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# Comprehensive increase in CO<sub>2</sub> release by drying–rewetting cycles among Japanese forests and pastureland soils and exploring predictors of increasing magnitude

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Received: 20 February 2024 – Discussion started: 9 April 2024 Revised: 7 October 2024 – Accepted: 26 November 2024 – Published: 16 January 2025

**Abstract.** It is still difficult to precisely quantify and predict the effects of drying–rewetting cycles (DWCs) on soil carbon dioxide  $(CO_2)$  release due to the paucity of studies using constant moisture conditions equivalent to the mean water content during DWC incubation. The present study was performed to evaluate overall trends in the effects of DWCs on CO<sub>2</sub> release and to explore environmental and soil predictors for variations in the effect size in 10 Japanese forests and pastureland soils variously affected by volcanic ash during their pedogenesis. Over an 84 d incubation period including three DWCs, CO<sub>2</sub> release was 1.3- to 3.7-fold greater than under continuous constant moisture conditions (p < 0.05) with the same mean water content as in the DWC incubations. Analysis of the relations between this increasing magnitude of CO<sub>2</sub> release by DWCs (IF<sub>CO<sub>2</sub></sub>) and various environmental and soil properties revealed significant positive correlations between IF<sub>CO2</sub> and soil organo-metal complex contents (p < 0.05), especially pyrophosphate-extractable aluminum (Alp) content (r = 0.74). Molar ratios of soil total carbon (C) and pyrophosphate-extractable C (Cp) to Alp contents and soil-carbon-contentspecific CO<sub>2</sub> release rate under continuous constant moisture conditions (qCO<sub>2</sub>\_soc) were also correlated with  $IF_{CO_2}$  (p < 0.05). The covariations among Alp, total Cp/Alp, and Cp/Alp molar ratios and qCO<sub>2</sub>\_soc suggested Alp to be the primary predictor of IF<sub>CO2</sub>. Additionally, soil microbial biomass C and nitrogen (N) levels were significantly lower in DWCs than under continuous constant moisture conditions, whereas there was no significant relation between the microbial biomass decrease and IF<sub>CO2</sub>. The present study showed a comprehensive increase in soil CO<sub>2</sub> release by DWC in Japanese forests and pastureland soils, suggesting that Alp is a predictor of the effect size, likely due to vulnerability of organo-Al complexes to DWC.

## 1 Introduction

There is accumulating evidence of climate-change-induced alterations in global water cycling (IPCC, 2021; Allan et al., 2020; Dai, 2012; Donat et al., 2016; Pfleiderer et al., 2019). Of the consequent water regime changes, decreasing precipitation frequency (e.g., number of rainy days) and increasing intensity (e.g., number of heavy rainy days) are becoming more frequent (Dai, 2012; Donat et al., 2016), although decadal trends in annual precipitation levels are not significant over global scales (IPCC, 2021). For example, the annual number of precipitation days in Japan has decreased by 15% during the past 120 years, whereas the annual number of heavy precipitation days (more than 100 mm in a day) has increased by 26 % (Ministry of Education, Science, and Technology (MEXT) and Japanese Meteorological Agency (JMA), 2020). There is a non-significant trend in annual precipitation level during the same period (MEXT and JAM, 2020). This changing pattern in precipitation is often observed in the temperate region of the Northern Hemisphere (IPCC, 2021) and is related to increased fluctuation of soil water environments, especially DWCs, and consequent alterations in ecosystem functions (Borken and Matzner, 2009; Jin et al., 2023; Zhang et al., 2020, 2023).

Carbon dioxide (CO<sub>2</sub>) release from soil is an ecosystem process that is sensitive to DWCs (Birch, 1958; Borken and Matzner, 2009; Lee et al., 2002; Nagano et al., 2019; Unger et al., 2010, 2012; Zhang et al., 2020, 2023) and has substantial feedback potential to the ongoing climate change due to its magnitude reaching as much as 7 times greater than anthropogenic CO<sub>2</sub> emission on a global scale (Bond-Lamberty and Thomson, 2010; Friedlingstein et al., 2020). The effects of DWCs on soil CO2 release were first shown by Birch (1958) as the marked increase in soil organic matter (SOM) decomposition and CO<sub>2</sub> release after the rapid rewetting of dried soil and has since been the subject of intensive investigation (Borken and Matzner, 2009; Kpemoua et al., 2023; Lee et al., 2002, 2004; Miller et al., 2005; Nagano et al., 2019; Unger et al., 2010, 2012; Xiang et al., 2008), including meta-analyses (Kim et al., 2012; Jin et al., 2023; Zhang et al., 2020). However, it is still difficult to precisely quantify and predict the effects of DWCs on soil CO<sub>2</sub> release.

The significant uncertainties in the effects of DWCs on soil CO<sub>2</sub> release include the inconsistent trends and sizes of effects likely due to the paucity of studies using constant moisture conditions equivalent to the mean water content during DWC incubation (Kpemoua et al., 2023; Zhang et al., 2020, 2023). According to a meta-analysis by Zhang et al. (2020) using 208 data from 34 sites in 29 reports, the effects of DWCs vary according to soil and water contents in continuous constant moisture conditions. In particular, changes in CO<sub>2</sub> release rate associated with DWCs ranged from -4% to +19% with an average of +4% in comparison with the medium level of constant moisture content, which should be the same as the mean water content during DWC incubation,

