition rate was clear, Sphagnum remnants being much more stable independently of the environmental conditions they underwent, which differed especially in terms of soil drainage. Hence, overall, vegetation appeared to be a good proxy for SOM quality. On this basis, monitoring the distribution of vegetation types in heathlands of Norway and elsewhere could be of particular interest for assessing the consequences of climate change on SOM stocks and dynamics. In the plausible scenario of a less continuous rainfall supply and a consequent contraction of Sphagnum-covered areas, the Sphagnum-released litter seems to have good short-term ability to resist decomposition under the two replacing types of vegetation, Molinia and Calluna. Long-term experiments addressing this issue are needed.

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References

Austnes, K., Kaste, O., Vestgarden, L. S., and Mulder, J.: Manipulation of snow in small headwater catchments at Storgama, Norway: Effects on leaching of total organic carbon and total organic nitrogen. Ambio 37: 38–47, 2008.

Ayres, E., Steltzer, H., Simmons, B. L., Simpson, R. T., Steinweg, J. M., Wallenstein, M. D., Mellor, N., Parton, W. J., Moore J. C., and Wall, D.H.: Home-field advantage accelerates leaf litter decomposition in forests. Soil Biol. Biochem. 41: 606–610, 2009.

Baldock, J. A., Oades, J. M., Waters, A. G., Peng, X., Vassallo, A. M., and Wilson, M. A.: Aspects of the chemical structure of soil organic materials as revealed by solid-state ^{13}C NMR spectroscopy. Biogeochemistry 16: 1–42, 1992.

Ballance, S., Kristiansen, K. A., Skogaker, N. T., Tvedt, K. E., and Christensen, B. E.: The localisation of pectin in Sphagnum moss leaves and its role in preservation. Carbohyd. Polym. 87: 1326–1332, 2012.

Berhe, A. A.: Effect of litterbags on rate of organic substrate decomposition along soil depth and geomorphic gradients. J. Soils Sediments 13: 629–640, 2013.

Brandstetter, A., Sletten, R. S., Mentler, A., and Wenzel, W. W.: Estimating dissolved organic carbon in natural waters by UV absorbance (254 nm). Z. Pflanz. Bodenkunde 159: 605–607, 1996.

Bridgham, S. D., Pastor, J., Dewey, B., Weltzin, J. F., and Updegraff, K.: Rapid carbon response of peatlands to climate change. Ecology 89: 3041–3048, 2008.

Certini, G., Corti, G., Agnelli, A., and Sanesi, G.: Carbon dioxide efflux and concentrations in two soils under temperate forests. Biol. Fert. Soils 37: 39–46, 2003.

Chambers, F. M., Mauquoy, D., and Todd, P. A.: Recent rise to dominance of *Molinia caerulea* in environmentally sensitive areas: new perspectives from palaeoecological data. J. Appl. Ecol. 36: 719–733, 1999.

Chin, Y. P., Aiken, G., and Oloughlin, E.: Molecular-weight, polydispersity, and spectroscopic properties of aquatic humic substances. Environ. Sci. Technol. 28: 1853–1858, 1994.

Chiti, T., Certini, G., Perugini, L., Mastrolonardo, G., Papale, D., and Valentini, R.: Soil carbon dynamics in a Mediterranean forest during the Kyoto Protocol commitment periods. Reg. Environ. Change 11: 371–376, 2011.

Chiti, T., Neubert, R. E. M., Janssens, I. A., Certini, G., Curiel Yuste, J., and Sirignano, C.: Radiocarbon dating reveals different past managements of adjacent forest soils in the Campine region, Belgium. Geoderma 149: 137–142, 2009.

Chodak, M., Khanna, P., and Beese, F.: Hot water extractable C and N in relation to microbiological properties of soils under beech forests. Biol. Fert. Soils 39: 123–130, 2003.

Couwenberg, J., Thiele, A., Tanneberger, F., Augustin, J., Barisch, S., Dubovik, D., Liashchynskaya, N., Michaelis, D., Minke, M., Skuratovich, A., and Joosten, H.:

Assessing greenhouse gas emissions from peatlands using vegetation as a proxy. Hydrobiologia 674: 67–89, 2011.

Curtin, D., Wright, C. E., Beare, M. H., and McCallum F. M.: Hot water-extractable nitrogen as an indicator of soil nitrogen availability. Soil Sci. Soc. Am. J. 70: 1512–1521, 2006.

De Deyn, G. B., Cornelissen, J. H. C., and Bardgett, R. D.: Plant functional traits and soil carbon sequestration in contrasting biomes. Ecol. Lett. 11: 516–531, 2008.

Delarue, F., Laggoun-Defarge, F., Disnar, J. R., Lottier, N., and Gogo, S.: Organic matter sources and decay assessment in a Sphagnum-dominated peatland (Le Forbonnet, Jura Mountains, France): impact of moisture conditions. Biogeochemistry 106: 39–52, 2011.

