SOIL, 1, 459–473, 2015 www.soil-journal.net/1/459/2015/ doi:10.5194/soil-1-459-2015 © Author(s) 2015. CC Attribution 3.0 License.





# Evaluation of vineyard growth under four irrigation regimes using vegetation and soil on-the-go sensors

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Received: 15 October 2014 – Published in SOIL Discuss.: 25 November 2014 Revised: 18 May 2015 – Accepted: 3 June 2015 – Published: 17 June 2015

**Abstract.** Precision agriculture is a useful tool to assess plant growth and development in vineyards. The present study focused on spatial and temporal analysis of vegetation growth variability, in four irrigation treatments with four replicates. The research was carried out in a vineyard located in the southwest of Spain during the 2012 and 2013 growing seasons. Two multispectral sensors mounted on an all-terrain vehicle (ATV) were used in the different growing seasons/stages in order to calculate the vineyard normalized difference vegetation index (NDVI). Soil apparent electrical conductivity (ECa) was also measured up to 0.8 m soil depth using an on-the-go geophysical sensor. All measured data were analysed by means of principal component analysis (PCA). The spatial and temporal NDVI and ECa variations showed relevant differences between irrigation treatments and climatological conditions.

# 1 Introduction

Terroir is a French concept that means that "there are unique aspects of a place that shape the quality of grapes and wine". Those aspects that impact on grapes and wine quality are usually associated with topography, soil, climate, plant management and plant genetics (Vaudour, 2002). According to several authors, the study of plant vegetative vigour is an essential parameter to successfully manage yield and grape/wine quality because plant growth integrates climate, soil, topography, available water and other plant controlling factors (Carbonneau, 1995; Cortell et al., 2005; Deloire et al., 2005; Smart, 1985). Consequently, appropriate management of soil and consideration of the main climatic variables are key factors to obtain good yields and, ultimately, quality wines. Vineyard canopy management practices such as pruning systems, shoot orientation, shoot thinning or leaf removal, have the capacity to modify climate factors around the plant and, consequently, to modify grape and wine quality (Dry, 2000).

Vineyard behaviours with regard to water management have been studied in recent decades in a wide range of environments and vineyard varieties because of the implications of irrigation on yield and quality of the final product (Smart and Coombe, 1983; Williams and Araujo, 2002; Mullins et al., 1992; Bravdo and Hepner, 1986; Intrigliolo and Castel, 2010). Previous authors also indicate that vine vegetative development is highly influenced by water availability, to the extent that this may become a limiting factor. However, with the same irrigation depth, sometimes the response between two close plants is not the same. This point should be considered when selecting methods to estimate crop water status in order to achieve better management and to meet the production objectives defined at the beginning of the growing season. On the one hand, covering all water needs is not recommended because this creates management problems, reduces crop quality and in general increases unnecessarily the cost of cultivation. On the other hand, increasing water availability to the vineyard causes not only grape production to rise, but also the costs of pruning and plant protection treatments, and usually results in reduction of grape quality. Thus, water stress has to be controlled to achieve good yield and quality of grapes as well as balanced growth while avoiding the problems of excess water. It is essential, therefore, to know the correct way to manage this crop.

Several studies related to spectral vegetation indices (VIs) have carried out analyses of vine canopy, shape, size and functional capacity in order to determine spatial and temporal management of vegetation as well as other productions factors such as water. Spectral VIs are able to predict a large number of plant features, such as leaf area index (LAI), vegetation fraction cover, fraction of absorbed photosynthetically active radiation (fAPAR), chlorophyll pigment concentration, plant stress and other related parameters (Baret and Guyot, 1991; Gitelson and Merzlyak, 2004; Jordan, 1969; Peñuelas et al., 1993; Rondeaux et al., 1996). These VIs, which are mathematical combinations of two or more plant reflectances at specific wavelengths, can be used in vine growth site-specific management enabling the optimization of grape quality and yield (Lamb and Bramley, 2001).

Nowadays, it is possible to obtain a plant spectral signature with a multispectral proximal sensor (Tardáguila and Diago, 2008) suitable for studying vine vegetation terroir. The normalized difference vegetation index (NDVI), developed by Rouse et al. (1973), is one of the most extensive indices used for vegetation growth analysis. It can be calculated as

$$NDVI = \frac{NIR - Red}{NIR + Red},$$
(1)

where Red and near infra-red (NIR) are the reflectance parameters in the red and NIR electromagnetic radiation bands, respectively. When electromagnetic radiation (natural or man-made) impacts on living green leaves, part of it is absorbed, another part is transmitted and the rest is reflected. The electromagnetic radiation spectral range that can be absorbed by plants is the photosynthetically active radiation (PAR), and is between 0.4 and 0.7  $\mu$ m (similar to visible range). Within this range, chlorophyll is efficient at capturing the red and blue ranges, and it normally reflects the green, the infrared (IR) and the NIR ranges. Thus, on the basis of NDVI, the greater the amount of vegetative cover or canopy, the higher the value in the index. However, the ability to absorb and reflect both bands depends not only on health plant status but also on plant size. Thus, a plant with water stress, or any other kind of stress (pests, disease, nutritional deficiencies, etc.), will have less capacity to absorb the red band through photosynthetic apparatus and to reflect the NIR band on the cell walls, and will consequently have a lower NDVI value. Therefore, expression of vineyard vegetative development can be related to NDVI. Several studies have shown the relationship between vegetative canopy parameters, such as LAI and fAPAR, and physiological factors, such as crop production and grape quality, in harvest or plant water status (Smart and Coombe, 1983; Jackson et al., 1983; Dry, 2000). Furthermore, NDVI is also largely related to density of the vegetative canopy of the vineyard (Dobrowski et al., 2002; Johnson, 2003; Hall et al., 2008), so that any change in the factors affecting growth and vineyard development could be estimated by NDVI.

Terroir is also affected by physical, chemical and biological soil properties; as a tool to interpret these soil property variations, soil apparent electrical conductivity (ECa) may be used. Soil ECa measurements can characterize soil spatial variability, with regard mainly to the physical features of the soil, and have been used by other authors to delineate homogeneous management zones (Terrón et al., 2013; Corwin and Lesch, 2003; Moral et al., 2010). Soil ECa measurements can be obtained through geoelectric sensors and this can be an easy and economical way of sampling the soil and guiding soil evaluators in their soil property analyses (Terrón et al., 2011).

According to Hall et al. (2002) the implementation of vineyard site-specific tools are needed in order to better manage vineyards. Thus, the present work makes use of precision agriculture tools to determine (i) the effects of different irrigation treatments on vine vegetation growth in two different climatic seasons and (ii) influence of the soil on vegetation growth expression.

# 2 Material and methods

#### 2.1 Study area and experimental design

The study was carried out during the 2012 and 2013 growing seasons, in a field belonging to the Agrarian Research Institute "La Orden – Valdesequera", in Extremadura (Spain)  $(38^{\circ}51' \text{ N}, 6^{\circ}40' \text{ E})$ . The climate is characterized by mild winters and hot summers, with maximum temperatures reaching 40 °C. Rainfall is irregular, with dry summers and often with an annual average below 500 mm.

The study area is located in a vineyard of 1.8 ha, varietal Tempranillo (*Vitis vinifera* L.) grafted on Richter 110. It was planted in 2001 by vertical trellis in bilateral cordon system, with 0.6 m stem height and 12 buds per plant. Cultivar Tempranillo is a vigorous variety adaptable to all types of soils, preferably slightly acid, and oriented towards areas with a high number of hours of sun per year.

The field is situated in the Guadiana River valley, whose soil morphology is typical of the Quaternary, carved in Tertiary sediments. Surface horizons have been artificially transformed for agricultural use with water management, which has resulted in destruction of these horizons and in burial of diagnostic horizons, even below 50 cm. These latter horizons are not differentiated and they have evolved edaphically from the sediments and materials of the lower terraces of the Guadiana River, which cover the coarser sediments. According to Soil Survey Staff (2006), the soil is in the order Entisol, suborder Orthent and the great group Xerorthent (Xeric).

There exists a soil study consisting of a 15-point database randomly distributed on the study area. Tables 1a and 1b show statistic values of soil samples of some physicochemical parameters at two depths (from 0 to 0.30 m and from 0.30 to 0.60 m respectively) which were analysed by official laboratory procedures.

From our measurements of a nearby well, we verified that there exists a water table placed at a depth of 6 m, in accordance with the river water level. However, from our experience in this area, we know that the level can vary due to rainfall and water use for irrigation.

The experimental design was randomized complete blocks, with four replicates (plots) per treatment. Each plot had 108 vines in 6 rows with 18 vines per row, where the distance between plants and rows were 1.20 and 2.50 m respectively, placed on a trellis with an east–west row direction. Water treatments were dependent on the growing season (Fig. 1):

- i. 2012 treatments were divided into four levels of irrigation, corresponding to four levels of crop evapotranspiration (ETc) rates:
  - a. fully watered, based on the application of 100% ETc;
  - regulated deficit irrigation (RDI) 50-20, based on the regulated deficit irrigation technique, with 50 % ETc before *véraison* and 20 % ETc after it;
  - c. RDI 50-0, based on the regulated deficit irrigation technique, with 50% ETc before *véraison* and 0% ETc after it;
  - d. non-watered, based on rainfed treatment; and
- ii. 2013 treatments were reduced to three levels of irrigation, corresponding to three levels of ETc rates:
  - a. fully watered, based on the application of 100% ETc;
  - b. RDI 30, based on the regulated deficit irrigation technic, with 30 % ETc throughout the season; and
  - c. non-watered, based on rainfed treatment.