whereas only 9 of 38 data representing CO<sub>2</sub> release rates were measured for such a medium level of constant moisture conditions (Zhang et al., 2020). Another 29 data were calculated as the average of two CO<sub>2</sub> release rates at the wettest and driest water contents of constant moisture conditions, which should be the same as the maximum and minimum water contents in DWC treatment, respectively (Zhang et al., 2020). In the experiment using three Alfisols from Chinese long-term experimental field studies, Zhang et al. (2023) showed similar or somewhat lower CO<sub>2</sub> release in the DWC compared with the constant moisture conditions with the same mean water content for the DWC incubation. Using two Luvisols from French long-term field experiment sites, Kpemoua et al. (2023) also showed similar features of changes in CO<sub>2</sub> release associated with DWCs, indicating the need for further comparison of CO<sub>2</sub> release between DWCs and constant moisture conditions with the same mean water content. In contrast to these studies, Nagano et al. (2019) found a 49% increase in CO<sub>2</sub> release rate associated with DWCs in an Andisol collected from a Japanese forest. This increase was more than double that of another non-volcanic ash soil from the same forest. Thus, there are substantial variations in trends of effects of DWCs in comparison with constant moisture conditions having the same mean water content during incubation, leaving knowledge gaps about environmental and soil predictors for variations in effect sizes. There are roughly three proposed mechanisms for CO<sub>2</sub> release increase by DWCs (Schimel, 2018; Barnard et al., 2020): (i) increase in available carbon source via the releases of cellular metabolites from microbial cells destroyed by rewetting after the strong drought, (ii) increase in available carbon source by the releases of carbon from macroaggregates destroyed by repeated DWC, and (iii) changes in the microbial communities in response to transient moisture conditions. Nevertheless, there are still substantial knowledge gaps for critical mechanisms or the relative importance of those mechanisms among multiple soils.

We perform the present study to evaluate overall trends in the effect of DWCs on soil CO<sub>2</sub> release and to explore the predictors of variations in its effect size among 10 Japanese forests and pastureland soils. These soils are variously affected by volcanic ash during their pedogenesis and, therefore, include several Andisols, which are known to have a high SOM storage capacity (Morisada et al., 2004), likely due to the protection of SOM from microbial decomposition by abundant reactive minerals and metals in these soils (Asano and Wagai, 2014; Imaya et al., 2007; Shirato et al., 2004). Reactive minerals and metals that contribute to the protection of SOM are iron (Fe) and aluminum (Al), constituting short-range-order minerals and organo-metal complexes (Asano and Wagai, 2014; Rasmussen et al., 2018; Shirato et al., 2004; Wagai et al., 2018). Although global coverage of Andisols is about 1 % (FAO/IIASA/ISRIC/ISS-CAS/JRC, 2009), determination of the responses of carbon cycling in Andisols to DWCs will help in understanding the



Figure 1. Locations of Japanese forests and pastureland where investigated soils were collected.

responses of non-volcanic ash soils because reactive minerals and metals are also essential in high carbon stocks of those soils (Rasmussen et al., 2018; Hall and Thompson, 2022) and may be sensitive to climate and land use management, including water regime (Georgiou et al., 2022; Kramer and Chadwick, 2018).

#### 2 Materials and methods

### 2.1 Site description and soil sampling

We collected 10 soil samples from depths of 0-5 or 0-10 cm in six forests and a pastureland located in Niigata (six soils from four forests), Ibaraki (two soils from a forest) (Nagano et al., 2019), and Oita (two soils from a forest and a pastureland) (Wijesinghe et al., 2021) prefectures in Japan. Figure 1 and Table 1 present the locations and site characteristics, i.e., elevation, mean annual temperature (MAT), mean annual precipitation (MAP), potential evapotranspiration (PET), and net primary production (NPP). Briefly, all of the investigated sites have a humid temperate climate with MAT of 9.1-10.8 °C and MAP of 1474-2930 mm. All of the forests are dominated by beech (Fagus crenata and Fagus japonica) and oak (Quercus serrata), except for Oita forest which is a deciduous/evergreen mixed forest dominated by Siebold's maple (Acer sieboldianum), Japanese snowbell (Styrax japonicus), and Japanese holly (Ilex crenata). The pastureland in Oita is dominated by Japanese lawn grass (Zoysia japonica), dwarf fountain grass (Pennisetum alopecuroides), cranesbill (Geranium thunbergii), white clover (Trifolium repens), and Indian strawberry (Potentilla indica).

Soil sampling was conducted in the snow-free season (April to October) of 2021. We also collected soil samples from layers below the target depth (i.e., 0–5 or 0–10 cm depth) down to 50 cm as the maximum depth to examine whether the soil could be classified as Andisol. According to the USDA (United States Department of Agriculture) Soil Taxonomy criteria (USDA Soil Survey Staff, 2022), soils

with 60% or more of the thickness containing more than 20 mg of acid-oxalate-extractable Al (Alo) and 1/2 Fe (Feo) per 1 g soil within a depth of 0-60 cm are classified as Andisols. According to these criteria, 4 of the 10 soils were Andisols (one each from Niigata and Ibaraki and two from Oita), with non-allophanic properties determined by a high ratio of pyrophosphate-extractable Al (Alp) content to Alo content (Alp/Alo > 0.5; USDA Soil Survey Staff (2022); Fig. S1 in the Supplement). Collected soil samples were transferred to the laboratory and stored at 4 °C before further analysis. Before analysis, soils were gently passed through a 4 mm sieve to remove gravel and plant tissue. Fine roots in the sieved samples were removed with tweezers. In our study, we considered that the soil water content at the soil sampling reflected the ability of soil to hold the water and thus the usual water contents in the field because the soil water content showed significantly positive correlation with water-holding capacity (WHC) (r = 0.87, p < 0.01). Therefore, CO<sub>2</sub> release rate for constant moisture conditions in the present study should represent the release rate under the usual field moisture conditions of each soil.