Dilling, J. and Kaiser, K.: Estimation of the hydrophobic fraction of dissolved organic matter in water samples using UV photometry. Water Res. 36: 5037–5044, 2002.

Forte, C., Piazzi, A., Pizzanelli, S., and Certini, G.: CP MAS C-13 spectral editing and relative quantitation of a soil sample. Solid State Nucl. Mag. 30: 81–88, 2006.

Gartner, T. B. and Cardon, Z. G.: Decomposition dynamics in mixed-species leaf litter. Oikos 104: 230–246, 2004.

Ghani, A., Dexter, M., and Perrott, K. W.: Hot-water extractable carbon in soils: a sensitive measurement for determining impacts of fertilisation, grazing and cultivation. Soil Biol. Biochem. 35: 1231–1243, 2003.

Giesler, R., Lundstrom, U. S., and Grip, H.: Comparison of soil solution chemistry assessment using zero-tension lysimeters or centrifugation. Eur. J. Soil Sci. 47: 395–405, 1996.

Hájek, T., Ballance, S., Limpens, J., Zijlstra, M., and Verhoeven, J. T. A.: Cell-wall polysaccharides play an important role in decay resistance of Sphagnum and actively depressed decomposition in vitro. Biogeochemistry 103: 45–57, 2011.

Hicks Pries, C. E., Schuur, E. A. G., Vogel, J. G., and Natali, S. M.: Moisture drives surface decomposition in thawing tundra. J. Geophys. Res.-Biogeo. 118: 1133–1143, 2013.

Hjelle, K. L., Halvorsen, L. S., and Overland, A.: Heathland development and relationship between humans and environment along the coast of western Norway through time. Quatern. Int. 220: 133–146, 2010.

Hobbie, S. E., Schimel, J. P., Trumbore, S. E., and Randerson, J. R.: Controls over carbon storage and turnover in high-latitude soils. Glob. Change Biol. 6 (Suppl. 1): 196–210, 2000.

Johnson, D. W. and Curtis, P. S.: Effects of forest management on soil C and N storage: meta-analysis. Forest Ecol. Manag. 140: 227–238, 2001.

Karltun, E., Harrison, A. F., Alriksson, A., Bryant, C., Garnett, M. H., and Olsson, M. T.: Old organic carbon in soil solution DOC after afforestation – evidence from C-14 analysis. Geoderma 127: 188–195, 2005.

Kaste, O., Austnes, K., Vestgarden, L.S., and Wright, R. F.: Manipulation of snow in small headwater catchments at Storgama, Norway: Effects on leaching of inorganic nitrogen. Ambio 37: 29–37, 2008.

Kļaviņa, L., Bikovens, O., Šteinberga, I., Maksimova, V., and Eglīte, L.: Characterization of chemical composition of some bryophytes common in Latvia. Environ. Exp. Biol. 10: 27–34, 2012.

Kleber, M.: What is recalcitrant soil organic matter? Environ. Chem. 7: 320–332, 2010.

Kleber, M., Nico, P. S., Plante, A., Filley, T., Kramer, M., Swanston, C., and Sollins, P.: Old and stable soil organic matter is not necessarily chemically recalcitrant: implications for modeling concepts and temperature sensitivity. Glob. Chang. Biol. 17: 1097–1107, 2011.

Laiho, R.: Decomposition in peatlands: Reconciling seemingly contrasting results on the impacts of lowered water levels. Soil Biol. Biochem. 638: 2011–2024, 2006.

Landgraf, D., Leinweber, P., and Makeschin, F.: Cold and hot water-extractable organic matter as indicators of litter decomposition in forest soils. J. Plant Nutr. Soil Sci. 169: 76–82, 2006.

Leavitt, S. W., Follett, R. F., and Paul, E. A.: Estimation of slow- and fast-cycling soil organic carbon pools from 6N HCl hydrolysis. Radiocarbon 38: 231–239, 1996.

Perez, G., Aubert, M., Decaens, T., Trap, J., and Chauvat, M.: Home-Field Advantage: A matter of interaction between litter biochemistry and decomposer biota. Soil Biol. Biochem. 67: 245–254, 2013.

Rosberg, I., Ovstedal, D. O., Seljelid, R., Schreiner, O., and Goksoyr, J.: Estimation of carbon flow in a Calluna Heath System. Oikos 37: 295–305, 1981.

Safarík, I., and Santrucková, H.: Direct determination of total soil carbohydrate content. Plant Soil 143: 109–114, 1992.

Scheffer, R. A., van Logtestijn, R. S. P., and Verhoeven, J. T. A.: Decomposition of Carex and Sphagnum litter in two mesotrophic fens differing in dominant plant species. Oikos 92: 44–54, 2001.

Schmidt, M. W. I., Torn, M. S., Abiven, S., and 11 others: Persistence of soil organic matter as an ecosystem property. Nature 478: 49–56, 2011.