The irrigation system was set up by drip irrigation with one emitter of  $4 L h^{-1}$  every 0.60 m (two emitters per vine) attached to a wire suspended 0.40 m above the ground. Full ETc was calculated by means of the weight differences recorded on a weighing lysimeter installed in the centre of the



**Figure 1.** Maps of treatments and respective plots: (**a**) map of treatments of 2012 growing season; (**b**) map of treatment of 2013 growing season, where "a" and "b" replicates of RDI 30 are in the same emplacement of the respective replicates of RDI 50-20 and RDI 50-0 of the previous season.

assay, corresponding to a fully watered treatment plot (Yrissarry and Naveso, 1999). Two grapevine plants were planted in the lysimeter container in order to provide the water balance as their canopy developed. Precipitation was collected by an agro-meteorological station located in a reference field near the vineyard.

Soil management was characterized by two annual cultivator treatments: one in winter dormancy and another at the bud break phenological stage. Later, spontaneous vegetation was controlled by herbicide treatments and  $250-350 \text{ kg ha}^{-1}$  of NPK fertilization (9-18-27) was added to the soil. With regard to canopy management, a spring pruning was performed to adjust the potential yield to the 12 initial shoots. Subsequently, before the *véraison* stage, growing shoots were introduced into the trellis to facilitate the passage of agricultural machinery. From *véraison* to harvest, plant protect treatments were made against cryptogamic diseases in cycles of 15 to 20 days.

# 2.2 Vegetation index and soil apparent electrical conductivity

The NDVI estimation was performed with two active proximal multi-spectral sensors mounted on an all-terrain vehicle (ATV). These sensors (OptRx ACS-430, Ag Leader Technology, USA) report directly the vineyard canopy NDVI calculated with red (0.67  $\mu$ m) and NIR (0.78  $\mu$ m) wavelengths. Data sets were collected using a personal digital assistant (PDA) data logger connected to the sensors with TopView software (Betop Topografía SL, Seville – Spain). Geographical coordinates were obtained by a dual-frequency global positioning system (GPS) (GGD Maxor JAVAD Javad GNSS Inc., USA) with real-time kinematic (RTK) differential corrections that reached a planimetric accuracy lower