### 2.2 Soil analysis

The soil properties analyzed were  $pH(H_2O)$ ; electrical conductivity; water content; water-holding capacity (WHC); total carbon (C) and nitrogen (N) contents; particle size distributions as relative compositions of clay, silt, and sand-sized particles; and selectively dissolved Al and Fe contents (Tables 2-4). The pH(H<sub>2</sub>O) was measured in soil and water mixtures consisting of 1 g of soil and 2.5 mL of water. For measurement of electrical conductivity, 5 mL of water was added to 1 g of soil. Water content was measured by determining the difference in soil weight before and after drying at 105 °C for 24 h. WHC was measured as the difference in soil weight before and after water saturation referring to the Hilgard method (Mabuhay et al., 2003; Ahn et al., 2008). Here, water contents when soil is completely saturated in the Hilgard method should equal zero pF value (0 kPa) as soil water potential. Soil total C and N contents were measured for airdried and well-ground soil samples using an elemental analyzer (vario PYRO cube; Elementar, Manchester, UK). Particle size distributions were determined using the sedimentation method based on Stokes' law (Miller et al., 1988; Nakai, 1997) using soil mineral particles after removing organic matter by hydrogen peroxide solution digestion. Selectively dissolved metals such as reactive Al and Fe were measured according to the procedure described previously in Nagano et al. (2023). Briefly, the contents of Al and Fe extractable with 2.0 M acid ammonium oxalate (i.e., Alo and Feo, respectively) were measured as contents of organo-metal complexes and short-range-order minerals, while Al and Fe extractable with 0.1 M sodium pyrophosphate solution (i.e., Alp and Fep, respectively) were measured as contents of organo-metal complexes (Asano and Wagai, 2014; Takahashi

| Prefecture | Soil              | Latitude<br>° N | Longitude<br>° E | Elevation<br>m | MAT<br>°C | MAP<br>mm | Annual PET<br>mm | MAP – PET<br>mm | Annual NPP<br>gCm <sup>-2</sup> |
|------------|-------------------|-----------------|------------------|----------------|-----------|-----------|------------------|-----------------|---------------------------------|
| Niigata    | Aga1, Aga2        | 37.55           | 139.51           | 474            | 10.6      | 1714      | 747              | 967             | 633                             |
|            | Tsunan1, Tsunan2  | 37.04           | 138.60           | 716            | 10.8      | 1899      | 795              | 1104            | 628                             |
|            | Asahi1            | 38.38           | 139.70           | 655            | 9.2       | 1887      | 697              | 1190            | 605                             |
|            | Oshirakawa1       | 37.35           | 139.16           | 627            | 9.1       | 1963      | 668              | 1295            | 563                             |
| Ibaraki    | Ogawa13, Ogawa 14 | 36.93           | 140.59           | 643            | 10.7      | 1474      | 764              | 710             | 785                             |
| Oita       | Kuju_pasture,     | 33.06           | 131.23           | 841            | 10.8      | 2930      | 786              | 2144            | 819                             |
|            | Kuju_forest       |                 |                  |                |           |           |                  |                 |                                 |

Table 1. Environmental properties of Japanese forests and pastureland where investigated soils were collected\*.

\* MAT, MAP, and PET were obtained as averages for 1981–2020 in a global data set of climate and climatic water balance (i.e., TerraClimate) by Abatzoglou et al. (2018). NPP data were obtained as averages for 2001–2020 in the global distribution of NPP estimated from MODIS observation products (i.e., MOD17A3HGF) by Running and Zhao (2021). MAT, mean annual temperature; MAP, mean annual precipitation; PET, potential evapotranspiration; NPP, net primary production.

and Dahlgren, 2016; Wagai et al., 2018). The difference between acid-oxalate- and pyrophosphate-extractable metals (i.e., Alo-p and Feo-p) represented the contents of shortrange-order minerals (Courchesne and Turmel, 2008). Contents of Al and Fe in the solution were measured with an inductively coupled plasma-optical emission spectrometer (ICP-OES) (5110; Agilent Technologies, Santa Clara, CA, USA). Carbon concentrations in pyrophosphate-extracted solution (Cp) were also measured with a total organic carbon (TOC) analyzer (TOC-L; Shimadzu, Kyoto, Japan). For soils from Kuju forest and grassland and two Ogawa forests, we also measured C concentration in free light density fraction (fLF) (Leuthold et al., 2023) obtained by the density fractionation method using sodium polytungstate solution at  $2.0 \,\mathrm{g}\,\mathrm{cm}^{-3}$  (Koarashi et al., 2012), in order to evaluate possible effects of labile C abundance in fLF on CO<sub>2</sub> release increase by DWCs.

For the soils after incubation (see Sect. 2.3), soil microbial biomass C and N were measured by the chloroform fumigation extraction method (Vance et al., 1987). Organic C and total N concentrations in 0.5 M potassium sulfate solution used for extraction of fumigated, and nonfumigated soils were measured using another TOC analyzer (TOC-L; Shimadzu) equipped with a total nitrogen (TN) unit (TNM-L, Shimadzu).

## 2.3 Incubation experiment and soil CO<sub>2</sub> release rate measurement

Soils were incubated aerobically at 20 °C for 84 d including three DWCs (i.e., 28 d per cycle). Simultaneously, soils were incubated in the same manner but without DWCs, during which water content of the soils was maintained at a constant level equivalent to the mean water content for the DWC treatment (Fig. 2). A pre-incubation was conducted at the constant water content for 7 d prior to the 84 d incubation. A post-incubation was also conducted at the constant water content for 28 d after the 84 d incubation to evaluate the remaining effect of DWCs on soil CO<sub>2</sub> release. We con-



**Figure 2.** Schematic time courses of soil moisture during incubation with constant moisture content (a) and DWCs (b) with measurement period of  $CO_2$  release rate.

sider that the incubated soils have been aerobic even after the rewetting to increase the water content by twice the WHC because the  $CO_2$  concentrations in our experiment never surpassed 1%; thus the oxygen concentrations in the incubation jar have likely never decreased below 19% or lower. Also, a sufficiently large volume of our incubation jar (1.0 L) compared to contained soil amounts (i.e., 5.31–10.63 g) and added water contents in the rewetting (i.e., ca. 6 to 7 mL) support the state of aerobic condition during the incubation.