Skjemstad, J. O., Clarke, P., Taylor, J. A., Oades, J. M., and Newman, R. H.: The removal of magnetic-materials from surface soils – A solid-state C-13 Cp/Mas nmr-study. Aust. J. Soil Res. 32: 1215–1229, 1994.

Soil Survey Staff: Keys to Soil Taxonomy. 11th Edition. USDA Natural Resources Conservation Service. Washington, DC., 2010.

Strand, L. T., Abrahamsen, G., and Stuanes, A. O.: Leaching from organic matter-rich soils by rain of different qualities: I. Concentrations. J. Environ. Qual. 31: 547–556, 2002.

Strand, L. T., Haaland, S., Kaste, Ø., and Stuanes, A. O.: Natural variability in soil and runoff from small headwater catchments at Storgama, Norway. Ambio 37: 18–28, 2008.

Stuanes, A. O., de Wit, H. A., Hole, L. R., Kaste, O., Mulder, J., Riise, G., and Wright, R. F.: Effect of climate change on flux of N and C: Air-land-freshwater-marine links: Synthesis. Ambio 37: 2–8, 2008.

Taylor, K., Rowland, A. P., and Jones, H. E.: *Molinia caerulea* (L.) Moench. J. Ecol. 89: 126–144, 2001.

Trumbore, S.: Age of soil organic matter and soil respiration: Radiocarbon constraints on belowground C dynamics. Ecol. Appl. 10: 399–411, 2000.

Trumbore, S.: Radiocarbon and soil carbon dynamics. Annu. Rev. Earth Pl. Sci. 37: 47–66, 2009.

van Vuuren, M. M. I. and Berendse, F.: Changes in soil organic-matter and net nitrogen mineralization in heathland soils, after removal, addition or replacement of litter from *Erica-tetralix* or *Molinia-caerulea*. Biol. Fert. Soils 15: 268–274, 1993.

van Vuuren, M. M. I. and van der Eerden, L. J.: Effects of 3 rates of atmospheric nitrogen deposition enriched with n-15 on litter decomposition in a heathland. Soil Biol. Biochem. 24: 527–532, 1992.

Verhoeven, J. T. A. and Toth, E.: Decomposition of carex and sphagnum litter in fens – effect of litter quality and inhibition by living tissue-homogenates. Soil Biol. Biochem. 27: 271–275, 1995.

Vestgarden, L. S. and Austnes, K.: Effects of freeze-thaw on C and N release from soils below different vegetation in a montane system: a laboratory experiment. Glob. Chang. Biol. 15: 876–887, 2009.

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Vestgarden, L. S., Austnes, K., and Strand, L. T.: Vegetation control on DOC, DON and DIN concentrations in soil water from a montane system, southern Norway. Boreal Environ. Res. 15: 565–578, 2010.

von Lützow, M., Kögel-Knabner, I., Ekschmitt, K., Flessa, H., Guggenberger, G., Matzner, E., and Marschner, B.: SOM fractionation methods: Relevance to functional pools and to stabilization mechanisms. Soil Biol. Biochem. 39: 2183–2207, 2007.

Wang, Q. K., Zhong, M., He, T.: Home-field advantage of litter decomposition and nitrogen release in forest ecosystems. Biol. Fert. Soils 49: 427–434, 2013.

Wickland, K. P., Neff, J. C., and Harden, J. W.: The role of soil drainage class in carbon dioxide exchange and decomposition in boreal black spruce (*Picea mariana*) forest stands. Can. J. Forest Res. 40: 2123–2134, 2010.

Figures captions:

Figure 1. a) A general view of the study area, Storgama, showing soil occurring in pockets and small depressions at the bedrock surface; note that close up vegetation at the bottom right is dominated by *Molinia caerulea* (L), the understorey of pines beyond is *Calluna vulgaris* (L) Hull, the basin in the background is covered by *Sphagnum* spp. **b**) A rare coalescence of the three dominant species, *Calluna vulgaris*, on the left, *Sphagnum* spp. L, at the bottom, and *Molinia caerulea*, on the right.

Figure 2. ^{13}C CPMAS NMR spectra of the aboveground biomass of the dominant plant species and soil.

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Figure 3. Residual mass in the litterbags as a function of time for different combinations of litter and vegetation cover. Cal in Cal means *Calluna* litter decomposing under *Calluna*, Cal in Mol means *Calluna* litter decomposing under *Molinia*, and so on. Error bars are standard deviations, while lower case letters indicate significant differences (p<0.05) between same litters decomposing under different types of vegetation. The trial was one year long.

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Figure 4. Carbon and nitrogen concentrations and C/N ratio in decaying biomass in the litterbags as a function of time for different combinations of litter and vegetation cover. Cal in Cal means *Calluna* litter decomposing in soil under *Calluna*, Cal in Mol means *Calluna* litter decomposing under *Molinia,* and so on. Upper case letters indicate significant differences (p<0.05) between different litters, whereas lower case letters indicate significant differences between same litters decomposing in soils covered by different types of vegetation.