<b>Table 1. (a)</b> Soil composition from 0 to 0.30 m of the vineyard assa	y. (b) Soil composition from 0.3	30 to 0.60 m of the vineyard assay
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(a) Sample	Clay	Sand	Silt	Texture	pH (1:2.5)	Organic matter	Total nitrogen	Assimilable phosphorus	Potassium $(K^+)$	CEC
	%	%	%			%	%	ppm	$meq (100 g)^{-1}$	$meq(100g)^{-1}$
1	21.4	51.14	27.46	Silty clay loam	6.91	1.35	0.038	22.93	0.164	8.82
2	17.76	55.03	27.21	Sandy loam	7.23	1.27	0.024	0.3	0.113	7.64
3	17.33	58.13	24.54	Sandy loam	7.38	0.83	0.02	0.3	0.109	7.68
4	21.35	43.6	35.05	Loam	7.27	0.94	0.022	2.42	0.168	7.57
5	22.13	41.67	36.2	Loam	6.74	0.88	0.013	15.09	0.171	8.24
6	21.93	40.63	37.44	Loam	7.13	0	0.014	7.26	0.159	9.45
7	25.87	39.02	35.11	Loam	7.31	0.08	0.019	21.22	0.235	10.54
8	20.45	47.05	32.5	Loam	7.37	0.16	0.022	0.3	0.205	10.2
9	17.25	41.65	41.1	Loam	6.87	1.2	0.03	24.04	0.201	10.91
10	19.35	48.98	31.67	Loam	6.82	1.3	0.03	35.23	0.259	11.07
11	20.22	51.02	28.76	Loam	6.72	1.12	0.022	74.13	0.268	10.87
12	19.47	52.49	28.04	Sandy loam	6.81	0.8	0.026	32.46	0.199	10.44
13	18.85	49.13	32.02	Loam	7.08	0.9	0.019	22.58	0.197	10.85
14	19.45	47.78	32.77	Loam	7.23	0.88	0.027	12.31	0.176	10.64
15	20.14	50.32	29.54	Loam	7.23	1	0.029	20.34	0.197	10.14
Mean	20.20	47.84	31.96		7.07	0.85	0.02	19.39	0.19	9.67
Median	20.14	48.98	32.02		7.13	0.90	0.02	20.34	0.20	10.20
SD	2.21	5.57	4.48		0.24	0.44	0.01	19.02	0.05	1.32
Skewness	3.03	-0.07	-1.98		-0.14	-0.91	0.41	1.62	0.08	-0.67
Kurtosis	11.38	2.20	7.62		1.53	2.78	3.02	6.03	2.77	1.85
(b) Sample	Clay	Sand	Silt	Texture	pH (1:2.5)	Organic matter	Total nitrogen	Assimilable phosphorus	Potassium (K <sup>+</sup> )	CEC
(b) Sample	Clay %	Sand %	Silt %	Texture	pH (1:2.5)	Organic matter %	Total nitrogen %	Assimilable phosphorus ppm	Potassium (K <sup>+</sup> ) meq $(100 \text{ g})^{-1}$	$\begin{array}{c} \text{CEC} \\ \text{meq} \left( 100  \text{g} \right)^{-1} \end{array}$
( <b>b</b> ) Sample	Clay % 16.86	Sand % 55.55	Silt % 27.48	Texture Sandy loam	рН (1:2.5) 7.39	Organic matter % 0.1	Total nitrogen % 0.003	Assimilable phosphorus ppm 0.2	Potassium $(K^+)$ meq $(100 g)^{-1}$ 0.105	$\frac{\text{CEC}}{\text{meq}(100\text{g})^{-1}}$ 8.51
( <b>b</b> ) Sample	Clay % 16.86 17.32	Sand % 55.55 46.08	Silt % 27.48 36.6	Texture Sandy loam Loam	pH (1:2.5) 7.39 6.98	Organic matter % 0.1 0.39	Total nitrogen % 0.003 0.007	Assimilable phosphorus ppm 0.2 0.2	Potassium $(K^+)$ meq $(100 g)^{-1}$ 0.105 0.115	$\begin{array}{c} \text{CEC} \\ \text{meq}  (100  \text{g})^{-1} \\ \\ 8.51 \\ 9.52 \end{array}$
(b) Sample	Clay % 16.86 17.32 14.17	Sand % 55.55 46.08 59.75	Silt % 27.48 36.6 26.08	Texture Sandy loam Loam Sandy loam	pH (1:2.5) 7.39 6.98 7.49	Organic matter % 0.1 0.39 0.1	Total nitrogen % 0.003 0.007 0.005	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2	Potassium $(K^+)$ meq $(100 g)^{-1}$ 0.105 0.115 0.099	CEC meq (100 g) <sup>-1</sup> 8.51 9.52 8.23
(b) Sample	Clay % 16.86 17.32 14.17 21.93	Sand % 55.55 46.08 59.75 44.7	Silt % 27.48 36.6 26.08 33.37	Texture Sandy loam Loam Sandy loam Loam	pH (1:2.5) 7.39 6.98 7.49 6.77	Organic matter % 0.1 0.39 0.1 0.45	Total nitrogen % 0.003 0.007 0.005 0.018	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} \text{Potassium } (\text{K}^+) \\ \text{meq} (100 \text{ g})^{-1} \\ 0.105 \\ 0.115 \\ 0.099 \\ 0.132 \end{array}$	CEC meq(100g) <sup>-1</sup> 8.51 9.52 8.23 10.92
(b) Sample	Clay % 16.86 17.32 14.17 21.93 22.92	Sand % 55.55 46.08 59.75 44.7 37.67	Silt % 27.48 36.6 26.08 33.37 39.41	Texture Sandy loam Loam Sandy loam Loam	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46	Organic matter % 0.1 0.3 0.1 0.45 0.31	Total nitrogen % 0.003 0.007 0.005 0.018 0.014	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} Potassium (K^+) \\ meq  (100  g)^{-1} \\ 0.105 \\ 0.115 \\ 0.099 \\ 0.132 \\ 0.137 \end{array}$	CEC meq (100 g) <sup>-1</sup> 8.51 9.52 8.23 10.92 11.81
(b) Sample	Clay % 16.86 17.32 14.17 21.93 22.92 19.86	Sand % 55.55 46.08 59.75 44.7 37.67 43.28	Silt % 27.48 36.6 26.08 33.37 39.41 36.86	Texture Sandy loam Loam Loam Loam Loam	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.86	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} Potassium (K^+)\\ meq (100g)^{-1}\\ 0.105\\ 0.115\\ 0.099\\ 0.132\\ 0.137\\ 0.117\end{array}$	CEC meq (100 g) <sup>-1</sup> 8.51 9.52 8.23 10.92 11.81 11.58
(b) Sample 1 2 3 4 5 6 7	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67	Texture Sandy loam Loam Loam Loam Loam Loam	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.86 6.83	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} Potassium (K^+) \\ meq  (100  g)^{-1} \\ 0.105 \\ 0.115 \\ 0.099 \\ 0.132 \\ 0.137 \\ 0.117 \\ 0.145 \end{array}$	CEC meq(100g) <sup>-1</sup> 8.51 9.52 8.23 10.92 11.81 11.58 11.66
(b) Sample	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.1	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67 42.13	Texture Sandy loam Loam Sandy loam Loam Loam Loam Loam Loam	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.86 6.83 7.19	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} Potassium (K^+)\\ meq (100g)^{-1}\\ 0.105\\ 0.115\\ 0.099\\ 0.132\\ 0.137\\ 0.117\\ 0.145\\ 0.151\end{array}$	CEC meq (100 g) <sup>-1</sup> 8.51 9.52 8.23 10.92 11.81 11.58 11.66 11.45
(b) Sample 1 2 3 4 5 6 7 8 9	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.1 25.4	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77 33.27	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67 42.13 41.33	Texture Sandy loam Loam Loam Loam Loam Loam Loam Loam	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.86 6.83 7.19 6.72	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39 0.38	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009 0.009	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} Potassium (K^+)\\ meq (100  g)^{-1}\\ 0.105\\ 0.115\\ 0.099\\ 0.132\\ 0.137\\ 0.117\\ 0.145\\ 0.151\\ 0.136\\ \end{array}$	CEC meq(100g) <sup>-1</sup> 8.51 9.52 8.23 10.92 11.81 11.58 11.66 11.45 11.81
(b) Sample 1 2 3 4 5 6 7 8 9 10	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.1 25.4 24.55	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77 33.27 39.19	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67 42.13 41.33 36.26	Texture Sandy loam Loam Loam Loam Loam Loam Loam Loam L	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.86 6.83 7.19 6.72 6.63	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39 0.38 0.64	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009 0.009 0.008 0.016	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} Potassium (K^+)\\ meq (100g)^{-1}\\ 0.105\\ 0.115\\ 0.099\\ 0.132\\ 0.137\\ 0.117\\ 0.145\\ 0.151\\ 0.136\\ 0.153\\ \end{array}$	$\begin{array}{c} \text{CEC} \\ \text{meq} \left( 100  \text{g} \right)^{-1} \\ 8.51 \\ 9.52 \\ 8.23 \\ 10.92 \\ 11.81 \\ 11.58 \\ 11.66 \\ 11.45 \\ 11.81 \\ 28.19 \end{array}$
(b) Sample 1 2 3 4 5 6 7 8 9 10 11	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.61 25.4 24.55 23.25	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77 33.27 39.19 41.11	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67 42.13 41.33 36.26 35.64	Texture Sandy loam Loam Loam Loam Loam Loam Loam Loam L	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.86 6.83 7.19 6.72 6.63 6.67	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39 0.38 0.64 0.65	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009 0.009 0.009 0.008 0.016 0.011	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} Potassium (K^+)\\ meq (100g)^{-1}\\ 0.105\\ 0.115\\ 0.099\\ 0.132\\ 0.137\\ 0.117\\ 0.145\\ 0.151\\ 0.156\\ 0.153\\ 0.149\\ \end{array}$	$\begin{array}{c} \text{CEC} \\ \text{meq} \left( 100  \text{g} \right)^{-1} \\ 8.51 \\ 9.52 \\ 8.23 \\ 10.92 \\ 11.81 \\ 11.58 \\ 11.66 \\ 11.45 \\ 11.81 \\ 28.19 \\ 19.53 \end{array}$
(b) Sample 1 2 3 4 5 6 7 8 9 10 11 12	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.1 25.4 24.55 23.25 19.44	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77 33.27 39.19 41.11 48.78	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67 42.13 41.33 36.26 35.64 31.78	Texture Sandy loam Loam Sandy loam Loam Loam Loam Loam Loam Loam Loam L	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.86 6.83 7.19 6.72 6.63 6.67 6.43	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39 0.38 0.64 0.65 0.66	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009 0.009 0.008 0.016 0.011 0.026	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} \text{Potassium} \ (\text{K}^+) \\ \text{meq} \ (100 \ \text{g})^{-1} \\ \hline 0.105 \\ 0.115 \\ 0.099 \\ 0.132 \\ 0.137 \\ 0.117 \\ 0.145 \\ 0.151 \\ 0.136 \\ 0.153 \\ 0.149 \\ 0.135 \end{array}$	$\begin{array}{c} \text{CEC} \\ \text{meq}(100\text{g})^{-1} \\ 8.51 \\ 9.52 \\ 8.23 \\ 10.92 \\ 11.81 \\ 11.58 \\ 11.66 \\ 11.45 \\ 11.81 \\ 28.19 \\ 19.53 \\ 9.55 \end{array}$
(b) Sample 1 2 3 4 5 6 7 8 9 10 11 12 13	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.1 25.4 24.55 23.25 19.44 19.82	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77 33.27 39.19 41.11 48.78 47.28	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67 42.13 41.33 36.26 35.64 31.78 32.9	Texture Sandy loam Loam Sandy loam Loam Loam Loam Loam Loam Loam Loam L	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.83 7.19 6.72 6.63 6.67 6.43 6.43	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39 0.38 0.64 0.65 0.66 0.69	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009 0.009 0.009 0.008 0.016 0.011 0.026 0.02	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} Potassium (K^+)\\ meq (100g)^{-1}\\ 0.105\\ 0.115\\ 0.099\\ 0.132\\ 0.137\\ 0.117\\ 0.145\\ 0.151\\ 0.136\\ 0.153\\ 0.149\\ 0.135\\ 0.14\end{array}$	CEC meq (100 g) <sup>-1</sup> 8.51 9.52 8.23 10.92 11.81 11.58 11.66 11.45 11.81 28.19 19.53 9.55 9.5
(b) Sample 1 2 3 4 5 6 7 8 9 10 11 12 13 14	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.4 24.55 23.25 19.44 19.82 19.82 19.43	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77 33.27 39.19 41.11 48.78 47.28 47.25	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67 42.13 41.33 36.26 35.64 31.78 32.9 33.12	Texture Sandy loam Loam Sandy loam Loam Loam Loam Loam Loam Loam Loam L	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.86 6.83 7.19 6.72 6.63 6.67 6.43 6.48 6.57	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39 0.38 0.64 0.65 0.66 0.69 0.53	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009 0.009 0.009 0.009 0.008 0.016 0.011 0.026 0.02 0.012	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} \text{Potassium}(\text{K}^+)\\ \text{meq}(100\text{g})^{-1}\\ 0.105\\ 0.115\\ 0.099\\ 0.132\\ 0.137\\ 0.117\\ 0.145\\ 0.151\\ 0.156\\ 0.153\\ 0.149\\ 0.135\\ 0.144\\ 0.134\\ \end{array}$	$\begin{array}{c} \text{CEC} \\ \text{meq}  (100  \text{g})^{-1} \\ 8.51 \\ 9.52 \\ 8.23 \\ 10.92 \\ 11.81 \\ 11.58 \\ 11.66 \\ 11.45 \\ 11.81 \\ 28.19 \\ 19.53 \\ 9.55 \\ 9.5 \\ 9.54 \end{array}$
(b) Sample 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.1 25.4 24.55 23.25 19.44 19.82 19.43 18.13	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77 33.27 39.19 41.11 48.78 47.28 47.28 47.25	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67 42.13 41.33 36.26 35.64 31.78 32.9 33.12	Texture Sandy loam Loam Loam Loam Loam Loam Loam Loam L	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.86 6.83 7.19 6.72 6.63 6.67 6.43 6.43 6.57 6.42	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39 0.38 0.64 0.65 0.66 0.69 0.53 0.73	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009 0.009 0.009 0.008 0.016 0.011 0.026 0.012 0.012	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} \text{Potassium}(\text{K}^+)\\ \text{meq}(100\text{g})^{-1}\\ 0.105\\ 0.115\\ 0.099\\ 0.132\\ 0.137\\ 0.117\\ 0.145\\ 0.151\\ 0.151\\ 0.153\\ 0.149\\ 0.135\\ 0.14\\ 0.134\\ 0.126\\ \end{array}$	CEC meq(100g) <sup>-1</sup> 8.51 9.52 8.23 10.92 11.81 11.58 11.66 11.45 11.81 28.19 19.53 9.55 9.55 9.54 9.32
(b) Sample 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 Mean	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.1 25.4 24.55 23.25 19.44 19.82 19.43 18.13 20.92	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77 33.27 39.19 41.11 48.78 47.28 47.28 47.45 48.75 44.02	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67 42.13 36.26 35.64 31.78 32.9 33.12 33.12 33.12 35.05	Texture Sandy loam Loam Sandy loam Loam Loam Loam Loam Loam Loam Loam L	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.83 7.19 6.72 6.63 6.63 6.63 6.63 6.43 6.43 6.42 6.79	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39 0.38 0.64 0.65 0.66 0.69 0.53 0.73 0.73 0.43	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009 0.009 0.008 0.016 0.011 0.026 0.012 0.012 0.012 0.01	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} \text{Potassium}(\text{K}^+)\\ \text{meq}(100\text{g})^{-1}\\ 0.105\\ 0.115\\ 0.099\\ 0.132\\ 0.137\\ 0.117\\ 0.145\\ 0.151\\ 0.136\\ 0.153\\ 0.149\\ 0.135\\ 0.14\\ 0.136\\ 0.126\\ 0.13\\ \end{array}$	$\begin{array}{c} \text{CEC} \\ \text{meq} \left(100 \text{ g}\right)^{-1} \\ 8.51 \\ 9.52 \\ 8.23 \\ 10.92 \\ 11.81 \\ 11.58 \\ 11.66 \\ 11.45 \\ 11.81 \\ 28.19 \\ 19.53 \\ 9.55 \\ 9.54 \\ 9.55 \\ 9.54 \\ 9.32 \\ 12.07 \end{array}$
(b) Sample	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.1 25.4 24.55 23.25 19.44 19.82 19.43 18.13 20.92 19.86	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77 33.27 39.19 41.11 48.78 47.28 47.45 48.75 44.02	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67 42.13 41.33 36.26 35.64 31.78 32.9 33.12 35.05 35.64	Texture Sandy loam Loam Sandy loam Loam Loam Loam Loam Loam Loam Loam L	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.86 6.83 7.19 6.72 6.63 6.67 6.43 6.67 6.43 6.67 6.42 6.79 6.72	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39 0.38 0.64 0.65 0.66 0.69 0.53 0.73 0.73 0.73 0.73	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009 0.009 0.008 0.016 0.011 0.026 0.012 0.012 0.012 0.011 0.01	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} \text{Potassium} \ (\text{K}^+) \\ \text{meq} \ (100 \ \text{g})^{-1} \\ \hline 0.105 \\ 0.115 \\ 0.099 \\ 0.132 \\ 0.137 \\ 0.117 \\ 0.145 \\ 0.151 \\ 0.136 \\ 0.153 \\ 0.145 \\ 0.135 \\ 0.14 \\ 0.134 \\ 0.126 \\ 0.13 \\ 0.14 \end{array}$	$\begin{array}{c} \text{CEC} \\ \text{meq} (100 \text{ g})^{-1} \\ 8.51 \\ 9.52 \\ 8.23 \\ 10.92 \\ 11.81 \\ 11.58 \\ 11.66 \\ 11.45 \\ 11.81 \\ 28.19 \\ 19.53 \\ 9.55 \\ 9.5 \\ 9.54 \\ 9.32 \\ 12.07 \\ 10.92 \end{array}$
(b) Sample 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 Mean Median SD	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.1 25.4 24.55 23.25 23.25 19.44 19.82 19.43 18.13 20.92 19.86 3.51	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77 33.27 39.19 41.11 48.78 47.28 47.28 47.45 48.75 44.00 7.81	Silt % 27.48 36.6 26.08 33.37 39.41 36.86 39.67 42.13 41.33 36.26 35.64 31.78 32.9 33.12 33.12 35.05 35.64 4.63	Texture Sandy loam Loam Sandy loam Loam Loam Loam Loam Loam Loam Loam L	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.86 6.83 7.19 6.72 6.63 6.67 6.43 6.67 6.43 6.67 6.43 6.48 6.57 6.42 6.79 6.72 0.34	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39 0.38 0.64 0.65 0.66 0.69 0.53 0.73 0.43 0.39 0.21	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009 0.009 0.009 0.009 0.008 0.016 0.011 0.026 0.02 0.012 0.012 0.011 0.01	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} \text{Potassium} \ (\text{K}^+) \\ \text{meq} \ (100 \ \text{g})^{-1} \\ \hline 0.105 \\ 0.115 \\ 0.099 \\ 0.132 \\ 0.137 \\ 0.115 \\ 0.137 \\ 0.145 \\ 0.151 \\ 0.136 \\ 0.153 \\ 0.149 \\ 0.135 \\ 0.14 \\ 0.126 \\ 0.13 \\ 0.14 \\ 0.126 \\ 0.13 \\ 0.14 \\ 0.02 \end{array}$	$\begin{array}{c} \text{CEC} \\ \text{meq} \left(100 \text{ g}\right)^{-1} \\ 8.51 \\ 9.52 \\ 8.23 \\ 10.92 \\ 11.81 \\ 11.58 \\ 11.66 \\ 11.45 \\ 11.81 \\ 28.19 \\ 19.53 \\ 9.55 \\ 9.5 \\ 9.54 \\ 9.32 \\ 12.07 \\ 10.92 \\ 5.20 \end{array}$
(b) Sample 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 Mean Median SD Skewness	Clay % 16.86 17.32 14.17 21.93 22.92 19.86 25.61 25.4 24.55 23.25 19.44 19.82 19.43 18.13 20.92 19.86 3.51 -0.09	Sand % 55.55 46.08 59.75 44.7 37.67 43.28 34.72 32.77 33.27 39.19 41.11 48.78 47.28 47.45 48.75 44.02 44.70 7.81 0.40	Silt % 27.48 36.6 26.08 33.37 39.41 36.26 39.67 42.13 41.33 36.26 35.64 31.78 32.9 33.12 35.05 35.64 4.63 -0.11	Texture Sandy loam Loam Sandy loam Loam Loam Loam Loam Loam Loam Loam L	pH (1:2.5) 7.39 6.98 7.49 6.77 6.46 6.83 7.19 6.72 6.63 6.67 6.43 6.43 6.43 6.43 6.48 6.57 6.42 6.79 6.72 0.34 0.88	Organic matter % 0.1 0.39 0.1 0.45 0.31 0.19 0.28 0.39 0.38 0.64 0.65 0.66 0.69 0.53 0.73 0.43 0.39 0.21 -0.26	Total nitrogen % 0.003 0.007 0.005 0.018 0.014 0.004 0.009 0.009 0.009 0.009 0.009 0.008 0.016 0.011 0.012 0.012 0.012 0.011 0.01 0.01	Assimilable phosphorus ppm 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	$\begin{array}{c} \text{Potassium} \ (\text{K}^+) \\ \text{meq} \ (100 \ \text{g})^{-1} \\ \hline 0.105 \\ 0.115 \\ 0.099 \\ 0.132 \\ 0.137 \\ 0.117 \\ 0.145 \\ 0.151 \\ 0.136 \\ 0.153 \\ 0.149 \\ 0.135 \\ 0.14 \\ 0.126 \\ 0.13 \\ 0.14 \\ 0.126 \\ 0.13 \\ 0.14 \\ 0.02 \\ -0.66 \end{array}$	$\begin{array}{c} \text{CEC} \\ \text{meq}  (100  \text{g})^{-1} \\ \hline 8.51 \\ 9.52 \\ 8.23 \\ 10.92 \\ 11.81 \\ 11.58 \\ 11.66 \\ 11.45 \\ 11.81 \\ 28.19 \\ 19.53 \\ 9.55 \\ 9.55 \\ 9.54 \\ 9.32 \\ 12.07 \\ 10.92 \\ 5.20 \\ 2.42 \end{array}$