Mason jars (1.0 L volume; Ball, Buffalo, NY, USA) with lids equipped with tube fitting systems for gas sample collection (Koarashi et al., 2012; Nagano et al., 2019) were used as incubation jars. Small vials (300 mL, SM sample glass vial;

| Soil         | Soil type*                       | pH(H <sub>2</sub> O) | EC                  | Water content         | WHC                   | Particle size distribution |      | ize<br>on |
|--------------|----------------------------------|----------------------|---------------------|-----------------------|-----------------------|----------------------------|------|-----------|
|              |                                  |                      | ~ _1                |                       |                       | Sand                       | Silt | Clay      |
|              |                                  |                      | µS cm <sup>-1</sup> | g water per gram soil | g water per gram soil |                            | %    |           |
| Aga1         | Inceptisols or Entisols          | 4.36                 | 23                  | 0.77                  | 0.80                  | 27                         | 30   | 43        |
| Aga2         | Inceptisols or Entisols          | 4.39                 | 27                  | 1.07                  | 1.12                  | 17                         | 59   | 23        |
| Tsunan1      | Inceptisols (brown forest soils) | 4.72                 | 17                  | 0.78                  | 1.06                  | 47                         | 44   | 9         |
| Tsunan2      | Inceptisols (brown forest soils) | 4.08                 | 39                  | 1.06                  | 1.32                  | 34                         | 48   | 18        |
| Asahi1       | Inceptisols (brown forest soils) | 4.07                 | 39                  | 1.70                  | 2.14                  | 29                         | 41   | 30        |
| Oshirakawa1  | Andisols                         | 4.66                 | 24                  | 1.07                  | 1.26                  | 38                         | 45   | 17        |
| Ogawa13      | Inceptisols (brown forest soils) | 5.29                 | 18                  | 0.46                  | 1.00                  | 66                         | 25   | 8         |
| Ogawa14      | Andisols                         | 5.30                 | 27                  | 1.14                  | 1.68                  | 68                         | 23   | 9         |
| Kuju_pasture | Andisols                         | 5.35                 | 133                 | 1.32                  | 1.43                  | 72                         | 22   | 5         |
| Kuju_forest  | Andisols                         | 3.80                 | 168                 | 1.49                  | 1.58                  | 70                         | 25   | 4         |

Table 2. Basic properties of 10 Japanese forests and pastureland soils collected from depths of 0-5 or 0-10 cm.

\* Andisols were determined according to the USDA Soil Taxonomy criteria (USDA Soil Survey Staff, 2022) based on acid-oxalate-extractable Al and 1/2 Fe contents (see text for details). All Andisols were non-allophanic. Other soil types were determined using a Japanese soil digital map, i.e., Japan Soil Inventory NARO (2023). EC: elemental carbon.

Table 3. Soil carbon (C) and nitrogen (N) properties.

| Soil         | Total C<br>% | Total N<br>% | Total C/N | C in free light density fraction (fLF)<br>to total C % |
|--------------|--------------|--------------|-----------|--|
| Agal         | 8.8          | 0.56         | 15.7      | No data  |
| Aga2         | 14.3         | 0.79         | 18.2      | No data  |
| Tsunan1      | 12.4         | 0.63         | 19.8      | No data  |
| Tsunan2      | 15.3         | 0.98         | 15.6      | No data  |
| Asahi1       | 23.7         | 1.41         | 16.9      | No data  |
| Oshirakawa1  | 15.1         | 0.76         | 19.8      | No data  |
| Ogawa13      | 6.8          | 0.31         | 22.2      | 10.0   |
| Ogawa14      | 16.1         | 0.86         | 18.6      | 10.0   |
| Kuju_pasture | 21.8         | 1.37         | 15.9      | 14.0   |
| Kuju_forest  | 22.3         | 1.18         | 18.9      | 5.9  |

Sansyo, Tokyo, Japan) containing 5.31-10.63 g of soil sample depending on water content were placed in mason jars. Under the constant water treatment, we surrounded the small vial with 20 mL of water within the incubation jar to prevent the soil from drying. For each DWC, day 1 to day 7 and day 18 to day 24 were drying stages (Fig. 2), during which the soils were incubated with silica gel  $(20 \text{ g jar}^{-1})$ , which lowered the water content to < 5% WHC by day 7. Day 8 to day 12, day 13 to day 17, and day 25 to day 28 were the driest, wettest, and moderately wet stages, respectively, and CO<sub>2</sub> release rates in these three stages were measured using a gas chromatograph equipped with a thermal conductivity detector (GC-14B; Shimadzu). The CO<sub>2</sub> release rates were also measured in the pre- and post-incubation periods. At the beginning of the wettest stage, soils were rapidly rewetted with distilled water to double the soil water content from the initial status of DWC incubation. For CO2 release rate measurements, at the start of each stage, the headspace of the incubation jar was flushed with CO2-free air for 15 min at a rate of  $0.5 \,\mathrm{L\,min^{-1}}$ , and the jar was closed. At the end of each stage, 15 mL of gas sample was collected from the jar using a 20 mL plastic syringe (Terumo, Kyoto, Japan) and stored in a pre-evacuated 5 mL glass vial (SVG-5; Nichiden Rika, Osaka, Japan). Then, the CO<sub>2</sub> release rate was determined from the increase in CO<sub>2</sub> concentration during this period. After gas sampling, the jars were flushed with CO<sub>2</sub>-free air and closed for the next incubation stage. For 84 d in the incubations with the constant moisture condition as controls, the CO<sub>2</sub> release rates were measured for day 1 to day 12, day 13 to day 28, day 29 to day 40, day 41 to day 56, day 57 to day 68, and day 69 to day 84, in addition to the pre- and postincubation periods. Soil water contents during the incubation were measured periodically and maintained by adding water to ensure the same mean water content between the two treatments. Even in the drying stage under the DWC treatment for day 1 to day 7 and day 18 to day 24, we conducted measurements of soil water content once or twice. The measurements were performed by weighing those soils. Based on these data, we confirmed that the mean soil water content during DWC incubation was equal to that during constant moisture incu-