than 0.03 m. To obtain vineyard canopy reflectance the active multi-spectral sensors were placed in nadir position and at a distance, from the top of the grapevines rows, of 0.80 m ( $\pm$ 0.20 m, depending on the vineyard height) (Fig. 2). The number of intra-year spectral data sets was fixed to 5 and, according to the season: (i) in 2012, they were started on 29 May and ended on 6 September; and (ii) in 2013, they were started on 30 May and ended on 2 September.

To validate the NDVI with the LAI, several measurements of the latter were taken throughout the ripening stage of the crop in both years. Measurements were recorded by a Plant Canopy Analyser LAI-2000 (LI-COR, Inc, USA), following the procedure of Mabrouk and Carbonneau (1996).

ECa measurements were conducted on 18 February 2011, with a VERIS 3150 Surveyor sensor (Fig. 3), simultaneously in two different soil levels: (i) shallow or ECs – to a depth of 0.30 m from the soil surface; and (ii) deep or ECd – to a depth

of 0.80 m from the surface. Sampling details can be found in Moral et al. (2010).

#### 2.3 Geostatistical and statistical processing

The samplings shown in this work, corresponding to each data set of both growing seasons, were statistically analysed by means of ArcGIS v.10.1 software (ESRI, USA) for geostatistical analyses, and SPSS v.17 software (SPSS Inc., USA), for inferential statistics analyses.

The geostatistical analysis of the multi-temporal NDVI samplings included the followings phases:

- i. Voronoi map a previous exploratory analysis of the samplings was performed to extract outliers.
- ii. ordinary Kriging interpolation the parameters used in the semivariograms of each sampling to generate the corresponding maps are shown in Table 2. Once ob-

Data set	Variable	Model	Lag size (m)	Nugget	Range (m)	Partial sill	RMSE
29 May 2012	NDVI	Spherical	6	0.009	36.5	0.003	0.098
6 July 2012	NDVI	Spherical	6	0.007	32.7	0.005	0.091
24 July 2012	NDVI	Spherical	6	0.005	31.0	0.007	0.083
14 August 2012	NDVI	Spherical	6	0.005	28.7	0.007	0.078
6 September 2012	NDVI	Spherical	6	0.003	33.2	0.005	0.063
30 May 2013	NDVI	Spherical	6	0.008	72.0	0.003	0.092
8 July 2013	NDVI	Spherical	6	0.004	72.0	0.003	0.068
22 July 2013	NDVI	Spherical	6	0.006	72.0	0.002	0.083
12 August 2013	NDVI	Spherical	6	0.005	72.0	0.003	0.074
2 September 2013	NDVI	Spherical	6	0.002	72.0	0.002	0.051
18 February 2011	ECs	Spherical	7	0.321	70.6	0.808	0.601
18 February 2011	ECd	Spherical	7	0.594	67.3	2.264	0.943

**Table 2.** Parameters corresponding to the theoretical semivariograms for the NDVI samplings in 2012 and 2013 growing seasons and for the CEa samplings in 2011 growing season.



Figure 2. ATV with two multi-spectral sensors for NDVI mapping of vineyard canopy.

tained, these maps were rasterized using a pixel size of 2 m.

iii. principal component analysis (PCA) – in this work, a PCA process was established separately for each of the years of study.

At each analysis, input raster data set included the five NDVI samplings of the growing seasons, and the output data were distributed in five principal components. Thus, the results of the PCA analyses obtained consisted of five principal components for each year, where the first principal component shows the NDVI spatial variability for all the mapping dates of each year.

Meanwhile, the ECa samplings were also geostatistically analysed. In this case, only the ordinary Kriging interpolation tool was used, from which the ECs and ECd maps of 2011 were obtained. The parameters used to interpolate the ECa samplings are also shown in Table 2.



Figure 3. Mobile sensor platform Veris 3150 for ECa mapping.

The NDVI samplings from both growing seasons and the ECa samplings at both depths acquired by Kriging were statistically analysed in two phases: (i) firstly, descriptive parameters of each water treatment at each sampling date were acquired to obtain global knowledge of the behaviour of each of the components that make up the statistical design; (ii) secondly, variance analyses of each treatment at each sampling date were made. These analyses allowed comparison of the previously mentioned spatial and temporal behaviours.

To analyse relationships among all the variables studied, values of the first principal component (PC1) of the NDVI variables in both sampling sequences (years 2012 and 2013) and of ECa (ECs and ECd) in the sampling points from the raster maps (Figs. 6 and 7) were extracted and are shown in Table 3.

Finally, in order to determine the importance of the local soil characteristics, given by the ECa and NDVI parameters respectively, in the vegetative expression of the vineyard, the



Figure 4. (a) Accumulation of rainfall and ETc of 2012 and 2013 growing seasons. (b) Temperature components recorded in both 2012 and 2013 growing seasons: Tmean, Tmax and Tmin are monthly average, maximum and minimum temperature respectively; GDD is the growing degree day reached the last day of the month.

**Table 3.** CEa, shallow and deep, and first principal component of the NDVI at soil sample localizations.

Sample	CEa (ms	Sm <sup>-1</sup> )	First pr compone	incipal nt NDVI
	Shallow	Deep	2012	2013
1	5.11	7.63	1.42	1.31
2	3.79	6.38	1.27	1.29
3	3.77	5.96	1.31	1.29
4	5.97	10.00	1.50	1.51
5	5.05	8.61	1.24	1.44
6	5.62	8.72	1.29	1.38
7	6.98	11.29	1.35	1.36
8	6.42	11.12	1.56	1.61
9	4.99	9.55	1.55	1.58
10	5.18	10.05	1.57	1.70
11	5.89	9.58	1.35	1.57
12	4.52	8.17	1.35	1.68
13	4.97	9.36	1.51	1.66
14	4.81	9.15	1.55	1.65
15	5.61	9.85	1.59	1.66
Mean	5.25	9.03	1.43	1.51
Median	5.11	9.36	1.42	1.57
SD	0.88	1.52	0.12	0.15
Skewness	-0.01	-0.61	-0.11	-0.41
Kurtosis	2.76	2.94	1.47	1.57

geographically weighted regression (GWR) tool, included in ArcGIS v.10.1 software (ESRI, USA), was used. The relationship between both variables resulted in maps of determination coefficient ( $R^2$ ) of each water treatment, growing season and depth. The chosen geometric resolution was of 4 m of spatial resolution, which led to goodness of fit in the influence of soil characteristics on the vegetative growth of vines in each of the irrigation treatments of the assay.

### 3 Results and discussions

Climatic variables logged by the weather station situated in a reference field near the tested vineyard recorded diverse behaviour during the 2-year test, with drier conditions in the first growing season. Figure 4a and b show cumulative annual rainfall, cumulative annual ETc, temperature parameters and growing degree days (GDDs) on both years. Focusing on the accumulation of precipitation, the total amount in the second year trial (2013) was more than double compared to the first year trial, where only in its first quarter it had the same amount of rainfall as the whole previous season. However, during the final stages of vegetative development and in the whole ripening phenologic stages, both years had a similarly low accumulation of precipitation. In fact, the temperature was not very different between both years. The observed climatological differences in both seasons influenced differentially the vineyard vegetative development with respect to the different irrigation treatments analysed in this study.

Despite the large difference in precipitations between the two growing seasons, however, for the second year of the test, which was the wettest, hydric demand was similar to the previous year. This result permitted comparison of vegetative response of two consecutive years that were very different in their climatology. Furthermore, if this premise remains constant over the years, it could be possible to know the total needs of the culture of vineyards under whatever climatological conditions, and appropriate reductions could be made in ETc for a watering schedule based on precipitation occurring at each moment of the campaign. Obviously, as Wample and Smithyman (2002) have reported, increases in hydric necessities at each phenological stage must be taken into account, as shown in the slope changes of the accumulation curve of ETc (Fig. 4a), and care must be taken in dry seasons not to bring about unwanted water stress.

In this study, Fig. 5 shows the relationship between LAI estimations and NDVI measurements at the ripening stage of grapes in both years. It can be confirmed that they are well related ( $R^2 = 0.81$ ), thus indicating the possibility to estimate the degree of development of vineyard crops by NDVI determinations obtained by proximal active sensors. These results are in agreement with several authors who have found a good NDVI–LAI relationship (Johnson, 2003).With regard to temporal variability, Fig. 6 shows the results obtained in the first principal component (PC1) of each PCA made on the

		Clay content (%)		CEC		ECs	ECd	First principa	al component
		0–0.3 m	0.3–0.6 m	0–0.3 m	0.3–0.6 m	0–0.3 m	0–0.8 m	NDVI 2012	NDVI 2013
Clay content (%)	0–0.3 m	1							
-	0.3–0.6 m	0.15	1						
CEC	0–0.3 m	0.00	0.27	1					
	0.3–0.6 m	0.00	0.29	0.18	1				
ECs	0–0.3 m	0.59	0.48	0.14	0.05	1			
ECd	0–0.8 m	0.27	0.68	0.38	0.13	0.79	1		
First principal component	NDVI 2012	0.05	0.08	0.27	0.04	0.07	0.31	1	
	NDVI 2013	0.05	0.15	0.52	0.13	0.03	0.30	0.50	1

**Table 4.** Correlation matrix  $(R^2)$  among properties and vegetative vigour expression of the vineyard assay area.



Figure 5. NDVI–LAI relationship of both 2012 and 2013 years.

different mapping dates in each growing season. According to these results, there were differences in plant development even when the same doses of irrigation and cultural practices were administered to the plots with different types of irrigation treatment.

Soil properties, spatial variation of ECa, both ECs and ECd, and PC1 of NDVI (both 2012 and 2013 growing seasons) were statistically analysed (Table 4). On the one hand, ECs and ECd results were correlated with the clay content from 0 to 0.30 m ( $R^2 = 0.59$  and  $R^2 = 0.27$ , respectively) and from 0.30 to 0.60 m ( $R^2 = 0.48$  and  $R^2 = 0.68$ , respectively). These results indicate that ECa could be used as a tool to estimate some soil properties – thus avoiding the time-consuming inconvenience and cost of soil sampling and analysis in the laboratory. On the other hand, it is of interest to note the correlation between CEC, from 0 to 0.30 m, and PC1 of NDVI of 2013 ( $R^2 = 0.53$ ), indicating the higher influence of soil fertility due to the existence of water resources in that year.

Results of ECa spatial variation (Fig. 7) seem to be a pattern consisting of a variation in ECa from the northern and southern boundaries of the assay up to the centre, and also from east to west, coinciding with some physicochemical parameters of soil. There exists, too, a pattern in the variability of soil characteristics due to the good relationship with ECa, in particular with clay content (Moral et al., 2010). Spatial variability of ECa, both shallow and deep, had also shown significant differences among the locations of the plots of the

Figure 6. NDVI first principal component of (a) 2012 and (b) 2013.

different irrigation treatments (Table 5), with different values for the soil properties that influenced vegetative growth of the grapevines. In general, the spatial variability pattern mentioned above was observed in the plots with the different treatments, with higher ECs or ECd values in those plots near the northern and southern boundaries of the vineyard test site. Because of this spatial variability, it was necessary to perform, even within plots of the same treatment, geostatistical analyses between NDVI and ECa to determine the extent of the influence of soil properties on vegetative growth of the vineyard in each of the irrigation treatments and their respective plots.

#### 3.1 Intra-year variability

#### 3.1.1 2012 growing season

Both temporal and spatial evolution of the NDVI index of the irrigation treatments and their respective plots in the 2012 growing season is shown in Fig. 8. At first glance, the results of NDVI mapping for this year show how all the treatments had a temporal evolution similar to a Gaussian function, the mean value of the index increasing as the campaign advanced, reaching a maximum value around the phenolog-



Table 5. Statistic descriptive analyses of shallow and deep soil ECa interpolated d	lata
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Data set	Treatment	Plot <sup>1</sup>	$\frac{Mean^2}{(mS m^{-1})}$	SD (mS m <sup>-1</sup> )	Minimum (mS <sup>-1</sup> )	Maximum (mS m <sup>-1</sup> )	Range (mS m <sup>-1</sup> )	Skewness
ECs	Fully watered	1	5.57cd	0.29	4.95	6.30	1.35	0.32
	Fully watered	2	5.49d	0.33	4.84	6.21	1.37	0.01
	Fully watered	3	6.63a	0.55	5.54	7.69	2.15	-0.02
	Fully watered	4	5.94b	0.48	5.05	6.83	1.78	0.16
	RDI 50-20 – RDI 30a	1	4.55h	0.17	4.23	5.06	0.83	0.40
	RDI 50-20 - RDI 30a	2	5.52d	0.42	4.60	6.75	2.15	-0.08
	RDI 50-20 - RDI 30a	3	6.59a	0.30	5.82	7.39	1.57	0.14
	RDI 50-20 – RDI 30a	4	5.61de	0.43	4.81	6.51	1.70	0.24
	RDI 50-0 – RDI 30b	1	5.29ef	0.49	4.50	6.26	1.76	0.27
	RDI 50-0 - RDI 30b	2	5.25f	0.23	4.72	5.63	0.91	-0.52
	RDI 50-0 - RDI 30b	3	5.14f	0.49	4.31	6.40	2.09	0.51
	RDI 50-0 – RDI 30b	4	5.72c	0.74	4.33	7.10	2.77	-0.16
	Non-watered	1	4.80g	0.30	4.39	5.71	1.32	0.66
	Non-watered	2	5.4de	0.45	4.27	6.45	2.18	-0.29
	Non-watered	3	5.60cd	0.51	4.61	6.50	1.89	0.19
	Non-watered	4	5.49d	0.30	5.03	6.60	1.57	0.76
ECd	Fully watered	1	9.90cd	0.77	8.79	13.81	5.02	2.69
	Fully watered	2	10.01c	0.39	9.06	10.83	1.77	0.21
	Fully watered	3	10.96b	0.77	8.95	12.49	3.54	-0.03
	Fully watered	4	9.96c	0.64	8.76	12.06	3.30	0.51
	RDI 50-20 – RDI 30a	1	8.62gh	0.33	8.07	9.62	1.55	0.76
	RDI 50-20 – RDI 30a	2	9.97c	0.49	9.00	11.23	2.23	0.15
	RDI 50-20 – RDI 30a	3	11.37a	0.36	9.62	12.00	2.38	-1.38
	RDI 50-20 – RDI 30a	4	8.82fg	1.05	7.11	10.82	3.71	0.23
	RDI 50-0 - RDI 30b	1	8.91f	0.48	7.75	10.15	2.40	0.35
	RDI 50-0 - RDI 30b	2	9.68d	0.37	9.01	10.33	1.32	-0.21
	RDI 50-0 - RDI 30b	3	9.76cd	0.48	8.73	10.55	1.82	-0.43
	RDI 50-0 – RDI 30b	4	8.88fg	1.51	6.08	11.50	5.42	-0.04
	Non-watered	1	8.53h	0.37	7.82	9.47	1.65	0.27
	Non-watered	2	9.40e	0.77	7.41	11.14	3.73	-0.20
	Non-watered	3	9.77cd	0.28	9.11	10.49	1.38	0.35
	Non-watered	4	8.72fgh	0.57	7.90	10.53	2.63	0.99