| Soil         | Acid-oxalate-<br>extractable metals |         | Pyrophosphate-<br>extractable metals |       | Acid oxalate<br>–pyrophosphate |          |                      | Alp/Alo | Pyrophosphate-<br>extractable C |                      |       |
|--------------|-------------------------------------|---------|--------------------------------------|-------|--------------------------------|----------|----------------------|---------|---------------------------------|----------------------|-------|
|              | Al                                  | Fe      | Al +                                 | Al    | Fe                             | Al +     | Al                   | Fe      | Al +                            |                      |       |
|              |                                     |         | 0.5Fe                                |       |                                | 0.5Fe    |                      |         | 0.5Fe                           |                      |       |
|              | mg pe                               | er gram | dry soil                             | mg pe | er gram                        | dry soil | mg per gram dry soil |         |                                 | mg per gram dry soil |       |
| Agal         | 3.3                                 | 7.9     | 7.2                                  | 2.8   | 5.0                            | 5.3      | 0.5                  | 2.8     | 2.0                             | 0.84                 | 21.0  |
| Aga2         | 6.9                                 | 10.6    | 12.2                                 | 6.8   | 10.5                           | 12.0     | 0.1                  | 0.2     | 0.2                             | 0.98                 | 42.9  |
| Tsunan1      | 8.9                                 | 13.8    | 15.8                                 | 8.5   | 11.3                           | 14.2     | 0.4                  | 2.5     | 1.6                             | 0.96                 | 39.9  |
| Tsunan2      | 6.9                                 | 8.5     | 11.1                                 | 7.1   | 7.2                            | 10.6     | -0.2                 | 1.4     | 0.5                             | 1.03                 | 47.4  |
| Asahi1       | 4.8                                 | 10.2    | 9.9                                  | 4.7   | 9.2                            | 9.3      | 0.1                  | 1.0     | 0.6                             | 0.98                 | 45.4  |
| Oshirakawa1  | 9.5                                 | 12.4    | 15.7                                 | 9.1   | 11.0                           | 14.6     | 0.4                  | 1.4     | 1.2                             | 0.95                 | 43.0  |
| Ogawa13      | 9.2                                 | 8.7     | 13.5                                 | 6.9   | 6.5                            | 10.2     | 2.3                  | 2.2     | 3.4                             | 0.75                 | 19.4  |
| Ogawa14      | 24.9                                | 16.6    | 33.2                                 | 23.2  | 12.7                           | 29.6     | 1.6                  | 3.9     | 3.6                             | 0.93                 | 59.1  |
| Kuju_pasture | 22.0                                | 20.3    | 32.2                                 | 19.0  | 14.1                           | 26.0     | 3.1                  | 6.3     | 6.2                             | 0.86                 | 96.7  |
| Kuju_forest  | 19.5                                | 19.1    | 29.0                                 | 19.0  | 16.2                           | 27.1     | 0.4                  | 2.9     | 2.0                             | 0.98                 | 109.8 |

Table 4. Selectively dissolved minerals and associated carbon contents in soils.

bation. All incubations were conducted with three replicates for each treatment and soil.

#### 2.4 Data processing and statistical analysis

The CO<sub>2</sub> release rates were compared between the DWC and constant water content treatments. For the DWC treatment, the CO<sub>2</sub> release rates in the drying stages (i.e., day 1 to day 7 and day 18 to day 24) could not be measured and therefore had to be estimated to evaluate the mean CO<sub>2</sub> release rates for the individual cycle and total of three cycles. The rates in the drying stages were estimated as the mean values of the CO<sub>2</sub> release rates measured before and after the period of interest, in the same manner as described previously (Nagano et al., 2019). Then, the effect size of the DWCs on  $CO_2$  release (defined here as the increase factor,  $IF_{CO_2}$ ) was quantified as the ratio of CO<sub>2</sub> release rate under the DWC condition to that under the constant water content condition (Nagano et al., 2019; Zhang et al., 2020). Here, doubling of  $CO_2$  release by DWC resulted in IF<sub>CO2</sub> of 2, while halving resulted in IF<sub>CO<sub>2</sub></sub> of 0.5.

The pairwise *t* test was applied to examine the statistical significance of differences in CO<sub>2</sub> release rates between the DWC and constant water content treatments with adjustment of site-by-site variations in the metrics. Differences in soil microbial biomass C and N between the two treatments were also evaluated with the pairwise *t* test. To explore predictors explaining the variation in IF<sub>CO2</sub> among soils, relations between IF<sub>CO2</sub> and environmental and soil properties were visualized on scatterplots and evaluated by linear correlation analysis. All statistical analyses were conducted with R 4.1.1 (R Core Team, Vienna, Austria), and *p* < 0.05 was taken to indicate statistical significance.

#### 3 Results

#### 3.1 Quantifying the effect of DWCs on soil CO<sub>2</sub> release

The CO<sub>2</sub> release rates under DWC conditions showed large fluctuations for all soils, along with fluctuations in soil water content (Fig. 3). The CO<sub>2</sub> release rates in the driest stages (3.5%-18.2%) of WHC depending on the soil) were  $3.0-41.5 \mu g C$  per gram dry soil per day. In contrast, CO<sub>2</sub> release rates in the wettest stages (90.8%-201.1%) of WHC) reached  $18.8-194.1 \mu g C$  per gram dry soil per day. In the moderately wet stages, CO<sub>2</sub> release rates were  $8.4-63.9 \mu g C$  per gram dry soil per day. For the constant water content treatment, CO<sub>2</sub> release rates observed for each soil showed little fluctuation during incubation although the rates varied from 7.3 to  $69.9 \mu g C$  per gram dry soil per day depending on the soil.

For all soils, the observed large fluctuations in CO<sub>2</sub> release under DWC treatment resulted in a greater CO<sub>2</sub> release rate than under constant water content treatment, although the  $IF_{CO_2}$  values varied among the soils (Fig. 4). At the wettest stage in the first DWC, the CO<sub>2</sub> release rates increased by 47.2–127.7 µgC per gram dry soil per day compared with those under the constant water content conditions, resulting in  $IF_{CO_2}$  values of 2.9–12.2. For the whole of the first cycle (i.e., the first 28 d), the  $CO_2$  release rates increased by 15.4-43.4 µgC per gram dry soil per day under the DWCs compared with the constant water content conditions, resulting in IF<sub>CO<sub>2</sub></sub> values of 1.6–5.2. For the whole incubation period (84 d) including three DWCs, IF<sub>CO2</sub> values were 1.3-3.7, with an increase in  $CO_2$  release rate by 7.4–23.8 µgC per gram dry soil per day by DWCs. These increases in CO2 release by DWCs were observed in all cycles during the 84 d incubation period, whereas no increase was observed in the 28 d post-incubation period after the three DWCs (Fig. S2 in the Supplement). Considering these results, we focused on



**Figure 3.** Time courses of soil  $CO_2$  release rates (**a**, **b**) and water contents (**c**, **d**) during 84 d incubation under constant water content (**a**, **c**) and three DWC conditions (**b**, **d**).

the  $IF_{CO_2}$  variations obtained for the whole incubation period, including three DWCs.

# 3.2 Exploring predictors of the effect sizes on the increase in CO<sub>2</sub> release

Among the environmental and soil physiochemical properties, reactive mineral and metal contents (especially, Alo + 0.5Feo, Alo, Feo, Alp + 0.5Fep, and Alp) in soils showed significant positive correlations with IF<sub>CO2</sub> (p <0.05; Table 5). In particular, Alp content appeared to be a key predictor of the variation in  $IF_{CO_2}$  among the soils, given that Alp accounted for most (73 %-99 %) of Alo (Table 4) and showed a higher correlation coefficient than others (Table 5). Scatterplots for Alp content and IF<sub>CO2</sub> values are presented in Fig. 5. In addition, the molar ratios of soil total C and Cp contents to Alp contents showed significant negative correlations to  $IF_{CO_2}$  (p < 0.05; Table 5, Fig. 5). Here, it should be noted that  $IF_{CO_2}$  had no significant relationship with soil water content at soil sampling and WHC, suggesting the variation in  $IF_{CO_2}$  among soils resulting from other than such hydrogenic properties of soils. The SOM quality, such as C/N ratios of total bulk soils and K<sub>2</sub>SO<sub>4</sub>-extractable fractions, also had no significant relationship with IF<sub>CO2</sub>. The insignificant relations of SOM quality to IF<sub>CO2</sub> were also supported by the fact that Kuju forest and grassland soils having more than doubled differences in fLF abundance showed only 12% differences in IF<sub>CO2</sub>, and two Ogawa forest soils having almost identical fLF abundances showed more than doubled differences in IF<sub>CO2</sub>. The amounts of clay and sandsized particles showed significant correlations with IF<sub>CO2</sub> after rewetting in the first cycle (p < 0.05, r = -0.66 for clay and 0.71 for sand particles). However, those correlations between the particle contents and IF<sub>CO2</sub> were insignificant for IF<sub>CO2</sub> for a total of three cycles. Thus, in the present study, reactive mineral and metal content, especially Alp content in soils, was the significant predictor for IF<sub>CO2</sub> variation among soils, rather than soil water environments, SOM quality, and particle size distributions.

Among the additional soil properties related to soil microbial activity and abundance, the soil-C-content-specific CO<sub>2</sub> release rate under constant moisture conditions (defined as the CO<sub>2</sub> release rate per unit of C in soil, qCO<sub>2</sub>\_soc, in  $\mu$ g CO<sup>2</sup>-C per milligram of soil carbon per day) showed significant negative correlations with IFCO2 values in all incubation stages (p < 0.01; Table 6, Fig. 6). Here qCO<sub>2</sub>\_soc should be an index for microbial availability of carbon substrate normalized by total C contents in soils. Thus, using qCO<sub>2</sub> soc, we can consider whether the microbially available carbon substrate in interested soil should be much more than in other soils. Microbial biomass N also showed significantly negative correlations with  $IF_{CO_2}$  values after rewetting in the first cycle and in the whole of the first cycle (p < 0.05) but not in the whole incubation period including three cycles (p = 0.12).

There were also considerable relations among Alp contents, total C/Alp molar ratio, and qCO<sub>2</sub>\_soc (Fig. 7). Soil Alp content and total C/Alp molar ratio showed a negative



**Figure 4.** Comparisons of mean CO<sub>2</sub> release rates after rewetting in the first cycle, the whole of the first cycle, and for the total of three cycles between DWC and constant water content conditions (**a**–**c**), and the factor of increase in CO<sub>2</sub> release by DWCs (IF<sub>CO2</sub>) for individual periods (**d**–**f**). Statistically significant differences (p < 0.05, pairwise *t* test) in CO<sub>2</sub> release rate between the two treatments are presented.

| Table 5. Pearson' | 's correlation | coefficients | between I | F <sub>CO</sub> , and | d environmental | and soil | properties |
|-------------------|----------------|--------------|-----------|-----------------------|-----------------|----------|------------|
|-------------------|----------------|--------------|-----------|-----------------------|-----------------|----------|------------|

| Environmental or soil property | After rewetting | Total          | Total           |
|--------------------------------|-----------------|----------------|-----------------|
|                                | in first cycle  | in first cycle | in three cycles |
| Elevation                      | 0.36            | 0.37           | 0.22            |
| MAT                            | 0.54            | 0.55           | 0.46            |
| MAP                            | 0.05            | 0.08           | -0.11           |
| MAP – PET                      | 0.01            | 0.04           | -0.14           |
| Water content at sampling      | -0.11           | -0.07          | -0.15           |
| WHC                            | 0.06            | 0.08           | 0.04            |
| pH(H <sub>2</sub> O)           | 0.54            | 0.51           | 0.57            |
| Electronic conductivity        | 0.19            | 0.21           | -0.01           |
| Total C                        | -0.03           | 0.01           | -0.11           |
| Total N                        | -0.07           | -0.03          | -0.12           |
| C/N ratio                      | 0.27            | 0.23           | 0.22            |
| Sand                           | 0.72*           | 0.70*          | 0.56            |
| Silt                           | -0.50           | -0.48          | -0.35           |
| Clay                           | -0.69*          | -0.68*         | -0.57           |
| Alo + 0.5Feo                   | 0.81**          | 0.82**         | 0.70*           |
| Alo                            | 0.84**          | 0.85**         | 0.73*           |
| Feo                            | 0.64*           | 0.66*          | 0.53            |
| Alp+0.5Fep                     | 0.81**          | 0.82**         | 0.70*           |
| Alp                            | 0.84**          | 0.85**         | 0.74*           |
| Fep                            | 0.55            | 0.58           | 0.54            |
| Alo-p + 0.5Feo-p               | 0.46            | 0.43           | 0.37            |
| Alo-p                          | 0.56            | 0.57           | 0.48            |
| Feo-p                          | 0.55            | 0.55           | 0.46            |
| Total C/Alp molar ratio        | -0.79**         | -0.78**        | -0.72*          |
| Ср                             | 0.39            | 0.42           | 0.23            |
| Cp/Alp molar ratio             | -0.79**         | -0.75*         | -0.74*          |



**Figure 5.** Relations between IF<sub>CO2</sub> and soil pyrophosphate-extractable Al (Alp) content (**a–c**), total C/Alp molar ratio (**d–f**), and pyrophosphate-extractable C (Cp)/Alp molar ratio (**g–i**). Significant correlation coefficients at p < 0.01 and p < 0.05 are indicated with single (\*) and double asterisks (\*\*), respectively.