Sampling was carried out on 18 February 2011. <sup>1</sup> Plots are numbered in a north–south orientation. <sup>2</sup> Variance analyses among treatments are made for each data set independently; a, b, c, and d mean significant difference at p value  $\leq 0.05$  in Tukey post hoc analysis.

ical stage of *véraison*, and then decreasing until harvest. In spite of this sigmoidal evolution, a positive relationship between NDVI and water dose occurred, in which the fully watered treatment maintained the higher mean value of NDVI, and the non-watered treatment the lower mean value, for all the mapping dates, these differences being, moreover, significant (Table 6). These results indicate that the greater the quantity of water in the vineyard the higher the vegetative development of its canopy.

The intermediate RDI 50-20 and RDI 50-0 irrigation treatments also showed significant differences between NDVI values with regard to the previous ones, with intermediate values. Both RDI treatments kept their NDVI values similar up to January, and then they became different as a result of the change in the water dose of the experimental design. At that moment, the RDI 50-0 treatment had a greater decrease in NDVI mean value and, consequently, in vegetative expression of the vineyard. Taking into account these aspects, and bearing in mind the existing relationship between vegetative growth of the vines and NDVI value, it can be considered that the latter increased its value when water doses were higher, and that variations in doses will result in changes in the vegetative expression of the vineyard.

On the other hand, despite the relation found between the water doses applied in the assay and the vegetative development of the vines, there were significant differences among

Data set	Treatment	Mean <sup>1</sup>	SD	Minimum	Maximum	Range <sup>2</sup>	Skewness
29 May 2012	Fully watered	0.643a	0.036	0.502	0.713	0.211	-0.319
	RDI 50-20	0.608b	0.039	0.507	0.691	0.184	-0.548
	RDI 50-0	0.597c	0.046	0.472	0.706	0.234	-0.265
	Non-watered	0.572d	0.044	0.446	0.677	0.231	-0.302
	Mean	0.605	0.041	0.482	0.697	0.215	
6 July 2012	Fully watered	0.729a	0.050	0.586	0.807	0.221	-0.535
	RDI 50-20	0.708b	0.042	0.579	0.780	0.201	-0.591
	RDI 50-0	0.714b	0.054	0.569	0.817	0.248	-0.311
	Non-watered	0.624c	0.060	0.453	0.766	0.313	-0.210
	Mean	0.694	0.052	0.547	0.793	0.246	
24 July 2012	Fully watered	0.750a	0.041	0.597	0.813	0.216	-0.998
	RDI 50-20	0.718b	0.046	0.452	0.789	0.337	-1.300
	RDI 50-0	0.721b	0.055	0.554	0.803	0.249	-0.767
	Non-watered	0.618c	0.064	0.430	0.730	0.300	-0.448
	Mean	0.702	0.052	0.508	0.784	0.276	
14 August 2012	Fully watered	0.742a	0.039	0.483	0.803	0.320	-1.853
	RDI 50-20	0.712b	0.048	0.577	0.794	0.217	-0.475
	RDI 50-0	0.696c	0.070	0.512	0.800	0.288	-0.828
	Non-watered	0.613d	0.054	0.404	0.731	0.327	-0.568
	Mean	0.691	0.053	0.494	0.782	0.288	
6 September 2012	Fully watered	0.701a	0.032	0.575	0.761	0.186	-0.825
	RDI 50-20	0.673b	0.045	0.534	0.740	0.206	-0.681
	RDI 50-0	0.647c	0.070	0.445	0.750	0.305	-0.917
	Non-watered	0.600d	0.056	0.417	0.707	0.290	-0.647
	Mean	0.655	0.051	0.493	0.740	0.247	
30 May 2013	Fully watered	0.671b	0.039	0.570	0.749	0.179	-0.454
	RDI 30a (previous 50-20)	0.680a	0.045	0.570	0.749	0.179	-0.728
	RDI 30b (previous 50-0)	0.665b	0.053	0.518	0.747	0.229	-0.573
	Non-watered	0.671b	0.050	0.528	0.761	0.233	-0.547
	Mean	0.672	0.047	0.547	0.752	0.205	
8 July 2013	Fully watered	0.779a	0.040	0.655	0.831	0.176	-0.827
	RDI 30a (previous 50-20)	0.766b	0.052	0.597	0.833	0.236	-1.000
	RDI 30b (previous 50-0)	0.754bc	0.069	0.555	0.832	0.277	-1.138
	Non-watered	0.761c	0.050	0.614	0.823	0.209	-0.808
	Mean	0.769	0.053	0.605	0.830	0.225	
22 July 2013	Fully watered	0.737a	0.034	0.646	0.794	0.148	-0.429
	RDI 30a (previous 50-20)	0.738a	0.049	0.607	0.792	0.185	-1.200
	RDI 30b (previous 50-0)	0.724b	0.063	0.547	0.802	0.255	-1.238
	Non-watered	0.728b	0.043	0.617	0.792	0.175	-0.659
	Mean	0.732	0.047	0.604	0.795	0.191	
12 August 2013	Fully watered	0.749a	0.042	0.632	0.822	0.190	-0.366
	RDI 30a (previous 50-20)	0.734b	0.053	0.570	0.797	0.227	-0.986
	RDI 30b (previous 50-0)	0.721c	0.071	0.542	0.810	0.268	-0.989
	Non-watered	0.718c	0.050	0.583	0.796	0.213	-0.735
	Mean	0.731	0.054	0.582	0.806	0.225	
2 September 2013	Fully watered	0.753a	0.030	0.656	0.795	0.139	-0.766
	RDI 30a (previous 50-20)	0.742b	0.035	0.624	0.790	0.166	-1.076
	RDI 30b (previous 50-0)	0.731c	0.054	0.564	0.791	0.227	-1.133
	Non-watered	0.725d	0.037	0.609	0.781	0.172	-0.543
	Mean	0.738	0.039	0.613	0.789	0.176	

Table 6. Statistic descriptive analyses of the NDVI interpolated data sets for 2012 and 2013 growing seasons (dimensionless).

 $\overline{1}$  Variance analyses among treatments are made for each data set independently; a, b, c, and d mean significant difference at *p* value  $\leq 0.05$  in Tukey post hoc analysis. <sup>2</sup> Statistical range of NDVI values (max-min).



**Figure 7.** Interpolated apparent electrical conductivity maps of 2011 growing season: (a) shallow ECa map and (b) deep ECa map.

the various plots of each water treatment (data not shown), indicating that spatial variability of the NDVI index existed, and consequently of vegetative growth too, which was dependent on other factors, even though the characteristics of the management were identical. In this respect, it can be observed in Fig. 8 how vegetative expression was not homogeneous in all the plots within a specific water treatment, but rather variations were found in the NDVI value depending on the geographical location of each of the plots. Thus, on a specific mapping date, some plots with different water treatments had similar mean values of NDVI, even between plots with fully watered and non-watered treatments. A factor associated with geographical location, therefore, has had some influence on the vegetative growth. The terroir effect, in which the physicochemical parameters of soil are included, could be one of the factors that have caused a certain influence on vegetative development, as indicated by van Leeuwen and Seguin (2006).

A priori, the global results on the relationship between NDVI and ECa indicate a low association when compared in the first 0.30 m of soil depth (ECs, Table 7), and a relatively high one when a large section of soil is considered (ECd, Table 7). These results suggest that the soil surface layer has little influence on vegetative expression of the vine-yard because of the deeper distributions of the roots, although it does influence other crops with shallow roots (Fortes et al., 2014). Furthermore, in the year when climatic quality involved drought (2012), ECa and NDVI values were lower, suggesting that the soil properties seem to be an influential factor but not a limiting one in vegetative expression, the availability of water resources being the principal limiting factor.



**Figure 8.** Interpolated NDVI maps of 2012 growing season: (a) 29 May; (b) 6 July; (c) 24 July; (d) 14 August; and (e) 6 September.



**Figure 9.** NDVI maps year 2013: (a) 30 May; (b) 8 July; (c) 22 July; (d) 12 August; and (e) 2 September.

# 3.1.2 2013 growing season

Figure 9 shows the spatial and temporal evolution of NDVI in the watered treatments and their respective plots in the 2013 growing season. In the same way as the previous year, increased water doses applied to the vineyard were associated with a higher NDVI mean value. However, in this season, the differences in this mean value were closer, being no higher than 0.10 points of index value. The intense precipitations between post-harvest of 2012 and flowering of 2013 decreased the possibility of water stress in the vines, so vegetative development was very similar at the beginning of the NDVI mappings, the only difference being the RDI 30 treatment that came from the RDI 50-20 of the previous growing season (Table 6). On the other hand, in the 2013 season, temporal



**Figure 10.** Local  $R^2$  of GWR analyses: (a) first principal component of NDVI in 2012 and ECs of 2011; (b) first principal component of NDVI in 2013 and ECs of 2011; (c) first principal component of NDVI in 2012 and ECd of 2011; (d) first principal component of NDVI in 2013 and ECd 2011.

evolution of the mean NDVI value of the whole treatments was more homogeneous for most of the season. Generally speaking, there was an initial increment of the NDVI value in all treatments up to the phenological stage of *véraison*, from which point that value was constant until the harvest. Both results, i.e. higher and constant NDVI values as compared to the previous season, may have been caused by the high groundwater recharge, which provided almost unlimited water to plants during the early stages of vineyard growth.

With regard to temporal behaviour of NDVI between water treatments, the mean value of the index resulted in slightly higher significant differences as the season progressed, with two different groups of treatments at *véraison* (Table 6): (i) fully watered and RDI 30a and (ii) non-watered and RD 30b. From this moment until harvest, the irrigation treatments of the first group showed significant differences in the mean NDVI value, while treatments of the second group had a similar value. In general, in the same way as the previous season, there were some factors, in this case climatological ones, that modified the expected trend of a vineyard managed under specific water conditions.

The irrigation treatments of the 2013 growing season also had significant differences in mean NDVI values among their respective plots spatially (data not shown), with a pattern of reduced values from north to south of the vineyard test area. Thus, for the same water treatment and mapping date, the mean NDVI values of each plot decreased the further south the plot was located, with, furthermore, significant differences among them. This result was already shown by Blanco

**Table 7.** Correlation matrix  $(R^2)$  between first principal components of 2012 and 2013 growing seasons and apparent electrical conductivities, shallow and deep, interpolated data of 2011.

Variable	First PC NDVI 2012	First PC NDVI 2013	ECs 2011	ECd 2011
First PC NDVI 2012	1.00			
First PC NDVI 2013	0.58	1.00		
ECs 2011	0.18	0.16	1.00	
ECd 2011	0.59	0.70	0.83	1.00

et al. (2012), who reported that vegetative growth of vines under the same management had different behaviours due to spatial changes in some influential factor, such us spatial variability of the physicochemical properties of soil. On the other hand, the influence of terroir, taking into account its climatic and edaphic factors, was so high in the 2013 season that it gave rise to similar mean NDVI values, with some exceptions, in plots with different irrigation treatments. Thus, for example, northern plots of fully watered and non-watered treatments gave similar NDVI values, as did the southern plots, but these values were statistically different between the two geographical locations. This behaviour can be seen in Fig. 9.

Figure 10 shows the local relationship between the PC1 of NDVI in each growing season and the ECa in 2011, both shallow and deep, throughout the test area, which is the level of influence of soil features on vegetative development in each water treatment. The highest ratios prevailed, again, in the northern and southern limits of the test area, in agreement with those zones where ECa reached the lowest values. Thus, the maximum values in the relationship between soil properties and vegetative growth were obtained during the 2013 season; values of  $R^2$  in the relationship between soil properties and vegetative growth were obtained during the 2013 season, with values of  $R^2$  of 0.55 and 0.64 points of ECs and ECd respectively, compared to 0.56 and 0.47 points reached in 2012. Thus, a lower maximum value of ECa was produced in the 2012 growing season. However, that year presented a good relationship between ECa and NDVI in a larger area than the 2013 growing season. These results could be due to the fact that soils with water limits in zones where ECa has low values (and lower expected clay content) (Sudduth et al., 2005; Terrón et al., 2011) tend to have higher water availability for the plant as compared to soils with higher ECa values (higher percentage of clay content).

#### 3.2 Between-year variability

The results of each mapping date of NDVI in both growing seasons (Figs. 8 and 9) show the behaviour of the vegetative development of the whole treatments established in the experimental designs. As has already been stated, NDVI values and, accordingly, vegetative growth of the vineyard were

# 470

Table 8. Correlation matri	ces between 2012 and 2013	NDVI surfaces of each irrigation treatmen
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			2012			
Treatment	Data set	29 May	6 July	24 July	4 August	6 September
Fully watered	29 May	1				
	6 July	0.47	1			
	24 July	0.33	0.65	1		
	14 August	0.42	0.35	0.47	1	
	6 September	0.28	0.57	0.59	0.57	1
RDI 50-20	29 May	1				
	6 July	0.74	1			
	24 July	0.61	0.72	1		
	14 August	0.69	0.79	0.70	1	
	6 September	0.70	0.81	0.66	0.84	1
RDI 50-0	29 May	1				
	6 July	0.59	1			
	24 July	0.69	0.86	1		
	14 August	0.69	0.89	0.86	1	
	6 September	0.68	0.86	0.83	0.95	1
Non-watered	29 May	1				
	6 July	0.70	1			
	24 July	0.68	0.83	1		
	14 August	0.66	0.83	0.81	1	
	6 September	0.65	0.82	0.79	0.90	1
			2013			
		30 May	8 July	22 July	12 August	2 September
Fully watered	30 May	1				
	8 July	0.76	1			
	22 July	0.61	0.61	1		
	12 August	0.58	0.66	0.67	1	
	2 September	0.64	0.79	0.63	0.76	1
RDI 30a	30 May	1				
	8 July	0.85	1			
	22 July	0.82	0.86	1		
	12 August	0.83	0.85	0.93	1	
	2 September	0.83	0.87	0.91	0.90	1
RDI 30b	30 May	1				
	8 July	0.90	1			
	22 July	0.87	0.93	1		
	12 August	0.88	0.94	0.95	1	
	2 September	0.89	0.95	0.95	0.96	1
Non-watered	30 May	1				
	8 July	0.80	1			
	22 July	0.77	0.88	1		
	12 August	0.84	0.89	0.88	1	
	2 September	0.73	0.83	0.85	0.86	1

influenced by soil properties (including groundwater level) in spatial components, and by climatic features in temporal ones.

With regard to the temporal variability, Fig. 6 shows the results obtained in the first principal component (PC1) of each PCA made for the different mapping dates in each growing season. This PC1 shows spatial variability of NDVI for

the whole of NDVI mapping dates for each year. Thus, each PC1 map for 2012 explains 80.57% of the temporal variability of each geographical location within the assay area, and 85.92% for the 2013 growing season. Thus, the PC1 for each year shows more than 80% of the mean variability of the NDVI values throughout both seasons in each of the irrigation treatments and their respective plots. In general, the PC1 map for 2013 shows higher and more homogeneous values than the 2012 one, indicating a higher and more homogeneous vegetative growth of grapevines.

Table 8 shows the level of relationship of NDVI values between the different mapping dates for each irrigation treatment. Generally speaking, in both 2012 and 2013 there was an increase in the determination coefficient  $(R^2)$  given by the NDVI values as the season progressed, indicating that the continuous development of the vineyard canopy was slowing; that is, the development rate or evolution of that canopy was increasingly smaller until the harvest stage was reached. However, the behaviour of the different irrigation treatments did not evolve equally either within years or between years. In 2012, the treatment with higher water doses (fully watered) had low values of correlation ( $R^2$  lower than 0.65) in all NDVI mapping dates due to a higher development rate versus the rest of water and rainfed treatments during the later phenological stages of the vineyard, thus indicating higher change rates. Conversely, Non-watered treatment had determination coefficients above 0.65 points, which suggests a low development rate due to lower hydric availability as a limiting factor. Meanwhile, the 2013 season showed a similar behaviour pattern in the extreme water treatments. Obviously, the determination coefficients were higher and more homogeneous than the previous season between the different mapping dates due to the intense precipitations, these being  $R^2 < 0.77$  for fully watered and  $R^2 > 0.73$  for non-watered treatments. These results point to a lower canopy development than in 2012 and, in the 2013 season, the differences between treatments were less pronounced.

With respect to differences in spatial variability of vegetative growth between the years tested, the 2013 season showed greater homogeneity, where the highest increase was found in the northern half of the test area, regardless of the water dose applied. Conversely, this vegetative development was lower in the south, where the southern plot of non-watered treatment did not have lower vegetative growth, although it did respond to a spatial pattern. Thus, the vegetation response in 2012 was more dependent on the irrigation treatments, while in 2013 it was more dependent on soil characteristics or other edaphic-climatic variables. In 2013, RDI 50-20 and RDI 50-0 treatments became RDI 30a and RDI 30b respectively, with water doses of 30 % of ETc during the whole irrigation period. In the same way that the rest of the treatments had higher NDVI values in 2013, RDI 30 also showed higher NDVI values than the RDI treatments of the previous season. However, despite having the same water dose, RDI 30b gave lower values than RDI 30a during most of the season (data not shown), thus suggesting once again that water dose must be redefined taking into account climate and soil properties.

According to Howell (2001), there must be an optimal method of crop management in any situation, in order to obtain the desired yields and qualities, but intra-year and between-year management must be performed in accordance with the terroir features of each year.