Table 6. Pearson's correlation coefficients between  $IF_{CO_2}$  and soil microbial properties.

| Soil microbial property  | After rewetting<br>in first cycle | Total in first cycle | Total in three cycles |
|--|-----------------------------------|----------------------|-----------------------|
| Total-C-content-specific CO <sub>2</sub> release rate under no water fluctuation, qCO <sub>2_soc</sub> | -0.87*                            | -0.86**              | -0.77*                |
| K <sub>2</sub> SO <sub>4</sub> -extractable C  | -0.25                             | -0.21                | -0.33                 |
| K <sub>2</sub> SO <sub>4</sub> -extractable N  | -0.63                             | -0.60                | -0.55                 |
| K <sub>2</sub> SO <sub>4</sub> -extractable C/N  | 0.15                              | 0.18                 | -0.01                 |
| Microbial biomass C  | -0.62                             | -0.61                | -0.48                 |
| Microbial biomass N  | -0.67*                            | -0.66*               | -0.52                 |
| Microbial biomass C/N  | 0.47                              | 0.48                 | -0.31                 |

correlation at p < 0.05. The qCO<sub>2</sub>\_soc showed a positive correlation with total C/Alp molar ratio and negative correlation with Alp content at p < 0.01. The Cp/Alp molar ratio also showed similar covariations with these variables, showing a strongly positive correlation with total C/Alp molar ratio (r = 0.90, p < 0.01; Fig. S3 in the Supplement).

It should be noted that both microbial biomass C and N were significantly lower in soils incubated under DWC conditions than constant water content conditions (p < 0.05; Fig. 8). Microbial biomass C under DWC conditions was lower by  $\geq 20.2 \%$  than that under constant water content conditions. Microbial biomass N under DWC conditions was  $\geq 12.6 \%$  lower than under constant water content conditions. Nevertheless, there were no significant correlations be-

tween the differences in microbial biomass and the value of  $IF_{CO_2}$  ( $p \ge 0.15$ ; Fig. 9).

## 4 Discussion

An increase in  $CO_2$  release due to DWCs was consistently observed across 10 forest and pastureland soils in Japan (Figs. 3 and 4). The comprehensive increases in  $CO_2$  release by DWCs were different from the findings of a recent metaanalysis of studies showing no significant increases in  $CO_2$ release under DWCs compared with constant water content with an equivalent mean water content during the period of interest (Zhang et al., 2020). Furthermore, our observations quantified the increase in  $CO_2$  release due to three DWCs as



**Figure 6.** Relations between IF<sub>CO2</sub> and total-C-content-specific CO<sub>2</sub> release rate under constant moisture content (qCO<sub>2</sub>\_soc). Significant correlation coefficients at p < 0.01 and p < 0.05 are indicated with single (\*) and double asterisks (\*\*), respectively.



**Figure 7.** Relations among soil Alp contents, total C/Alp molar ratio, and qCO<sub>2</sub>\_soc. Significant correlation coefficients at p < 0.01 and p < 0.05 are indicated with single (\*) and double asterisks (\*\*), respectively.

 $IF_{CO_2}$  values of 1.3–3.7. The observed effect size of DWCs on the CO<sub>2</sub> release from soils was large given that even a 20 % increase in CO<sub>2</sub> release from the world's soils can exceed the annual CO<sub>2</sub> emission from anthropogenic processes (Friedlingstein et al., 2020).

Furthermore, analyzing the relations between  $IF_{CO_2}$  and fundamental soil properties showed a significantly positive correlation between IF<sub>CO2</sub> and soil Alp content (Table 5, Fig. 5). Of the organo-metal complexes measured as pyrophosphate-extractable metals, Fep is known to be sensitive to DWCs, especially in seasonally flooded forests and wetlands, likely due to their vulnerability to redox potential changes caused by alterations to the water regime (Lacroix et al., 2019; Chen et al., 2017, 2018; Chen and Thompson, 2018). Less is known about the vulnerability of Alp to DWCs. However, soil Alp content may be affected by DWCs through changes in soil acidity. A previous field survey conducted in Japanese forest and arable soils by Takahashi et al. (2006) showed that liming of non-allophanic Andisols increased soil pH and decreased Alp content. Miyazawa et al. (2013) verified this behavior by a laboratory incubation experiment for Andisols. Although we did not monitor pH during incubation in the present study, increases in pH after DWCs have been widely observed in upland agricultural (Wang et al., 2020; Meng et al., 2020), seasonally submerged paddy, and wetland soils (Majumdar et al., 2023). Therefore, the acidity mitigation by DWCs would destroy the organo-Al complexes and increase microbially available C through the release of C protected by the organo-Al complexes or other soil elements, such as macro- and microaggregates, which are tightly bonded by organo-Al complexes and thus physically protect organic C from microbial decomposition (Asano and Wagai, 2014; Takahashi and Dahlgren, 2016; Wagai et al., 2018).

Covariations among soil Alp contents, total C/Alp molar ratio, and qCO<sub>2</sub>\_soc (Fig. 7) also support the state of soil Alp content as the primary predictor of variations in IFCO2. Negative correlations of soil Alp contents with both total C/Alp molar ratio and qCO<sub>2</sub>\_soc suggested that more Alp to total C (i.e., low total C/Alp values) strengthens binding between Alp and organic matter, resulting in resistance of organic matter to microbial decomposition under constant water conditions (i.e., low qCO2\_soc values). These features among reactive Al and SOM dynamics would result in negative correlations of qCO<sub>2</sub>\_soc and total C/Alp molar ratio with  $IF_{CO_2}$ (Table 6, Figs. 5 and 6). However, such an ability of Alp-rich soils to protect SOM from microbial decomposition likely does not persist under conditions of increased water fluctuations associated with DWCs, as suggested above. Given that amounts of pyrophosphate-extractable C (Cp) in investigated soils (>  $19000 \,\mu g$  per gram dry soil; Table 4), which represented C associated with the organo-metal complex, were substantially greater than observed CO<sub>2</sub> release increase by DWCs (620–1999 µgC per gram dry soil per 84 d), limited and specific C substrates in the organo-metal complex were likely associated with CO<sub>2</sub> increase by DWCs.



**Figure 8.** Comparisons of microbial biomass C (a) and N (b) after incubation between DWC and constant water content conditions. Significant differences (p < 0.05 by pairwise *t* test) in microbial biomass between two treatments are presented.



**Figure 9.** Relations between  $IF_{CO_2}$  and microbial biomass C or N decrease by DWCs. There were no statistically significant correlations (p > 0.05).

We also found substantially lower microbial biomass in soils subjected to DWCs than constant water content conditions (Fig. 8), suggesting a decrease in microbial biomass through the destruction of microbial cells by DWCs (Kaiser et al., 2015; Marumoto et al., 1977, 1982; Marumoto, 1984; Nagano et al., 2023; Unger et al., 2010, 2012). The destruction of microbial cells is expected to release soluble organic matter available for microbes that have survived the DWC and to cause a marked increase in  $CO_2$  release after rewetting (Marumoto et al., 1977, 1982; Marumoto, 1984; Nagano et al., 2019; Unger et al., 2010, 2012). Nevertheless, the contribution of microbially derived substances to the increase in  $CO_2$  release remained unclear in the present study because of the lack of a significant correlation between  $IF_{CO_2}$  and the decrease in microbial biomass (Fig. 9). Whereas there was also no significant correlation between the decrease in

microbial biomass and the increase in  $CO_2$  release due to DWCs (Fig. S4 in the Supplement), the decrease in microbial biomass C (246–1134 µg C per gram dry soil) was within the amount of increase in  $CO_2$  release (620–1999 µg C per gram dry soil per 84 d). Therefore, the strict mechanisms of these carbon sources to  $CO_2$  release increase, including the persistence and timing of their contribution, still require further work (Schimel, 2018; Barnard et al., 2020), considering the significant contribution of more than two carbon pools to the  $CO_2$  release increase (Slessarev and Schimel, 2020; Warren and Manzoni, 2023), as well as carbon likely released by destructions of organo-metal complexes and microbial biomass by DWC, as suggested in the present study.

It should be noted that shortcomings from unmeasured CO<sub>2</sub> release during the drying periods for DWCs treatments (i.e., day 1 to day 7 and day 18 to day 24 in each cycle) should be minor, while the linear changes in CO<sub>2</sub> release rate during the drying also assumed in other studies (e.g., Nagano et al., 2019; Zhang et al., 2020; Jin et al., 2023). This is because we observed significant relationships between  $IF_{CO_2}$  and the organo-Al complex, also in  $IF_{CO_2}$ , after the rewetting in addition to the  $IF_{CO_2}$  for the first cycle and three cycles in total (Table 5).

## 5 Conclusions

Through the present study, a comprehensive increase in  $CO_2$  release by DWCs (i.e., 1.3–3.7-fold greater than  $CO_2$  release under constant water conditions) was observed in Japanese forest and pastureland soils. These magnitudes of increase in  $CO_2$  release were strongly correlated with soil Alp content, total C/Alp molar ratio, and total-C-content-specific  $CO_2$  release rate under constant water conditions (i.e.,  $qCO_2$ \_soc), suggesting the possible vulnerability of SOM protection by organo-Al complexes against DWCs. A decrease in microbial biomass by DWCs was also suggested, whereas their relations with the increase in  $CO_2$  release remain to be determined in future studies.

**Data availability.** The data that support the findings of this study are available from the corresponding author upon reasonable request.

**Supplement.** The supplement related to this article is available online at: https://doi.org/10.5194/soil-11-35-2025-supplement.

**Author contributions.** YS and HN established the basic research design; conducted all data analyses, including software preparation, validation, and visualization; and wrote the original manuscript. SH, MAA, JK, and HN conducted soil sampling. YS, SH, MAA, JK, and HN contributed to the detailed research design, soil analysis, data validation, interpretation of the results, and editing of the

manuscript. TY and YK provided essential support in analysis of soil properties and data. All authors contributed to editing the article and approved the submitted version.

**Competing interests.** The contact author has declared that none of the authors has any competing interests.

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Acknowledgements. The authors thank Kouki Hikosaka and Hirofumi Kajino of Tohoku University for providing the vegetation information on the Oita forest site. Ayako Tamaki and Masahiro Otaki at Niigata University supported sample preparation and analysis during the experiment and text editing in preparing the manuscript. Rei Shibata at Niigata University and Masami Tsukahara, Kosuke Ito, and Tatsuki Tanaka at Niigata Prefectural Forest Research Institute helped in site selection and soil sampling in Niigata Prefecture, Japan. Naoki Harada, Kazuki Suzuki, and Asiloglu Rasit of Niigata University and Hiroyuki Sase and Rieko Urakawa of the Asia Center for Air Pollution Research (ACAP) provided valuable preliminary discussion in writing the manuscript. The authors also thank Misuzu Kaminaga, Kikuko Yoshigaki, Makiko Ishihara, and Kazumi Matsumura at JAEA for support with laboratory work.

**Financial support.** This research has been supported by the Japan Society for the Promotion of Science (JSPS) KAKENHI (grant nos. 21H02231, 21H05313, and 22H05717).

**Review statement.** This paper was edited by Axel Don and reviewed by Haicheng Zhang and one anonymous referee.

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