# 4 Conclusions

Water level and vegetative growth were clearly related; greater availability of water resources gave rise to greater vegetative development of the vineyard. However, spatialtemporal changes in climatic quality or in soil properties also affect vegetative expression. To the already estimated differences in vegetative growth of grapevines between different water doses, one must add the effects that climate and soil properties have on plants. Consequently, the application of the same cultural practices in each growing season makes it unfeasible to attain stable goals, i.e. the same level of quality in grapes and wines or similar yields every season. The application of some precision agriculture techniques to the vineyard crop, through real-time measurements of NDVI and ECa, makes it possible to determine homogeneous zones of growth and development in the vineyard dependent on climatic and soil characteristics for a specific irrigation treatment. Thus, according to the results of this study, the following can be concluded:

- i. In global terms, the higher the water doses the higher the NDVI values and, hence, the greater the vegetative growth of the vineyard.
- ii. Vegetative development is not homogeneous, even when the same cultural practices are being used, but spatial and temporal variability occur depending on climatic and soil characteristics and their interactions.
- iii. It is necessary for crop management to adapt to the variability of agronomic factors in order to achieve homogeneous vegetative growth even in zones where the soil characteristics are different.

An irrigation schedule based on real-time NDVI results, and knowledge of the variability of soil characteristics could be the basis for improved vineyard management.

Acknowledgements. This work was carried out with funding from the RITECA Project, Transboundary Research Network Extremadura, Center and Alentejo, co-financed by the European Regional Development Fund (ERDF), by the Spain–Portugal Border Cooperation Operational Programme (POCTEP) 2007–2013 and by the Government of Extremadura.

This research was also co-financed by the Government of Extremadura and the European Regional Development Fund (ERDF) through the project GR10038 (Research Group TIC008). The vineyard irrigation project which has complemented this work is INIA RTA2009-00026-C02-02 and was co-financed by the European Regional Development Fund (ERDF).

Edited by: E. Costantini

# References

- Baret, F. and Guyot, G.: Potentials and limits of vegetation indices for LAI and APAR assessment, Remote Sens. Environ., 35, 161– 173, 1991.
- Blanco, J., Terrón, J. M., Pérez, F. J., Galea, F., Salgado, J. A., Moral, F. J., Marques da Silva, J. R., and Silva, L. L.: Variabilidad espacial y temporal del vigor vegetativo en viñedo sin restricciones hídricas en la demanda evapotranspirativa, VII Congreso Ibérico de Agroingeniería y Ciencias Hortícolas, Madrid, 2012.
- Bravdo, B. and Hepner, Y.: Irrigation management and fertigation to optimize grape composition and vine performance, Symposium on Grapevine Canopy and Vigor Management, XXII IHC 206, 49–68, 1986
- Carbonneau, A.: General relationship within the whole-plant: examples of the influence of vigour status, crop load and canopy exposure on the sink "berry maturation" for the grapevine, Strategies to Optimize Wine Grape Quality 427, 99–118, 1995.
- Cortell, J. M., Halbleib, M., Gallagher, A. V., Righetti, T. L., and Kennedy, J. A.: Influence of Vine Vigor on Grape (*Vitis vinifera* L. Cv. Pinot Noir) and Wine Proanthocyanidins, J. Agr. Food Chem., 53, 5798–5808, 2005.
- Corwin, D. and Lesch, S.: Application of soil electrical conductivity to precision agriculture, Agron. J., 95, 455–471, 2003.
- Deloire, A., Vaudour, E., Carey, V., Bonnardot, V., and van Leeuwen, C.: Grapevine responses to terroir: a global approach, J. Int. Sci. Vigne Vin, 39, 149–162, 2005.
- Dobrowski, S. Z., Ustin, S. L., and Wolpert, J. A.: Remote estimation of vine canopy density in vertically shoot-positioned vineyards: determining optimal vegetation indices, Aust. J. Grape Wine R., 8, 117–125, 2002.
- Dry, P. R.: Canopy management for fruitfulness, Aust. J. Grape Wine R., 6, 109–115, 2000.
- Fortes, R., Prieto, M., Terron, J., Blanco, J., Millan, S., and Campillo, C.: Using apparent electric conductivity and NDVI measurements for yield estimation of processing tomato crop, T. ASABE, 57, 827–835, 2014.
- Gitelson, A. A. and Merzlyak, M. N.: Non-Destructive Assessment of Chlorophyll Carotenoid and Anthocyanin Content in Higher Plant Leaves: Principles and Algorithms, in: Remote Sensing for Agriculture and the Environment, edited by: Stamatiadis, S., Lynch, J. M., and Schepers, J. S., Ella, Greece, 78–94, 2004.
- Hall, A., Lamb, D. W., Holzapfel, B., and Louis, J.: Optical remote sensing applications in viticulture – a review, Aust. J. Grape Wine R., 8, 36–47, 2002.
- Hall, A., Louis, J., and Lamb, D. W.: Low-resolution remotely sensed images of winegrape vineyards map spatial variability in planimetric canopy area instead of leaf area index, Aust. J. Grape Wine R., 14, 9–17, 2008.
- Howell, G. S.: Sustainable grape productivity and the growth-yield relationship: A review, Am. J. Enol. Viticult., 52, 165–174, 2001.
- Intrigliolo, D. S. and Castel, J. R.: Response of grapevine cv. "Tempranillo" to timing and amount of irrigation: water relations, vine

SOIL, 1, 459–473, 2015

growth, yield and berry and wine composition, Irrigation Sci., 28, 113–125, 2010.

- Jackson, R. D., Slater, P. N., and Pinter Jr., P. J.: Discrimination of growth and water stress in wheat by various vegetation indices through clear and turbid atmospheres, Remote Sens. Environ., 13, 187–208, 1983.
- Johnson, L. F.: Temporal stability of an NDVI-LAI relationship in a Napa Valley vineyard, Aust. J. Grape Wine R., 9, 96–101, 2003.
- Jordan, C. F.: Derivation of Leaf-Area Index from Quality of Light on Forest Floor, Ecology, 50, 663–666, 1969.
- Lamb, D. and Bramley, R. G. V.: Managing and monitoring spatial variability in vineyard variability, Nat. Res. Man., 4, 25–30, 2001.
- Mabrouk, H. and Carbonneau, A.: A simple method for determination of grapevine Vitis vinifera L. leaf area, Progres Agricole et Viticole, 113, 392–398, 1996.
- Moral, F. J., Terrón, J. M., and Marques da Silva, J. R.: Delineation of management zones using mobile measurements of soil apparent electrical conductivity and multivariate geostatistical techniques, Soil and Tillage Research, 106, 335–343, 2010.
- Mullins, M. G., Bouquet, A., and Williams, L. E.: Biology of the Grapevine, Cambridge University Press, 1992.
- Peñuelas, J., Filella, I., Biel, C., Serrano, L., and Save, R.: The reflectance at the 950–970 nm region as an indicator of plant water status, Int. J. Remote Sens., 14, 1887–1905, 1993.
- Rondeaux, G., Steven, M., and Baret, F.: Optimization of soiladjusted vegetation indices, Remote Sens. Environ., 55, 95–107, 1996.
- Rouse, J. W., Haas, R. H., Schell, J. A., and Deering, D. W.: Monitoring the vernal advancement and retrogradation (green wave effect) of natural vegetation, Remote Sensing Center, Texas A&M Univ., College Station, 93 pp., 1973.
- Smart, R. E. and Coombe, B. G.: Water relations of grapevines, in: Water deficits and plant growth, edited by: Kozlowski, T. T., Academic Press, 137–196, 1983.
- Smart, R. E.: Principles of Grapevine Canopy Microclimate Manipulation with Implications for Yield and Quality. A Review, Am. J. Enol. Viticult., 36, 230–239, 1985.
- Soil Survey Staff: Keys to soil taxonomy, USDA-Natural Resources Conservation Service, Washington, D.C., 333 pp., 2006.
- Sudduth, K., Kitchen, N., Wiebold, W., Batchelor, W., Bollero, G., Bullock, D., Clay, D., Palm, H., Pierce, F., and Schuler, R.: Relating apparent electrical conductivity to soil properties across the north-central USA, Comput. Electron. Agr., 46, 263–283, 2005.
- Tardáguila, J. and Diago, M.: Viticultura de precisión: principios y tecnologías aplicadas en el viñedo, VI World Wine Forum, Logroño, Spain, 2008.
- Terrón, J., da Silva, J. M., Moral, F., and García-Ferrer, A.: Soil apparent electrical conductivity and geographically weighted regression for mapping soil, Precis. Agric., 12, 750–761, 2011.
- Terrón, J., Moral, F., da Silva, J. M., and Rebollo, F.: Analysis of spatial pattern and temporal stability of soil apparent electrical conductivity and relationship with yield in a soil of high clay content, 3rd Global Workshop on Proximal Soil Sensing, 2013.
- van Leeuwen, C. and Seguin, G.: The concept of terroir in viticulture, Journal of Wine Research, 17, 1–10, 2006.
- Vaudour, E.: The Quality of Grapes and Wine in Relation to Geography: Notions of Terroir at Various Scales, Journal of Wine Research, 13, 117–141, 2002.

- Wample, R. and Smithyman, R.: Regulated deficit irrigation as a water management strategy in *Vitis vinifera* production, in: Deficit irrigation practices, Food and Agricultural Organization of the United Nations (FAO), Rome, Italy, 89–100, 2002.
- Williams, L. E. and Araujo, F. J.: Correlations among Predawn Leaf, Midday Leaf, and Midday Stem Water Potential and their Correlations with other Measures of Soil and Plant Water Status in Vitis vinifera, J. Am. Soc. Hortic. Sci., 127, 448–454, 2002.
- Yrissarry, J. J. B. and Naveso, F. S.: Use of weighing lysimeter and Bowen-Ratio Energy-Balance for reference and actual crop evapotranspiration measurements, III International Symposium on Irrigation of Horticultural Crops, 537, 143–150, 1999.